

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization International Bureau



(43) International Publication Date
31 December 2003 (31.12.2003)

PCT

(10) International Publication Number
WO 2004/001010 A2

- (51) International Patent Classification⁷: C12N
- (21) International Application Number: PCT/US2003/019760
- (22) International Filing Date: 23 June 2003 (23.06.2003)
- (25) Filing Language: English
- (26) Publication Language: English
- (30) Priority Data:
- | | | |
|------------|---------------------------|----|
| 60/390,747 | 21 June 2002 (21.06.2002) | US |
| 60/398,180 | 24 July 2002 (24.07.2002) | US |
| 60/398,287 | 24 July 2002 (24.07.2002) | US |
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- (81) Designated States (national): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, OM, PH, PL, PT, RO, RU, SC, SD, SE, SG, SK, SL, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, YU, ZA, ZM, ZW.
- (84) Designated States (regional): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SI, SZ, TZ, UG, ZM, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HU, IE, IT, LU, MC, NL, PT, RO, SE, SI, SK, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

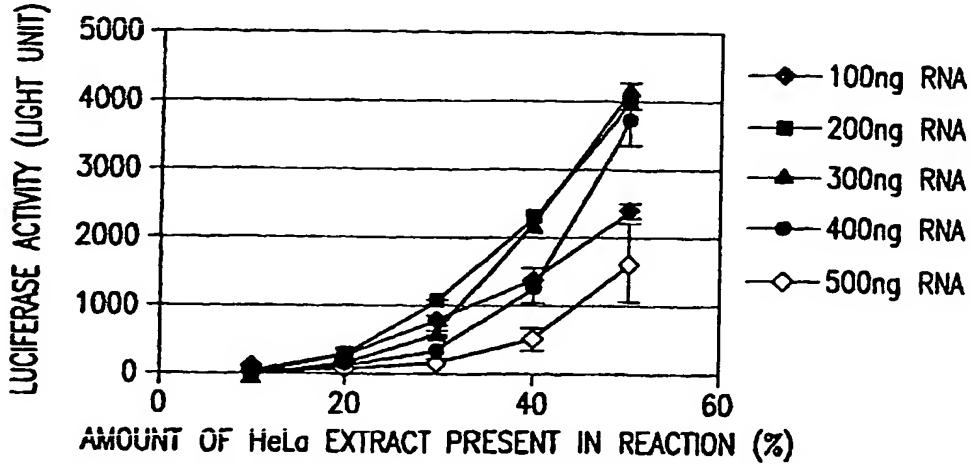
Published:

— without international search report and to be republished upon receipt of that report

[Continued on next page]

(54) Title: METHODS FOR IDENTIFYING SMALL MOLECULES THAT MODULATE PREMATURE TRANSLATION TERMINATION AND NONSENSE MEDIATED mRNA DECAY

WILD-TYPE RNA IN HeLa TRANSLATION EXTRACT



WO 2004/001010 A2

(57) Abstract: The present invention relates to methods for identifying compounds that modulate premature translation termination and/or nonsense-mediated mRNA decay by screening and identifying compounds that modulate the post-transcriptional expression of any gene with a premature translation stop codon. The invention particularly relates to using any gene encoding a premature stop codon to identify compounds that modulate premature translation termination and/or nonsense-mediated mRNA decay. A compound that modulates premature translation termination and/or nonsense-mediated mRNA decay of a target gene is identified using standard methods known in the art to measure changes in translation or mRNA stability of the gene product or mRNA of the gene with the premature stop codon. The methods of the present invention provide a simple, sensitive assay for high-throughput screening of libraries of compounds to identify pharmaceutical leads.

METHODS FOR IDENTIFYING SMALL MOLECULES THAT MODULATE
PREMATURE TRANSLATION TERMINATION AND NONSENSE MEDIATED mRNA
DECAY

This application is entitled to and claims priority benefit to U.S. Provisional Patent Application No. 60/390,747, filed June 21, 2002, U.S. Provisional Patent Application 5 No. 60/398,180, filed July 24, 2002 and U.S. Provisional Patent Application No. 60/398,287, filed July 24, 2002, each of which are incorporated herein by reference in their entirety.

1. INTRODUCTION

The present invention relates to a method for screening and identifying 10 compounds that modulate premature translation termination and/or nonsense-mediated messenger ribonucleic acid ("mRNA") decay by screening and identifying compounds that modulate the post-transcriptional expression of any gene with a premature translation stop codon. A compound that modulates premature translation termination and/or nonsense-mediated mRNA decay of a target gene is identified using standard methods known in the 15 art to measure changes in translation or mRNA stability of the gene product or mRNA of the gene with the premature stop codon. The methods of the present invention provide a simple, sensitive assay for high-throughput screening of libraries of compounds to identify pharmaceutical leads.

2. BACKGROUND OF THE INVENTION

20 Protein synthesis encompasses the processes of translation initiation, elongation, and termination, each of which has evolved to occur with great accuracy and has the capacity to be a regulated step in the pathway of gene expression. Recent studies, including those suggesting that events at termination may regulate the ability of ribosomes to recycle to the start site of the same mRNA, have underscored the potential of termination 25 to regulate other aspects of translation. The RNA triplets UAA, UAG, and UGA are non-coding and promote translational termination. Termination starts when one of the three termination codons enters the A site of the ribosome signaling the polypeptide chain release factors to bind and recognize the termination signal. Subsequently, the ester bond between

the 3' nucleotide of the transfer RNA ("tRNA") located in the ribosome's P site and the nascent polypeptide chain is hydrolyzed, the completed polypeptide chain is released, and the ribosome subunits are recycled for another round of translation.

Nonsense-mediated mRNA decay is a surveillance mechanism that
5 minimizes the translation and regulates the RNA stability of nonsense RNAs that contain chain termination mutations (see, e.g., Hentze & Kulozik, 1999, Cell 96:307-310; Culbertson, 1999, Trends in Genetics 15:74-80; Li & Wilkinson, 1998, Immunity 8:135-141; and Ruiz-Echevarria *et al.*, 1996, Trends in Biological Sciences, 21:433-438). Chain termination mutations are caused by a base substitution or frameshift mutation changes a
10 codon into a termination codon, *i.e.*, a stop codon that causes translational termination. In nonsense-mediated mRNA decay, mRNAs with premature stop codons are subject to degradation. In some cases, a truncated protein is produced if the premature stop codon is located near the end of an open reading frame.

Certain classes of known antibiotics have been characterized and found to interact with RNA. For example, the antibiotic thiostreptone binds tightly to a 60-mer from ribosomal RNA (Cundliffe *et al.*, 1990, in *The Ribosome: Structure, Function & Evolution* (Schlessinger *et al.*, eds.) American Society for Microbiology, Washington, D.C. pp. 479-490). Bacterial resistance to various antibiotics often involves methylation at specific rRNA sites (Cundliffe, 1989, Ann. Rev. Microbiol. 43:207-233). Aminoglycosidic aminocyclitol
15 (aminoglycoside) antibiotics and peptide antibiotics are known to inhibit group I intron splicing by binding to specific regions of the RNA (von Ahsen *et al.*, 1991, Nature (London) 353:368-370). Some of these same aminoglycosides have also been found to inhibit hammerhead ribozyme function (Stage *et al.*, 1995, RNA 1:95-101). In addition, certain aminoglycosides and other protein synthesis inhibitors have been found to interact
20 with specific bases in 16S rRNA (Woodcock *et al.*, 1991, EMBO J. 10:3099-3103). An oligonucleotide analog of the 16S rRNA has also been shown to interact with certain aminoglycosides (Purohit *et al.*, 1994, Nature 370:659-662). A molecular basis for hypersensitivity to aminoglycosides has been found to be located in a single base change in mitochondrial rRNA (Hutchin *et al.*, 1993, Nucleic Acids Res. 21:4174-4179).
25 Aminoglycosides have also been shown to inhibit the interaction between specific structural RNA motifs and the corresponding RNA binding protein. Zapp *et al.* (Cell, 1993, 74:969-978) has demonstrated that the aminoglycosides neomycin B, lividomycin A, and tobramycin can block the binding of Rev, a viral regulatory protein required for viral gene expression, to its viral recognition element in the IIB (or RRE) region of HIV RNA. This

blockage appears to be the result of competitive binding of the antibiotics directly to the RRE RNA structural motif.

Aminoglycosides have also been found to promote nonsense suppression (see, e.g., Bedwell *et al.*, 1997, Nat. Med. 3:1280-1284 and Howard *et al.*, 1996, Nat. Med.

5 2:467-469). Nonsense mutations cause approximately 10 to 30 percent of the individual cases of virtually all inherited diseases. Although nonsense mutations inhibit the synthesis of a full length protein to one percent or less of wild-type levels, minimally boosting the expression levels of the full length protein to between five and fifteen percent of normal levels can greatly reduce the severity or eliminate the disease. Clinical approaches that
10 target the translation termination event to promote nonsense suppression have recently been described for model systems of cystic fibrosis and muscular dystrophy. Gentamicin is an aminoglycoside antibiotic that causes translational misreading and allowed the insertion of amino acids at the site of the nonsense codon in models of cystic fibrosis, Hurlers Syndrome, and muscular dystrophy (see, e.g., Barton-Davis *et al.*, 1999, J. Clin. Invest.
15 104:375-381). These results strongly suggest that drugs that promote nonsense suppression by altering translation termination efficiency of a premature termination codon can be therapeutically valuable in the treatment of diseases caused by nonsense mutations.

Citation or identification of any reference in Section 2 of this application is not an admission that such reference is available as prior art to the present invention.

20 3. **SUMMARY OF THE INVENTION**

The present invention provides methods for identifying a compound that modulates premature translation termination and/or nonsense-mediated mRNA decay. In particular, the invention provides methods for identifying a compound that suppresses premature translation termination and/or nonsense-mediated mRNA decay. The invention
25 encompasses the use of the compounds identified utilizing the methods of the invention for the prevention, treatment, management or amelioration of a disorder associated with, characterized by or caused by premature translation termination and/or nonsense-mediated mRNA decay, or a symptom thereof.

The invention provides cell-based and cell-free assays for the identification
30 of a compound that modulates premature translation termination and/or nonsense-mediated mRNA decay. In general, the level of expression of a reporter gene product past the premature termination codon and/or activity of such a gene product in the reporter gene based-assays described herein is indicative of the effect of the compound on premature

translation termination and/or nonsense-mediated mRNA decay. The reporter gene-based assays described herein for the identification of compounds that modulate premature translation termination and/or nonsense-mediated mRNA decay are well suited for high-throughput screening.

5 The reporter gene cell-based assays may be conducted by contacting a compound with a cell containing a nucleic acid sequence comprising a reporter gene, wherein the reporter gene comprises a premature stop codon or nonsense mutation, and measuring the expression of the reporter gene. The reporter gene cell-based assays may also be conducted by: (a) contacting a compound with a cell containing a first nucleic acid
10 sequence and a second nucleic acid sequence, wherein the first nucleic acid sequence comprises a regulatory element operably linked to a reporter gene and the second nucleic acid sequence comprises a nucleotide sequence with a premature stop codon or nonsense mutation that encodes a regulatory protein that binds to the regulatory element of the first nucleic acid sequence and regulates the expression of the reporter gene; and (b) measuring
15 the expression of the reporter gene.

The reporter gene cell-based assays may also be conducted by: (a) contacting a compound with a cell containing a first nucleic acid sequence, a second nucleic acid sequence and a third nucleic acid sequence, wherein (i) the first nucleic acid sequence comprises a nucleotide sequence encoding a first fusion protein comprising a DNA binding domain and a first protein, the nucleotide sequence of the DNA binding domain or the first protein containing a premature stop codon or nonsense mutation, (ii) the second nucleic acid sequence comprises a nucleotide sequence encoding a second fusion protein comprising an activation domain and a second protein, the second protein interacting with the first protein to produce a regulatory protein, and (iii) the third nucleic acid sequence comprises a regulatory element operably linked to a reporter gene, the expression of the reporter gene being regulated by the binding of the regulatory protein to the regulatory element: and (b) measuring the expression of the reporter gene. Further, the reporter gene cell-based assays may be conducted by: (a) contacting a compound with a cell containing a first nucleic acid sequence, a second nucleic acid sequence and a third nucleic acid sequence, wherein (i) the first nucleic acid sequence comprises a nucleotide sequence encoding a first fusion protein comprising a DNA binding domain and a first protein, (ii) the second nucleic acid sequence comprises a nucleotide sequence encoding a second fusion protein comprising an activation domain and a second protein, the nucleotide sequence of the activation domain or the second protein containing a premature stop codon or nonsense mutation, and the second
20 25 30

protein interacting with the first protein to produce a regulatory protein, and (iii) the third nucleic acid sequence comprises a regulatory element operably linked to a reporter gene, the expression of the reporter gene being regulated by the binding of the regulatory protein to the regulatory element: and (b) measuring the expression of the reporter gene.

5 The reporter gene cell-free assays may be conducted by contacting a compound with a cell-free extract and a nucleic acid sequence comprising a reporter gene, wherein the reporter gene comprises a premature stop codon or nonsense mutation, and measuring the expression of the reporter gene. The reporter gene cell-free assays may also be conducted by contacting a compound with a cell-free extract and an *in vitro* transcribed 10 RNA of a reprotoer gene, wherein the RNA product contains a premature stop codon or nonsense mutation, and measuring the expression of the protein encoded by the RNA product. The reporter gene cell-free assays may also be conducted by: (a) contacting a compound with a cell-free extract, a first nucleic acid sequence and a second nucleic acid sequence, wherein the first nucleic acid sequence comprises a regulatory element operably 15 linked to a reporter gene and the second nucleic acid sequence comprises a nucleotide sequence with a premature stop codon or nonsense mutation that encodes a regulatory protein that binds to the regulatory element of the first nucleic acid sequence and regulates the expression of the reporter gene; and (b) measuring the expression of the reporter gene.

 The reporter gene cell-free assays may also be conducted by: (a) contacting a compound with a cell-free extract, a first nucleic acid sequence, a second nucleic acid sequence and a third nucleic acid sequence, wherein (i) the first nucleic acid sequence comprises a nucleotide sequence encoding a first fusion protein comprising a DNA binding domain and a first protein, the nucleotide sequence of the DNA binding domain or the first protein containing a premature stop codon or nonsense mutation, (ii) the second nucleic acid 20 sequence comprises a nucleotide sequence encoding a second fusion protein comprising an activation domain and a second protein, the second protein interacting with the first protein to produce a regulatory protein, and (iii) the third nucleic acid sequence comprises a regulatory element operably linked to a reporter gene, the expression of the reporter gene being regulated by the binding of the regulatory protein to the regulatory element: and (b) 25 measuring the expression of the reporter gene. The reporter gene cell-free assays may also be conducted by: (a) contacting a compound with a cell-free extract, a first nucleic acid sequence, a second nucleic acid sequence and a third nucleic acid sequence, wherein (i) the first nucleic acid sequence comprises a nucleotide sequence encoding a first fusion protein comprising a DNA binding domain and a first protein, (ii) the second nucleic acid sequence 30 comprising a DNA binding domain and a first protein, (ii) the second nucleic acid sequence

comprises a nucleotide sequence encoding a second fusion protein comprising an activation domain and a second protein, the nucleotide sequence of the activation domain or the second protein containing a premature stop codon or nonsense mutation, and the second protein interacting with the first protein to produce a regulatory protein, and (iii) the third 5 nucleic acid sequence comprises a regulatory element operably linked to a reporter gene, the expression of the reporter gene being regulated by the binding of the regulatory protein to the regulatory element; and (b) measuring the expression of the reporter gene.

In the cell-based and cell-free reporter gene assays described herein, the alteration in reporter gene expression or activity relative to a previously determined 10 reference range, or to the expression or activity of the reporter gene in the absence of the compound or the presence of an appropriate control (e.g., a negative control) indicates that a particular compound modulates premature translation termination and/or nonsense-mediated mRNA decay. In particular, a decrease in reporter gene expression or activity relative to a previously determined reference range, or to the expression in the absence of the compound 15 or the presence of an appropriate control (e.g., a negative control) may, depending upon the parameters of the reporter gene assay, indicate that a particular compound reduces or suppresses premature translation termination and/or nonsense-mediated mRNA decay. In contrast, an increase in reporter gene expression or activity relative to a previously determined reference range, or to the expression in the absence of the compound or the 20 presence of an appropriate control (e.g., a negative control) may, depending upon the parameters of the reporter gene-based assay, indicate that a particular compound enhances premature translation termination and/or nonsense-mediated mRNA decay.

The invention relates to the identification of compounds that modulate premature translation termination or nonsense-mediated mRNA decay, using, in some 25 instances, a reporter based assay. The invention provides for the identification of compounds that modulated premature translation termination via a nonsense stop codon in a nucleic acids. Such nucleic acids include, but are not limited to, DNA and RNA. In a more certain embodiment, the nucleic acid is RNA. In another embodiment, the nucleic acid is single stranded. In other embodiments, the nucleic acids are single stranded. In yet other 30 embodiments, the nucleic acids are more than single stranded, e.g., double, triple or quadruple stranded.

In one embodiment, the invention provides a method for identifying a compound that modulates premature translation termination and/or nonsense-mediated mRNA decay, said method comprising: (a) expressing a nucleic acid sequence comprising a

reporter gene in a cell, wherein the reporter gene comprises a premature stop codon; (b) contacting said cell with a member of a library of compounds; and (c) detecting the expression of said reporter gene, wherein a compound that modulates premature translation termination and/or nonsense-mediated mRNA decay is identified if the expression of said reporter gene in the presence of a compound is altered relative to a previously determined reference range, or the expression of said reporter gene in the absence of the compound or the presence of an appropriate control (e.g., a negative control such as phosphate buffered saline).

In another embodiment, the invention provides a method for identifying a compound that modulates premature translation termination and/or nonsense-mediated mRNA decay, said method comprising: (a) contacting a member of a library of compounds with a cell containing a nucleic acid sequence comprising a reporter gene, wherein the reporter gene comprises a premature stop codon; and (b) detecting the expression of said reporter gene, wherein a compound that modulates premature translation termination and/or nonsense-mediated mRNA decay is identified if the expression of said reporter gene in the presence of a compound is altered relative to a previously determined reference range, or the expression of said reporter gene in the absence of said compound or the presence of an appropriate control (e.g., a negative control).

In another embodiment, the invention provides a method for identifying a compound that modulates premature translation termination and/or nonsense-mediated mRNA decay, said method comprising: (a) contacting a member of a library of compounds with a cell-free extract and a nucleic acid sequence comprising a reporter gene, wherein the reporter gene comprises a premature stop codon; and (b) detecting the expression of said reporter gene, wherein a compound that modulates premature translation termination and/or nonsense-mediated mRNA decay is identified if the expression of said reporter gene in the presence of a compound is altered relative to a previously determined reference range, or the expression of said reporter gene in the absence of said compound or the presence of an appropriate control (e.g., a negative control). In accordance with this embodiment, the cell-extract is preferably isolated from cells that have been incubated at about 0°C to about 10°C and/or an S10 to S30 cell-free extract.

In another embodiment, the invention provides a method for identifying a compound that modulates premature translation termination and/or nonsense-mediated mRNA decay, said method comprising: (a) contacting a member of a library of compounds with a cell containing a first nucleic acid sequence and a second nucleic acid sequence,

wherein the first nucleic acid sequence comprises a regulatory element operably linked to a reporter gene and the second nucleic acid sequence comprises a nucleotide sequence with a premature stop codon that encodes a regulatory protein that binds to the regulatory element of the first nucleic acid sequence and regulates the expression of the reporter gene; and (b) 5 detecting the expression of the reporter gene, wherein a compound that modulates premature translation termination and/or nonsense-mediated mRNA decay is identified if the expression of said reporter gene in the presence of a compound is altered relative to a previously determined reference range, or the expression of said reporter gene in the absence of said compound or the presence of an appropriate control (e.g., a negative 10 control).

In another embodiment, the invention provides a method for identifying a compound that modulates premature translation termination and/or nonsense-mediated mRNA decay, said method comprising: (a) contacting a member of a library of compounds with a cell containing a first nucleic acid sequence, a second nucleic acid sequence and a third nucleic acid sequence, wherein (i) the first nucleic acid sequence comprises a nucleotide sequence encoding a first fusion protein comprising a DNA binding domain and a first protein, the nucleotide sequence of the DNA binding domain or the first protein comprising a premature stop codon, (ii) the second nucleic acid sequence comprises a nucleotide sequence encoding a second fusion protein comprising an activation domain and a second protein, the second protein interacting with the first protein to produce a regulatory protein, and (iii) the third nucleic acid sequence comprises a regulatory element operably linked to a reporter gene, the expression of the reporter gene being regulated by the binding of the regulatory protein to the regulatory element; and (b) detecting the expression of the reporter gene, wherein a compound that modulates premature translation termination and/or nonsense-mediated mRNA decay is identified if the expression of said reporter gene in the presence of a compound is altered relative to a previously determined reference range, or the expression of said reporter gene in the absence of said compound or the presence of an appropriate control (e.g., a negative control). 15 20 25

In another embodiment, the invention provides a method for identifying a compound that modulates premature translation termination and/or nonsense-mediated mRNA decay, said method comprising: (a) contacting a member of a library of compounds with a cell containing a first nucleic acid sequence, a second nucleic acid sequence and a third nucleic acid sequence, wherein (i) the first nucleic acid sequence comprises a nucleotide sequence encoding a first fusion protein comprising a DNA binding domain and

a first protein, (ii) the second nucleic acid sequence comprises a nucleotide sequence encoding a second fusion protein comprising an activation domain and a second protein, the nucleotide sequence of the activation domain or the second protein containing a premature stop codon, and the second protein interacting with the first protein to produce a regulatory protein, and (iii) the third nucleic acid sequence comprises a regulatory element operably linked to a reporter gene, the expression of the reporter gene being regulated by the binding of the regulatory protein to the regulatory element; and (b) detecting the expression of the reporter gene, wherein a compound that modulates premature translation termination and/or nonsense-mediated mRNA decay is identified if the expression of said reporter gene in the presence of a compound is altered relative to a previously determined reference range, or the expression of said reporter gene in the absence of said compound or the presence of an appropriate control (e.g., a negative control).

In another embodiment, the invention provides a method for identifying a compound that modulates premature translation termination and/or nonsense-mediated mRNA decay, said method comprising: (a) contacting a member of a library of compounds with a cell-free extract, a first nucleic acid sequence and a second nucleic acid sequence, wherein the first nucleic acid sequence comprises a regulatory element operably linked to a reporter gene and the second nucleic acid sequence comprises a nucleotide sequence with a premature stop codon that encodes a regulatory protein that binds to the regulatory element of the first nucleic acid sequence and regulates the expression of the reporter gene; and (b) detecting the expression of the reporter gene, wherein a compound that modulates premature translation termination and/or nonsense-mediated mRNA decay is identified if the expression of said reporter gene in the presence of a compound is altered relative to a previously determined reference range, or the expression of said reporter gene in the absence of said compound or the presence of an appropriate control (e.g., a negative control).

In another embodiment, the invention provides a method for identifying a compound that modulates premature translation termination and/or nonsense-mediated mRNA decay, said method comprising: (a) contacting a member of a library of compounds with a cell-free extract, a first nucleic acid sequence, a second nucleic acid sequence and a third nucleic acid sequence, wherein (i) the first nucleic acid sequence comprises a nucleotide sequence encoding a first fusion protein comprising a DNA binding domain and a first protein, the nucleotide sequence of the DNA binding domain or the first protein comprising a premature stop codon, (ii) the second nucleic acid sequence comprises a

nucleotide sequence encoding a second fusion protein comprising an activation domain and a second protein, the second protein interacting with the first protein to produce a regulatory protein, and (iii) the third nucleic acid sequence comprises a regulatory element operably linked to a reporter gene, the expression of the reporter gene being regulated by the binding of the regulatory protein to the regulatory element; and (b) detecting the expression of the reporter gene, wherein a compound that modulates premature translation termination and/or nonsense-mediated mRNA decay is identified if the expression of said reporter gene in the presence of a compound is altered relative to a previously determined reference range, or the expression of said reporter gene in the absence of said compound or the presence of an appropriate control (e.g., a negative control).

In another embodiment, the invention provides a method for identifying a compound that modulates premature translation termination and/or nonsense-mediated mRNA decay, said method comprising: (a) contacting a member of a library of compounds with a cell-free extract, a first nucleic acid sequence, a second nucleic acid sequence and a third nucleic acid sequence, wherein (i) the first nucleic acid sequence comprises a nucleotide sequence encoding a first fusion protein comprising a DNA binding domain and a first protein, (ii) the second nucleic acid sequence comprises a nucleotide sequence encoding a second fusion protein comprising an activation domain and a second protein, the nucleotide sequence of the activation domain or the second protein containing a premature stop codon, and the second protein interacting with the first protein to produce a regulatory protein, and (iii) the third nucleic acid sequence comprises a regulatory element operably linked to a reporter gene, the expression of the reporter gene being regulated by the binding of the regulatory protein to the regulatory element; and (b) detecting the expression of the reporter gene, wherein a compound that modulates premature translation termination and/or nonsense-mediated mRNA decay is identified if the expression of said reporter gene in the presence of a compound is altered relative to a previously determined reference range, or the expression of said reporter gene in the absence of said compound or the presence of an appropriate control (e.g., a negative control).

In accordance with the invention, the step of contacting a compound with a cell, or cell-free extract and a nucleic acid sequence in the reporter gene-based assays described herein is preferably conducted in an aqueous solution comprising a buffer and a combination of salts (such as KCl, NaCl and/or MgCl₂). The optimal concentration of each salt used in the aqueous solution is dependent on, e.g., the protein, polypeptide or peptide encoded by the nucleic acid sequence (e.g., the regulatory protein) and the compounds used,

and can be determined using routine experimentation. In a specific embodiment, the aqueous solution approximates or mimics physiologic conditions. In another specific embodiment, the aqueous solution further comprises a detergent or a surfactant.

The assays of the present invention can be performed using different

- 5 incubation times. In the a cell-based system, the cell and a compound or a member of a library of compounds may be incubated together for at least 0.2 hours, 0.25 hours, 0.5 hours, 1 hour, 2 hours, 3 hours, 4 hours, 5 hours, 6 hours, 8 hours, 10 hours, 12 hours, 18 hours, at least 1 day, at least 2 days or at least 3 days before the expression and/or activity of a reporter gene is measured. In a cell-free system, the cell-free extract and the nucleic acid
- 10 sequence(s) (e.g., a reporter gene) can be incubated together before the addition of a compound or a member of a library of compounds. In certain embodiments, the cell-free extract are incubated with a nucleic acid sequence(s) (e.g., a reporter gene) before the addition of a compound or a member of a library of compounds for at least 0.2 hours, 0.25 hours, 0.5 hours, 1 hour, 2 hours, 3 hours, 4 hours, 5 hours, 6 hours, 8 hours, 10 hours, 12
- 15 hours, 18 hours, or at least 1 day. In other embodiments, the cell-free extract, or the nucleic acid sequence(s) (e.g., a reporter gene) is incubated with a compound or a member of a library of compounds before the addition of the nucleic acid sequence(s) (e.g., a reporter gene), or the cell-free extract, respectively. In certain embodiments, a compound or a member of a library of compounds is incubated with a nucleic acid sequence(s) (e.g., a
- 20 reporter gene) or cell-free extract before the addition of the remaining component, i.e., cell-free extract, or a nucleic acid sequence(s) (e.g., a reporter gene), respectively, for at least 0.2 hours, 0.25 hours, 0.5 hours, 1 hour, 2 hours, 3 hours, 4 hours, 5 hours, 6 hours, 8 hours, 10 hours, 12 hours, 18 hours, or at least 1 day. Once the reaction vessel comprises the components, i.e., a compound or a member of a library of compounds, the cell-free extract
- 25 and the nucleic acid sequence(s) (e.g., a reporter gene), the reaction may be further incubated for at least 0.2 hours, 0.25 hours, 0.5 hours, 1 hour, 2 hours, 3 hours, 4 hours, 5 hours, 6 hours, 8 hours, 10 hours, 12 hours, 18 hours, or at least 1 day.

- The progress of the reaction in the reporter gene-based assays can be measured continuously. Alternatively, time-points may be taken at different times of the
- 30 reaction to monitor the progress of the reaction in the reporter gene-based assays.

The reporter gene-based assays described herein may be conducted in a cell genetically engineered to express a reporter gene or *in vitro* utilizing a cell-free extract. Any cell or cell line of any species well-known to one of skill in the art may be utilized in accordance with the methods of the invention. Further, a cell-free extract may be derived

from any cell or cell line of any species well-known to one of skill in the art. Examples of cells and cell types include, but are not limited to, human cells, cultured mouse cells, cultured rat cells or Chinese hamster ovary ("CHO") cells.

The reporter gene constructs utilized in the reporter gene-based assays

5 described herein may comprise the coding region of a reporter gene and a premature stop codon that results in premature translation termination and/or nonsense-mediated mRNA decay. Preferably, the premature stop codon is N-terminal to the native stop codon of the reporter gene and is located such that the suppression of the premature stop codon is readily detectable. In a specific embodiment, a reporter gene construct utilized in the reporter gene-based assays described herein comprises the coding region of a reporter gene containing a premature stop codon at least 15 nucleotides, preferably 25 to 50 nucleotides, 50 to 75 nucleotides or 75 to 100 nucleotides from the start codon in the open reading frame of the reporter gene. In another embodiment, a reporter gene construct utilized in the reporter gene-based assays described herein comprises the coding region of a reporter gene
10 containing a premature stop codon at least 15 nucleotides, preferably 25 to 50 nucleotides, 50 to 75 nucleotides, 75 to 100 nucleotides, or 100 to 150 nucleotides from the native stop codon in the open reading frame of the reporter gene. In another embodiment, a reporter gene construct utilized in the reporter gene-based assays described herein comprises the coding region of a reporter gene containing a UAG and/or UGA premature stop codon. In
15 yet another embodiment, a reporter gene construct utilized in the reporter gene based assays described herein comprises the coding region of a reporter gene, containing a premature stop codon in the context of UGAA, UGAC, UGAG, UGAU, UAGA, UAGC, UAGG, UAGU, UAAA, UAAC, UAAG or UAAU..

Alternatively, the reporter gene constructs utilized in the reporter gene-based

20 assays described herein comprise a regulatory element that is responsive to a regulatory protein encoded by a nucleic acid sequence containing a premature stop codon. Preferably, the premature stop codon in the nucleotide sequence of a regulatory protein or a component or subunit thereof is N-terminal to the native stop codon of the regulatory protein or component or subunit thereof and the location of the premature stop codon is such that it
25 alters the biological activity of the regulatory protein (e.g., the ability of the regulatory protein to bind to its regulatory element). In a specific embodiment, the premature stop codon in the nucleotide sequence of a regulatory protein or a component or subunit thereof is at least 15 nucleotides preferably 25 to 50 nucleotides, 50 to 75 nucleotides or 75 to 100 nucleotides from the start codon in the open reading frame of the regulatory protein,
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component or subunit thereof. In another embodiment, the premature stop codon in the nucleotide sequence of a regulatory protein or a component or subunit thereof is at least 15 nucleotides, preferably 25 to 50 nucleotides, 50 to 75 nucleotides, 75 to 100 nucleotides, or 100 to 150 nucleotides from the native stop codon in the open reading frame of the

5 regulatory protein, component or subunit thereof. In another embodiment, the premature stop codon in the nucleotide sequence of regulatory protein or a component or subunit thereof is UAG or UGA. Any reporter gene well-known to one of skill in the art may be utilized in the reporter gene constructs described herein. Examples of reporter genes include, but are not limited to, the gene encoding firefly luciferase, the gene coding renilla

10 luciferase, the gene encoding click beetle luciferase, the gene encoding green fluorescent protein, the gene encoding yellow fluorescent protein, the gene encoding red fluorescent protein, the gene encoding cyan fluorescent protein, the gene encoding blue fluorescent protein, the gene encoding beta-galactosidase, the gene encoding beta-glucuronidase, the gene encoding beta-lactamase, the gene encoding chloramphenicol acetyltransferase, and

15 the gene encoding alkaline phosphatase.

The compounds utilized in the assays described herein may be members of a library of compounds. In specific embodiment, the compound is selected from a combinatorial library of compounds comprising peptoids; random biooligomers; diversomers such as hydantoins, benzodiazepines and dipeptides; vinylogous polypeptides; 20 nonpeptidal peptidomimetics; oligocarbamates; peptidyl phosphonates; peptide nucleic acid libraries; antibody libraries; carbohydrate libraries; and small organic molecule libraries. In a preferred embodiment, the small organic molecule libraries are libraries of benzodiazepines, isoprenoids, thiazolidinones, metathiazanones, pyrrolidines, morpholino compounds, or diazepindiones.

25 In certain embodiments, the compounds are screened in pools. Once a positive pool has been identified, the individual compounds of that pool are tested separately. In certain embodiments, the pool size is at least 2, at least 5, at least 10, at least 25, at least 50, at least 75, at least 100, at least 150, at least 200, at least 250, or at least 500 compounds.

30 Once a compound that modulates premature translation termination and/or nonsense-mediated mRNA decay is identified, the structure of the compound may be determined utilizing well-known techniques or by referring to a predetermined code. For example, the structure of the compound may be determined by mass spectroscopy, NMR, vibrational spectroscopy, or X-ray crystallography.

A compound identified in accordance with the methods of the invention may directly bind to the mRNA translation machinery. Alternatively, a compound identified in accordance with the methods of invention may bind to the premature stop codon. A compound identified in accordance with the methods of invention may also disrupt an interaction between a premature stop codon and the mRNA translation machinery. In a preferred embodiment, a compound identified in accordance with the methods of the invention suppresses premature translation termination and/or nonsense-mediated mRNA decay of a gene encoding a protein, polypeptide or peptide whose expression is beneficial to a subject. In another preferred embodiment, a compound identified in accordance with the methods of the invention increases premature translation termination and/or nonsense-mediated mRNA decay of a gene encoding a protein, polypeptide or peptide whose expression is detrimental to a subject. In a specific embodiment, a compound identified in accordance with the methods of the invention preferentially or differentially modulates premature translation termination and/or nonsense-mediated mRNA decay of a specific nucleotide sequence of interest relative to another nucleotide sequence, as measured by an assay described herein or well known to one of skill in the art under the same or similar assay conditions.

In a specific embodiment, a compound identified in accordance with the invention suppresses premature translation termination or nonsense-mediated mRNA decay of a specific nucleotide sequence of interest by at least 5%, preferably at least 10%, at least 15%, at least 20%, at least 25%, at least 30%, at least 35%, at least 40%, at least 45%, at least 50%, at least 55%, at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, or at least 95%, relative to an appropriate control (*e.g.*, a negative control such as PBS), in an assay described herein under the same or similar assay conditions. In accordance with this embodiment, preferably, the compound differentially or preferentially suppresses the nucleotide sequence of interest relative to another nucleotide sequence.

In certain embodiments of the invention, the compound identified using the assays described herein is a small molecule. In a preferred embodiment, the compound identified using the assays described herein is not known to affect premature translation termination and/or nonsense-mediated mRNA decay of a nucleic acid sequence, in particular a nucleic acid sequence of interest. In another preferred embodiment, the compound identified using the assays described herein has not been used as or suggested to be used in the prevention, treatment, management and/or amelioration of a disorder

associated with, characterized by or caused by a premature stop codon. In another preferred embodiment, the compound identified using the assays described herein has not been used as or suggested to be used in the prevention, treatment, management and/or amelioration of a particular disorder described herein.

5 A compound identified in accordance with the methods of the invention may be tested in *in vitro* and/or *in vivo* assays well-known to one of skill in the art or described herein to determine the prophylactic or therapeutic effect of a particular compound for a particular disorder. In particular, a compound identified utilizing the assays described herein may be tested in an animal model to determine the efficacy of the compound in the 10 prevention, treatment or amelioration of a disorder associated with, characterized by or caused by a premature stop codon, or a disorder described herein, or a symptom thereof. In addition, a compound identified utilizing the assays described herein may be tested for its toxicity in *in vitro* and/or *in vivo* assays well-known to one of skill in the art.

15 The invention provides for methods for preventing, treating, managing or ameliorating a disorder associated with, characterized by or caused by a premature stop codon or a symptom thereof, said method comprising administering to a subject in need thereof a therapeutically or prophylactically effective amount of a compound, or a pharmaceutically acceptable salt thereof, identified according to the methods described herein.

20 The present invention may be understood more fully by reference to the detailed description and examples, which are intended to illustrate non-limiting embodiments of the invention.

3.1. Terminology

25 As used herein, the term "compound" refers to any agent or complex that is being tested for its ability to modulate premature translation termination and/or nonsense-mediated mRNA decay or has been identified as modulating premature translation termination and/or nonsense-mediated mRNA decay.

30 As used herein, the terms "disorder" and "disease" are to refer to a condition in a subject. In a specific embodiment, the terms disease and disorder refer to a condition in a subject that is associated with, characterized by, or caused by premature translation termination and/or nonsense-mediated mRNA decay of one or more gene products. Non-limiting examples of such disease and disorders are described herein below.

As used herein, the term "effective amount" refers to the amount of a compound which is sufficient to (i) reduce or ameliorate the progression, severity and/or

duration of a disorder associated with, characterized by or caused by premature translation termination and/or nonsense-mediated mRNA decay, or one or more symptoms thereof, (ii) prevent the development, recurrence or onset of a disorder associated with, characterized by or caused by premature translation termination and/or nonsense-mediated mRNA decay, or 5 one or more symptoms thereof, (iii) prevent the advancement of a disorder associated with, characterized by or caused by premature translation termination and/or nonsense-mediated mRNA decay, or one or more symptoms thereof, or (iv) enhance or improve the therapeutic(s) effect(s) of another therapy.

As used herein, the term "host cell" includes a particular subject cell 10 transfected with a nucleic acid molecule and the progeny or potential progeny of such a cell. Progeny of such a cell may not be identical to the parent cell transfected with the nucleic acid molecule due to mutations or environmental influences that may occur in succeeding generations or integration of the nucleic acid molecule into the host cell genome.

As used herein, the term "in combination" refers to the use of more than one 15 therapy (e.g., prophylactic and/or therapeutic agents). The use of the term "in combination" does not restrict the order in which therapies (e.g., prophylactic and/or therapeutic agents) are administered to a subject with a disorder associated with, characterized by or caused by premature translation termination and/or nonsense-mediated mRNA decay. A first therapy (e.g., a prophylactic or therapeutic agent such as a compound identified in accordance with 20 the methods of the invention) can be administered prior to (e.g., 5 minutes, 15 minutes, 30 minutes, 45 minutes, 1 hour, 2 hours, 4 hours, 6 hours, 12 hours, 24 hours, 48 hours, 72 hours, 96 hours, 1 week, 2 weeks, 3 weeks, 4 weeks, 5 weeks, 6 weeks, 8 weeks, or 12 weeks before), concomitantly with, or subsequent to (e.g., 5 minutes, 15 minutes, 30 minutes, 45 minutes, 1 hour, 2 hours, 4 hours, 6 hours, 12 hours, 24 hours, 48 hours, 72 hours, 96 hours, 1 week, 2 weeks, 3 weeks, 4 weeks, 5 weeks, 6 weeks, 8 weeks, or 12 weeks after) the administration of a second therapy (e.g., a prophylactic or therapeutic agent such as a chemotherapeutic agent or a TNF- α antagonist) to a subject with a disorder 25 associated with, characterized by or caused by premature translation termination and/or nonsense-mediated mRNA decay.

As used herein, the term "library" in the context of compounds refers to a 30 plurality of compounds. A library can be a combinatorial library, e.g., a collection of compounds synthesized using combinatorial chemistry techniques, or a collection of unique chemicals of low molecular weight (less than 1000 daltons) that each occupy a unique three-dimensional space.

As used herein, the terms "manage", "managing" and "management" refer to the beneficial effects that a subject derives from a therapy (*e.g.*, a prophylactic or therapeutic agent) which does not result in a cure of the disorder associated with, characterized by or caused by premature translation termination and/or nonsense-mediated mRNA decay. In certain embodiments, a subject is administered one or more therapies to "manage" a disease or disorder so as to prevent the progression or worsening of the disease or disorder.

As used herein, the phrase "modulation of premature translation termination and/or nonsense-mediated mRNA decay" refers to the regulation of gene expression by altering the level of nonsense suppression. For example, if it is desirable to increase production of a defective protein encoded by a gene with a premature stop codon, *i.e.*, to permit readthrough of the premature stop codon of the disease gene so translation of the gene can occur, then modulation of premature translation termination and/or nonsense-mediated mRNA decay entails up-regulation of nonsense suppression. Conversely, if it is desirable to promote the degradation of an mRNA with a premature stop codon, then modulation of premature translation termination and/or nonsense-mediated mRNA decays entails down-regulation of nonsense suppression.

As used herein, the terms "non-responsive" and "refractory" describe patients treated with a currently available therapy (*e.g.*, prophylactic or therapeutic agent) for a disorder associated with, characterized by or caused by premature translation termination and/or nonsense-mediated mRNA decay (*e.g.*, cancer), which is not clinically adequate to relieve one or more symptoms associated with such disorder. Typically, such patients suffer from severe, persistently active disease and require additional therapy to ameliorate the symptoms associated with their disorder.

As used herein, "nonsense-mediated mRNA decay" refers to any mechanism that mediates the decay of mRNAs containing a premature translation termination codon.

As used herein, a "nonsense mutation" is a point mutation changing a codon corresponding to an amino acid to a stop codon.

As used herein, "nonsense suppression" refers to the inhibition or suppression of premature translation termination and/or nonsense-mediated mRNA decay.

The terms "nucleic acid," "nucleic acid sequence," "nucleotide sequence," and analogous terms as used herein include DNA molecules (*e.g.*, cDNA or genomic DNA), RNA molecules (*e.g.*, mRNA), combinations of DNA and RNA molecules of hybrid DNA/RNA molecules, and analogs of DNA or RNA molecules. Such analogs can be

generated using, for example, nucleotide analogs, which include, but are not limited to, inosine or tritylated bases. Such analogs can also comprise DNA or RNA molecules comprising modified backbones that lend beneficial attributes to the molecules, such as, for example, nuclease resistance or an increased ability to cross cellular membranes. The 5 nucleic acids, nucleic acid sequences or nucleotide sequences can be single-stranded, double-stranded, may contain both single-stranded and double-stranded portions, and may contain triple-stranded portions, but preferably is double-stranded DNA. In one embodiment, the nucleotide sequences comprise a contiguous open reading frame encoding a reporter gene, e.g., a cDNA molecule.

10 As used herein, the phrase "pharmaceutically acceptable salt(s)," includes, but is not limited to, salts of acidic or basic groups that may be present in compounds identified using the methods of the present invention. Compounds that are basic in nature are capable of forming a wide variety of salts with various inorganic and organic acids. The acids that can be used to prepare pharmaceutically acceptable acid addition salts of such 15 basic compounds are those that form non-toxic acid addition salts, i.e., salts containing pharmacologically acceptable anions, including but not limited to sulfuric, citric, maleic, acetic, oxalic, hydrochloride, hydrobromide, hydroiodide, nitrate, sulfate, bisulfate, phosphate, acid phosphate, isonicotinate, acetate, lactate, salicylate, citrate, acid citrate, tartrate, oleate, tannate, pantothenate, bitartrate, ascorbate, succinate, maleate, gentisinate, 20 fumarate, gluconate, glucaronate, saccharate, formate, benzoate, glutamate, methanesulfonate, ethanesulfonate, benzenesulfonate, p-toluenesulfonate and pamoate (i.e., 1,1'-methylene-bis-(2-hydroxy-3-naphthoate)) salts. Compounds that include an amino moiety may form pharmaceutically acceptable salts with various amino acids, in addition to the acids mentioned above. Compounds that are acidic in nature are capable of forming 25 base salts with various pharmacologically acceptable cations. Examples of such salts include alkali metal or alkaline earth metal salts and, particularly, calcium, magnesium, sodium lithium, zinc, potassium, and iron salts.

As used herein, "premature translation termination" refers to the result of a mutation that changes a codon corresponding to an amino acid to a stop codon.

30 As used herein, the terms "prevent", "preventing" and "prevention" refer to the prevention of the development, recurrence or onset of a disorder associated with, characterized by or caused by premature translation termination and/or nonsense-mediated mRNA decay or one or more symptoms thereof resulting from the administration of one or more compounds identified in accordance the methods of the invention or the

administration of a combination of such a compound and a known therapy for such a disorder.

As used herein, the term "previously determined reference range" refers to a reference range for the readout of a particular assay. In a specific embodiment, the term refers to a reference range for the expression of a reporter gene and/or the activity of a reporter gene product by a particular cell or in a particular cell-free extract. Each laboratory will establish its own reference range for each particular assay, each cell type and each cell-free extract. In a preferred embodiment, at least one positive control and at least one negative control are included in each batch of compounds analyzed.

As used herein, the terms "prophylactic agent" and "prophylactic agents" refer to any agent(s) which can be used in the prevention of a disorder associated with, characterized by or caused by premature translation termination and/or nonsense-mediated mRNA decay. In certain embodiments, the term "prophylactic agent" refers to a compound identified in the screening assays described herein. In certain other embodiments, the term "prophylactic agent" refers to an agent other than a compound identified in the screening assays described herein which is known to be useful for, or has been or is currently being used to prevent or impede the onset, development and/or progression of a disorder associated with, characterized by or caused by premature translation termination and/or nonsense-mediated mRNA decay or one or more symptoms thereof.

As used herein, the phrase "prophylactically effective amount" refers to the amount of a therapy (e.g., a prophylactic agent) which is sufficient to result in the prevention of the development, recurrence or onset of one or more symptoms associated with a disorder associated with, characterized by or caused by premature translation termination and/or nonsense-mediated mRNA decay.

As used herein, the term "purified" in the context of a compound, e.g., a compound identified in accordance with the method of the invention, refers to a compound that is substantially free of chemical precursors or other chemicals when chemically synthesized. In a specific embodiment, the compound is 60%, preferably 65%, 70%, 75%, 80%, 85%, 90%, or 99% free of other, different compounds. In a preferred embodiment, a compound identified in accordance with the methods of the invention is purified.

As used herein, a "premature termination codon" or "premature stop codon" refers to the occurrence of a stop codon instead of a codon corresponding to an amino acid.

As used herein, a "reporter gene" refers to a gene by which modulation of premature translation termination and/or nonsense-mediated mRNA decay is ascertained.

In a preferred embodiment, the expression of a reporter gene is easily assayed and has an activity which is not normally found in the organism of which the translation extract is derived.

As used herein, the term "small molecule" and analogous terms include, but
5 are not limited to, peptides, peptidomimetics, amino acids, amino acid analogs,
polynucleotides, polynucleotide analogs, nucleotides, nucleotide analogs, organic or
inorganic compounds (*i.e.*, including heterorganic and/or ganometallic compounds) having
a molecular weight less than about 10,000 grams per mole, organic or inorganic compounds
having a molecular weight less than about 5,000 grams per mole, organic or inorganic
10 compounds having a molecular weight less than about 1,000 grams per mole, organic or
inorganic compounds having a molecular weight less than about 500 grams per mole, and
salts, esters, and other pharmaceutically acceptable forms of such compounds.

As used herein, the terms "subject" and "patient" are used interchangeably
herein. The terms "subject" and "subjects" refer to an animal, preferably a mammal
15 including a non-primate (*e.g.*, a cow, pig, horse, cat, dog, rat, and mouse) and a primate
(*e.g.*, a chimpanzee, a monkey such as a cynomolgous monkey and a human), and more
preferably a human. In one embodiment, the subject is refractory or non-responsive to
current therapies for a disorder associated with, characterized by or caused by premature
translation termination and/or nonsense-mediated mRNA decay. In another embodiment,
20 the subject is a farm animal (*e.g.*, a horse, a cow, a pig, etc.) or a pet (*e.g.*, a dog or a cat).
In a preferred embodiment, the subject is a human.

As used herein, the term "synergistic" refers to a combination of a compound
identified using one of the methods described herein, and another therapy (*e.g.*, a
prophylactic or therapeutic agent), which combination is more effective than the additive
25 effects of the therapies. A synergistic effect of a combination of therapies (*e.g.*,
prophylactic or therapeutic agents) permits the use of lower dosages of one or more of the
therapies and/or less frequent administration of said therapies to a subject with a
proliferative disorder. The ability to utilize lower dosages of a therapy (*e.g.*, a prophylactic
30 or therapeutic agent) and/or to administer said therapy less frequently reduces the toxicity
associated with the administration of said therapy to a subject without reducing the efficacy
of said therapies in the prevention, treatment, management or amelioration of a disorder
associated with, characterized by or caused by premature translation termination and/or
nonsense-mediated mRNA decay. In addition, a synergistic effect can result in improved
efficacy of therapies (*e.g.*, agents) in the prevention, treatment, management or amelioration

of a disorder associated with, characterized by or caused by premature translation termination and/or nonsense-mediated mRNA decay. Finally, a synergistic effect of a combination of therapies (e.g., prophylactic or therapeutic agents) may avoid or reduce adverse or unwanted side effects associated with the use of either therapy alone.

5 As used herein, the terms "therapeutic agent" and "therapeutic agents" refer to any agent(s) which can be used in the prevention, treatment, management or amelioration of one or more symptoms of a disorder associated with, characterized by or caused by premature translation termination and/or nonsense-mediated mRNA decay. In certain embodiments, the term "therapeutic agent" refers to a compound identified in the screening assays described herein. In other embodiments, the term "therapeutic agent" refers to an 10 agent other than a compound identified in the screening assays described herein which is known to be useful for, or has been or is currently being used to prevent, treat, manage or ameliorate a proliferative disorder or one or more symptoms thereof.

As used herein, the term "therapeutically effective amount" refers to that 15 amount of a therapy (e.g., a therapeutic agent) sufficient to result in (i) the amelioration of one or more symptoms of a disorder associated with, characterized by or caused by premature translation termination and/or nonsense-mediated mRNA decay, (ii) prevent advancement of a disorder associated with, characterized by or caused by premature translation termination and/or nonsense-mediated mRNA decay, (iii) cause regression of a 20 disorder associated with, characterized by or caused by premature translation termination and/or nonsense-mediated mRNA decay, or (iv) to enhance or improve the therapeutic effect(s) of another therapy (e.g., therapeutic agent).

As used herein, the terms "treat", "treatment" and "treating" refer to the reduction or amelioration of the progression, severity and/or duration of a disorder 25 associated with, characterized by or caused by premature translation termination and/or nonsense-mediated mRNA decay or one or more symptoms thereof resulting from the administration of one or more compounds identified in accordance the methods of the invention, or a combination of one or more compounds identified in accordance with the invention and another therapy.

30 As used herein, the terms "therapy" and "therapies" refer to any method, protocol and/or agent that can be used in the prevention, treatment, management or amelioration of a disease or disorder or one or more symptoms thereof. In certain embodiments, such terms refer to chemotherapy, radiation therapy, surgery, supportive therapy and/or other therapies useful in the prevention, treatment, management or

amelioration of a disease or disorder or one or more symptoms thereof known to skilled medical personnel.

4. BRIEF DESCRIPTION OF THE FIGURES

- 5 FIG. 1. Translation of a wild-type luciferase RNA in the *in vitro* translation reaction. Reaction mixtures were prepared containing varying amounts of wild-type luciferase RNA and varying amounts of HeLa cell extract. The amount of luciferase produced was monitored in a Turner luminometer by the addition of luciferase substrate (Promega).
- 10 FIG. 2. Translation of a nonsense containing (UGA) luciferase RNA in the *in vitro* translation reaction. Reaction mixtures were prepared containing varying amounts of luciferase RNA harboring a UGA nonsense mutation and varying amounts of HeLa cell extract. The amount of luciferase was monitored in a Turner luminometer by the addition of luciferase substrate (Promega).
- 15 FIG. 3. Translation of wild-type luciferase RNA by incubating the cells on ice prior to lysis. HeLa cell pellets were incubated on ice or not incubated on ice prior to lysis and the effect of the incubation on the translation activity of the cell-extract was measured in an *in vitro* translation reaction for luciferase production
- 20 FIG. 4. Translation of a nonsense (UGA) containing luciferase RNA in the *in vitro* translation reaction. Reaction mixtures were prepared with luciferase RNA containing a UGA nonsense mutation. Gentamicin was (GENT) or was not added (UNT) added to the reaction mixture and the amount of luciferase produced was monitored in a Viewlux luminometer by the addition of luciferase substrate (Promega).
- 25 FIG. 5. The amount of luciferase produced was monitored in a Viewlux luminometer by the addition of luciferase substrate (Promega).
- FIG. 6A-6B. 6A: Nonsense suppression in cells harboring a luciferase nonsense allele. Stable cell lines harboring the UGA, UAA and UAG nonsense alleles of luciferase were treated overnight with Compound A, Compound B, and Gentamicin. The following day, the level of suppression was determined by measuring the amount of luminescence produced. The fold suppression above control cells treated with solvent was calculated and plotted vs.

concentration of compound. 6B: Nonsense suppression in cells harboring a luciferase nonsense allele. Stable cell lines harboring the UGA, UAA and UAG nonsense alleles of luciferase were treated overnight with Compound A, and gentamicin. The following day, the level of suppression was determined by measuring the amount of luminescence produced. The fold suppression above control cells treated with solvent was calculated and plotted vs. concentration of compound.

5 FIG. 7A-7B. Chemical footprinting analysis of Compound A on the human 28S rRNA.

10 100 pmol of ribosomes were incubated with 100 μ M compound, followed by treatment with chemical modifying agents (dimethyl sulfate [DMS] and kethoxal [KE]). Following chemical modification, rRNA was prepared and analyzed in primer extension reactions using end-labeled oligonucleotides hybridizing to rRNA. Panel A (lanes 1-3 DMS modification; lanes 4-6 KE modification): Lanes 1 and 4, DMSO treated; 2 and 5, paromomycin treated; 15 3 and 6, Compound A treated; 4. A sequencing reaction (indicated by lanes GATC in panel A) was run in parallel as a marker.

FIG. 8. 15 Functional CFTR expression monitored as cAMP-induced anion efflux using the halide-sensitive fluorophore 6-methoxy-N-(3-sulphopropyl) quinolinium (SPQ). Compound A increases cAMP-stimulated chloride channel activity

20 in cells expressing the W1282X mutation. Cells were initially loaded in a hypotonic buffer containing SPQ and sodium iodide; iodide quenches SPQ fluorescence (Yang *et al.*, 1993, Hum Mol Genet. 2(8):1253-1261). Sodium iodide in the bath was replaced by sodium nitrate at 2 min; since nitrate does not interact with SPQ, fluorescence increased as cell iodide is lost to the bath. A cAMP stimulation cocktail (10 μ M forskolin, 100 μ M cpt-cAMP and 100 μ M IBMX) was added at 6 min. Fluorescence was then quenched again by returning sodium iodide to the bath at 10 min. Functional CFTR expression was monitored as the dequenching of SPQ fluorescence caused by cAMP-induced iodide efflux.

25 FIG. 9. 30 Immunohistochemistry of myotubes from primary cell culture from mdx muscle. The presence of dystrophin was detected by mAb to the COOH-terminus of dystrophin (F192A12) followed by a rhodamine-conjugated anti-mouse IgG. Dystrophin was present in mdx myotubes treated with 20 μ M Compound A (left) and in mdx myotubes

treated with 200 μ M gentamicin (center). Little dystrophin was detected in untreated mdx myotubes (right).

FIG.10A-10F. Immunohistochemistry of muscle cross-sections to view dystrophin. C57 control tibialis anterior (TA) muscle displayed positive staining for dystrophin (panel D). Muscle cross-sections from mdx mice treated with gentamicin (200 μ M, panel A) and Compound A (10 μ M panel B; 20 μ M panel C) displayed positive staining for dystrophin. Muscle from untreated mdx mice (panel E) or from cross sections not treated with primary antibody (panel F) show only minimal staining.

10 5. **DETAILED DESCRIPTION OF THE INVENTION**

The present invention provides methods for identifying compounds that modulate premature translation termination and/or nonsense-mediated mRNA decay. In particular, the invention provides simple, rapid and sensitive methods for identifying compounds that modulate premature translation termination and/or nonsense-mediated mRNA decay. Any gene encoding a premature stop codon can be used in the cell-based and cell-free assays described herein to identify compounds that modulate premature translation termination and/or nonsense-mediated mRNA decay. The cell-based and cell-free assays described herein can be utilized in a high throughput format to screen libraries of compounds to identify those compounds that modulate premature translation termination and/or nonsense-mediated mRNA decay.

Reporter gene-based assays can be utilized to identify a compound that modulates premature translation termination and/or nonsense-mediated mRNA decay. The reporter gene-based assays described herein may be conducted by contacting a compound with a cell containing a nucleic acid sequence comprising a reporter gene, wherein said reporter gene comprises a premature stop codon, and measuring the expression and/or activity of the reporter gene. Alternatively, the reporter gene-based assays may be conducted by contacting a compound with a cell-free extract and a nucleic acid sequence comprising a reporter gene, wherein said reporter gene comprises a premature stop codon, and measuring the expression of said reporter gene. The reporter gene-based assays may also be conducted by: (a) contacting a compound with a cell containing a first nucleic acid sequence and a second nucleic acid sequence, wherein the first nucleic acid sequence comprises a reporter gene operably linked to a regulatory element and the second nucleic acid sequence comprises a nucleotide sequence encoding a regulatory protein or a subunit

thereof with a premature stop codon and the regulatory protein regulates the expression of the reporter gene; and (b) measuring the expression and/or activity of the reporter gene. Further, the reporter gene-based assays may be conducted by: (a) contacting a compound with a cell-free extract, a first nucleic acid sequence and a second nucleic acid sequence, 5 wherein the first nucleic acid sequence comprises a reporter gene operably linked to a regulatory element and the second nucleic acid sequence comprises a nucleotide sequence encoding a regulatory protein or a subunit thereof with a premature stop codon and the regulatory protein regulates the expression of the reporter gene; and (b) measuring the expression and/or activity of the reporter gene. The alteration in reporter gene expression 10 relative to a previously determined reference range, or the expression of the reporter gene in the absence of the compound or an appropriate control (*e.g.*, a negative control) in such reporter-gene based assays indicates that a particular compound modulates premature translation termination and/or nonsense-mediated mRNA decay.

The structure of the compounds identified in the assays described herein that 15 modulate changes in post-transcriptional gene regulation can be determined utilizing assays well-known to one of skill in the art or described herein. The methods used will depend, in part, on the nature of the library screened. For example, assays or microarrays of compounds, each having an address or identifier, may be deconvoluted, *e.g.*, by cross-referencing the positive sample to an original compound list that was applied to the 20 individual test assays. Alternatively, the structure of the compounds identified herein may be determined using mass spectrometry, nuclear magnetic resonance (“NMR”), X ray crystallography, or vibrational spectroscopy.

The invention encompasses the use of the compounds that modulate premature translation termination and/or nonsense-mediated mRNA decay that were 25 identified in accordance with the methods described herein. In particular, the invention encompasses the use of compounds identified as lead compounds for the development of prophylactic or therapeutic agents in the prevention, treatment, management and/or amelioration of a disease associated with, characterized by or caused by a nonsense mutation. Such diseases include, but are not limited to, cystic fibrosis, muscular dystrophy, 30 heart disease, cancer, retinitis pigmentosa, collagen disorders, Tay-Sachs disease, blood disorders, kidney stones, ataxia-telangiectasia, lysosomal storage diseases, and tuberous sclerosis.

Section 5.1 describes genes with premature translation stop codons and cells and cell-free extracts that are useful in the methods of the invention. Section 5.2 describes

libraries of compounds. Section 5.4 describes reporter gene-based assays for identifying compounds that modulate premature translation termination and/or nonsense-mediated mRNA decay. Section 5.5 describes naturally occurring genes with premature stop codons and examples of diseases associated with such genes. Section 5.6 describes secondary 5 biological screens. Section 5.7 describes the methods for designing congeners or analogs of compounds identified in accordance with the methods of the invention. Section 5.8 describes uses of compounds identified in accordance with the methods of the invention for preventing, treating, managing or ameliorating a disease or abnormal condition in a subject associated with, characterized by or caused by a premature stop codon. Section 5.9 10 describes methods of administering compounds identified in accordance with the invention to a subject in need thereof.

5.1. Reporter Gene Constructs, Transfected Cells and Cell-Free Extracts

The invention provides for reporter genes to ascertain the effects of a compound on premature translation termination and/or nonsense-mediated mRNA decay. 15 In general, the level of expression and/or activity of a reporter gene product is indicative of the effect of the compound on premature translation termination and/or nonsense-mediated mRNA decay.

The invention provides for specific vectors comprising a reporter gene operably linked to one or more regulatory elements and host cells transfected with the 20 vectors. The invention also provides for the *in vitro* translation of a reporter gene flanked by one or more regulatory elements. A reporter gene may or may not contain a premature stop codon depending on the assay conducted. Techniques for practicing this specific aspect of this invention will employ, unless otherwise indicated, conventional techniques of molecular biology, microbiology, and recombinant DNA manipulation and production, which are routinely practiced by one of skill in the art. See, e.g., Sambrook, 1989, 25 Molecular Cloning, A Laboratory Manual, Second Edition; DNA Cloning, Volumes I and II (Glover, Ed. 1985); Oligonucleotide Synthesis (Gait, Ed. 1984); Nucleic Acid Hybridization (Hames & Higgins, Eds. 1984); Transcription and Translation (Hames & Higgins, Eds. 1984); Animal Cell Culture (Freshney, Ed. 1986); Immobilized Cells and Enzymes (IRL 30 Press, 1986); Perbal, A Practical Guide to Molecular Cloning (1984); Gene Transfer Vectors for Mammalian Cells (Miller & Calos, Eds. 1987, Cold Spring Harbor Laboratory); Methods in Enzymology, Volumes 154 and 155 (Wu & Grossman, and Wu, Eds., respectively), (Mayer & Walker, Eds., 1987); Immunochemical Methods in Cell and Molecular Biology (Academic Press, London, Scopes, 1987), Expression of Proteins in

Mammalian Cells Using Vaccinia Viral Vectors in Current Protocols in Molecular Biology, Volume 2 (Ausubel et al., Eds., 1991).

5.1.1. Reporter Genes

Any reporter gene well-known to one of skill in the art may be used in reporter gene constructs to ascertain the effect of a compound on premature translation termination. Reporter genes refer to a nucleotide sequence encoding a protein, polypeptide or peptide that is readily detectable either by its presence or activity. Reporter genes may be obtained and the nucleotide sequence of the elements determined by any method well-known to one of skill in the art. The nucleotide sequence of a reporter gene can be obtained, e.g., from the literature or a database such as GenBank. Alternatively, a polynucleotide encoding a reporter gene may be generated from nucleic acid from a suitable source. If a clone containing a nucleic acid encoding a particular reporter gene is not available, but the sequence of the reporter gene is known, a nucleic acid encoding the reporter gene may be chemically synthesized or obtained from a suitable source (e.g., a cDNA library, or a cDNA library generated from, or nucleic acid, preferably poly A+ RNA, isolated from, any tissue or cells expressing the reporter gene) by PCR amplification. Once the nucleotide sequence of a reporter gene is determined, the nucleotide sequence of the reporter gene may be manipulated using methods well-known in the art for the manipulation of nucleotide sequences, e.g., recombinant DNA techniques, site directed mutagenesis, PCR, etc. (see, for example, the techniques described in Sambrook et al., 1990, Molecular Cloning, A Laboratory Manual, 2d Ed., Cold Spring Harbor Laboratory, Cold Spring Harbor, NY and Ausubel et al., eds., 1998, Current Protocols in Molecular Biology, John Wiley & Sons, NY, which are both incorporated by reference herein in their entireties), to generate reporter genes having a different amino acid sequence, for example to create amino acid substitutions, deletions, and/or insertions.

In a specific embodiment, a reporter gene is any naturally-occurring gene with a premature stop codon. Genes with premature stop codons that are useful in the present invention include, but are not limited to, the genes described below. In an alternative embodiment, a reporter gene is any gene that is not known in nature to contain a premature stop codon. Examples of reporter genes include, but are not limited to, luciferase (e.g., firefly luciferase, renilla luciferase, and click beetle luciferase), green fluorescent protein ("GFP") (e.g., green fluorescent protein, yellow fluorescent protein, red fluorescent protein, cyan fluorescent protein, and blue fluorescent protein), beta-galactosidase ("beta-gal"), beta-glucuronidase, beta-lactamase, chloramphenicol acetyltransferase ("CAT"), and

alkaline phosphatase ("AP"). Alternatively, a reporter gene can also be a protein tag, such as, but not limited to, myc, His, FLAG, or GST, so that nonsense suppression will produce the peptide and the protein can be monitored by an ELISA, a western blot, or any other immunoassay to detect the protein tag. Such methods are well known to one of skill in the art. In a preferred embodiment, the reporter gene is easily assayed and has an activity which is not normally found in the gene of interest. Table 1 below lists various reporter genes and the properties of the products of the reporter genes that can be assayed. In a preferred embodiment, a reporter gene utilized in the reporter constructs is easily assayed and has an activity which is not normally found in the cell or organism of interest.

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TABLE 1: Reporter Genes and the Properties of the Reporter Gene Products

| Reporter Gene | Protein Activity & Measurement |
|---|--|
| CAT (chloramphenicol acetyltransferase) | Transfers radioactive acetyl groups to chloramphenicol or detection by thin layer chromatography and autoradiography |
| GAL (beta-galactosidase) | Hydrolyzes colorless galactosides to yield colored products. |
| GUS (beta-glucuronidase) | Hydrolyzes colorless glucuronides to yield colored products. |
| LUC (luciferase) | Oxidizes luciferin, emitting photons |
| GFP (green fluorescent protein) | Fluorescent protein without substrate |
| SEAP (secreted alkaline phosphatase) | Luminescence reaction with suitable substrates or with substrates that generate chromophores |
| HRP (horseradish peroxidase) | In the presence of hydrogen oxide, oxidation of 3,3',5,5'-tetramethylbenzidine to form a colored complex |
| AP (alkaline phosphatase) | Luminescence reaction with suitable substrates or with substrates that generate chromophores |

| | |
|---|--|
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| AP (alkaline phosphatase) | Luminescence reaction with suitable substrates or with substrates that generate chromophores |

Described hereinbelow in further detailed are specific reporter genes and characteristics of those reporter genes.

5.1.1.1. Luciferase

15 Luciferases are enzymes that emit light in the presence of oxygen and a substrate (luciferin) and which have been used for real-time, low-light imaging of gene expression in cell cultures, individual cells, whole organisms, and transgenic organisms (reviewed by Greer & Szalay, 2002, Luminescence 17(1):43-74).

As used herein, the term “luciferase” is intended to embrace all luciferases, or recombinant enzymes derived from luciferases which have luciferase activity. The luciferase genes from fireflies have been well characterized, for example, from the *Photinus* and *Luciola* species (see, e.g., International Patent Publication No. WO 95/25798 for 5 *Photinus pyralis*, European Patent Application No. EP 0 524 448 for *Luciola cruciata* and *Luciola lateralis*, and Devine et al., 1993, *Biochim. Biophys. Acta* 1173(2):121-132 for *Luciola mingrelica*). Other eucaryotic luciferase genes include, but are not limited to, the click beetle (*Photinus plagiophthalmus*, see, e.g., Wood et al., 1989, *Science* 244:700-702), the sea panzy (*Renilla reniformis*, see, e.g., Lorenz et al., 1991, *Proc Natl Acad Sci U 10 S A* 88(10):4438-4442), and the glow worm (*Lampyris noctiluca*, see e.g., Sula-Newby et al., 1996, *Biochem J.* 313:761-767). The click beetle is unusual in that different members 15 of the species emit bioluminescence of different colors, which emit light at 546 nm (green), 560 nm (yellow-green), 578 nm (yellow) and 593 nm (orange) (see, e.g., U.S. Patent Nos. 6,475,719; 6,342,379; and 6,217,847, the disclosures of which are incorporated by reference in their entireties). Bacterial luciferin-luciferase systems include, but are not limited to, the bacterial lux genes of terrestrial *Photorhabdus luminescens* (see, e.g., Manukhov et al., 2000, *Genetika* 36(3):322-30) and marine bacteria *Vibrio fischeri* and *Vibrio harveyi* (see, e.g., Miyamoto et al., 1988, *J Biol Chem.* 263(26):13393-9, and Cohn et al., 1983, *Proc Natl Acad Sci USA.*, 80(1):120-3, respectively). The luciferases encompassed by the present 20 invention also includes the mutant luciferases described in U.S. Patent No. 6,265,177 to Squirrell et al., which is hereby incorporated by reference in its entirety.

In a specific embodiment, the luciferase is a firefly luciferase, a renilla luciferase, or a click beetle luciferase, as described in any one of the references listed *supra*, the disclosures of which are incorporated by reference in their entireties.

25 5.1.1.2. Green Fluorescent Protein

Green fluorescent protein (“GFP”) is a 238 amino acid protein with amino acid residues 65 to 67 involved in the formation of the chromophore which does not require additional substrates or cofactors to fluoresce (see, e.g., Prasher et al., 1992, *Gene* 111:229-233; Yang et al., 1996, *Nature Biotechnol.* 14:1252-1256; and Cody et al., 1993, *Biochemistry* 32:1212-1218).

As used herein, the term “green fluorescent protein” or “GFP” is intended to embrace all GFPs (including the various forms of GFPs which exhibit colors other than green), or recombinant enzymes derived from GFPs which have GFP activity. In a preferred embodiment, GFP includes green fluorescent protein, yellow fluorescent protein,

red fluorescent protein, cyan fluorescent protein, and blue fluorescent protein. The native gene for GFP was cloned from the bioluminescent jellyfish *Aequorea victoria* (see, e.g., Morin et al., 1972, J. Cell Physiol. 77:313-318). Wild type GFP has a major excitation peak at 395 nm and a minor excitation peak at 470 nm. The absorption peak at 470 nm allows 5 the monitoring of GFP levels using standard fluorescein isothiocyanate (FITC) filter sets. Mutants of the GFP gene have been found useful to enhance expression and to modify excitation and fluorescence. For example, mutant GFPs with alanine, glycine, isoleucine, or threonine substituted for serine at position 65 result in mutant GFPs with shifts in excitation maxima and greater fluorescence than wild type protein when excited at 488 nm (see, e.g., 10 Heim et al., 1995, Nature 373:663-664; U.S. Patent No. 5,625,048; Delagrange et al., 1995, Biotechnology 13:151-154; Cormack et al., 1996, Gene 173:33-38; and Cramer et al., 1996, Nature Biotechnol. 14:315-319). The ability to excite GFP at 488 nm permits the use of GFP with standard fluorescence activated cell sorting ("FACS") equipment. In another embodiment, GFPs are isolated from organisms other than the jellyfish, such as, but not 15 limited to, the sea pansy, *Renilla reniformis*.

Techniques for labeling cells with GFP in general are described in U.S. Patent Nos. 5,491,084 and 5,804,387, which are incorporated by reference in their entireties; Chalfie et al., 1994, Science 263:802-805; Heim et al., 1994, Proc. Natl. Acad. Sci. USA 91:12501-12504; Morise et al., 1974, Biochemistry 13:2656-2662; Ward et al., 20 1980, Photochem. Photobiol. 31:611-615; Rizzuto et al., 1995, Curr. Biology 5:635-642; and Kaether & Gerdes, 1995, FEBS Lett 369:267-271. The expression of GFPs in *E. coli* and *C. elegans* are described in U.S. Patent No. 6,251,384 to Tan et al., which is incorporated by reference in its entirety. The expression of GFP in plant cells is discussed in Hu & Cheng, 1995, FEBS Lett 369:331-33, and GFP expression in *Drosophila* is 25 described in Davis et al., 1995, Dev. Biology 170:726-729.

5.1.1.3. Beta Galactosidase

Beta galactosidase ("beta-gal") is an enzyme that catalyzes the hydrolysis of beta-galactosides, including lactose, and the galactoside analogs o-nitrophenyl-beta-D-galactopyranoside ("ONPG") and chlorophenol red-beta-D-galactopyranoside ("CPRG") 30 (see, e.g., Nielsen et al., 1983 Proc Natl Acad Sci USA 80(17):5198-5202; Eustice et al., 1991, Biotechniques 11:739-742; and Henderson et al., 1986, Clin. Chem. 32:1637-1641). The beta-gal gene functions well as a reporter gene because the protein product is extremely stable, resistant to proteolytic degradation in cellular lysates, and easily assayed. When

ONPG is used as the substrate, beta-gal activity can be quantitated with a spectrophotometer or microplate reader.

As used herein, the term "beta galactosidase" or "beta-gal" is intended to embrace all beta-gals, including *lacZ* gene products, or recombinant enzymes derived from 5 beta-gals which have beta-gal activity. The beta-gal gene functions well as a reporter gene because the protein product is extremely stable, resistant to proteolytic degradation in cellular lysates, and easily assayed. In an embodiment where ONPG is the substrate, beta-gal activity can be quantitated with a spectrophotometer or microplate reader to determine the amount of ONPG converted at 420 nm. In an embodiment when CPRG is the substrate, 10 beta-gal activity can be quantitated with a spectrophotometer or microplate reader to determine the amount of CPRG converted at 570 to 595 nm. In yet another embodiment, the beta-gal activity can be visually ascertained by plating bacterial cells transformed with a beta-gal construct onto plates containing Xgal and IPTG. Bacterial colonies that are dark blue indicate the presence of high beta-gal activity and colonies that are varying shades of 15 blue indicate varying levels of beta-gal activity.

5.1.1.4. Beta-Glucuronidase

Beta-glucuronidase ("GUS") catalyzes the hydrolysis of a very wide variety of beta-glucuronides, and, with much lower efficiency, hydrolyzes some beta-galacturonides. GUS is very stable, will tolerate many detergents and widely varying ionic 20 conditions, has no cofactors, nor any ionic requirements, can be assayed at any physiological pH, with an optimum between 5.0 and 7.8, and is reasonably resistant to thermal inactivation (see, e.g., U.S. Patent No. 5,268,463, which is incorporated by reference in its entirety).

In one embodiment, the GUS is derived from the *Escherichia coli* beta-glucuronidase gene. In alternate embodiments of the invention, the beta-glucuronidase 25 encoding nucleic acid is homologous to the *E. coli* beta-glucuronidase gene and/or may be derived from another organism or species.

GUS activity can be assayed either by fluorescence or spectrometry, or any other method described in U.S. Patent No. 5,268,463, the disclosure of which is 30 incorporated by reference in its entirety. For a fluorescent assay, 4-trifluoromethylumbelliferyl beta-D-glucuronide is a very sensitive substrate for GUS. The fluorescence maximum is close to 500 nm--bluish green, where very few plant compounds fluoresce or absorb. 4-trifluoromethylumbelliferyl beta-D-glucuronide also fluoresces much more strongly near neutral pH, allowing continuous assays to be performed more readily

than with MUG. 4-trifluoromethylumbelliferyl beta-D-glucuronide can be used as a fluorescent indicator *in vivo*. The spectrophotometric assay is very straightforward and moderately sensitive (Jefferson et al., 1986, Proc. Natl. Acad. Sci. USA 83:8447-8451). A preferred substrate for spectrophotometric measurement is p-nitrophenyl beta-D-glucuronide, which when cleaved by GUS releases the chromophore p-nitrophenol. At a pH greater than its pK_a (around 7.15) the ionized chromophore absorbs light at 400-420 nm, giving a yellow color.

5.1.1.5. Beta-Lactamases

Beta-lactamases are nearly optimal enzymes in respect to their almost diffusion-controlled catalysis of beta-lactam hydrolysis, making them suited to the task of an intracellular reporter enzyme (see, e.g., Christensen et al., 1990, Biochem. J. 266: 853-861). They cleave the beta-lactam ring of beta-lactam antibiotics, such as penicillins and cephalosporins, generating new charged moieties in the process (see, e.g., O'Callaghan et al., 1968, Antimicrob. Agents. Chemother. 8: 57-63 and Stratton, 1988, J. Antimicrob. Chemother. 22, Suppl. A: 23-35). A large number of beta-lactamases have been isolated and characterized, all of which would be suitable for use in accordance with the present invention (see, e.g., Richmond & Sykes, 1978, Adv.Microb.Physiol. 9:31-88 and Ambler, 1980, Phil. Trans. R. Soc. Lond. [Ser.B.] 289: 321-331, the disclosures of which are incorporated by reference in their entireties).

The coding region of an exemplary beta-lactamase employed has been described in U.S. Patent No. 6,472,205, Kadonaga et al., 1984, J.Biol.Chem. 259: 2149-2154, and Sutcliffe, 1978, Proc. Natl. Acad. Sci. USA 75: 3737-3741, the disclosures of which are incorporated by reference in their entireties. As would be readily apparent to those skilled in the field, this and other comparable sequences for peptides having beta-lactamase activity would be equally suitable for use in accordance with the present invention. The combination of a fluorogenic substrate described in U.S. Patent Nos. 6,472,205, 5,955,604, and 5,741,657, the disclosures of which are incorporated by reference in their entireties, and a suitable beta-lactamase can be employed in a wide variety of different assay systems, such as are described in U.S. Patent No. 4,740,459, which is hereby incorporated by reference in its entirety.

5.1.1.6. Chloramphenicol Acetyltransferase

Chloramphenicol acetyl transferase ("CAT") is commonly used as a reporter gene in mammalian cell systems because mammalian cells do not have detectable levels of

CAT activity. The assay for CAT involves incubating cellular extracts with radiolabeled chloramphenicol and appropriate co-factors, separating the starting materials from the product by, for example, thin layer chromatography ("TLC"), followed by scintillation counting (see, e.g., U.S. Patent No. 5,726,041, which is hereby incorporated by reference in its entirety).

As used herein, the term "chloramphenicol acetyltransferase" or "CAT" is intended to embrace all CATs, or recombinant enzymes derived from CAT which have CAT activity. While it is preferable that a reporter system which does not require cell processing, radioisotopes, and chromatographic separations would be more amenable to high through-put screening, CAT as a reporter gene may be preferable in situations when stability of the reporter gene is important. For example, the CAT reporter protein has an *in vivo* half life of about 50 hours, which is advantageous when an accumulative versus a dynamic change type of result is desired.

5.1.1.7. Secreted Alkaline Phosphatase

The secreted alkaline phosphatase ("SEAP") enzyme is a truncated form of alkaline phosphatase, in which the cleavage of the transmembrane domain of the protein allows it to be secreted from the cells into the surrounding media. In a preferred embodiment, the alkaline phosphatase is isolated from human placenta.

As used herein, the term "secreted alkaline phosphatase" or "SEAP" is intended to embrace all SEAP or recombinant enzymes derived from SEAP which have alkaline phosphatase activity. SEAP activity can be detected by a variety of methods including, but not limited to, measurement of catalysis of a fluorescent substrate, immunoprecipitation, HPLC, and radiometric detection. The luminescent method is preferred due to its increased sensitivity over calorimetric detection methods. The advantages of using SEAP is that a cell lysis step is not required since the SEAP protein is secreted out of the cell, which facilitates the automation of sampling and assay procedures. A cell-based assay using SEAP for use in cell-based assessment of inhibitors of the Hepatitis C virus protease is described in U.S. Patent No. 6,280,940 to Potts et al. which is hereby incorporated by reference in its entirety.

30 5.1.2. Proteins That Regulate the Expression of Reporter Genes

The invention provides a nucleic acid sequence comprising a nucleotide sequence encoding a regulatory protein or a component or subunit thereof, which regulatory protein binds to a regulatory element operably linked to a reporter gene and regulates the

expression of the reporter gene. The expression of the full-length regulatory protein or component or subunit thereof is suppressed or inhibited in the absence of a compound that suppresses premature translation termination and/or nonsense-mediated mRNA decay because of the presence of a premature stop codon or nonsense mutation within the open reading frame of the nucleotide sequence encoding the regulatory protein. The expression of the full-length regulatory protein or component or subunit thereof is, thus, contingent on the suppression of the premature stop codon or nonsense mutation by a compound. As the expression of the reporter gene is regulated by a regulatory element responsive to the full-length regulatory protein, reporter gene expression should only be detected in the presence 5 of a compound that suppresses the premature stop codon or nonsense mutation.

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The location of the premature stop codon or nonsense mutation is N-terminal to the native stop codon of the regulatory protein or component or subunit thereof. In a specific embodiment, the premature stop codon or nonsense mutation is at least 15 nucleotides, preferably at least 25 nucleotides, at least 50 nucleotides, at least 75 nucleotides 15 or at least 100 nucleotides from the start codon in the open reading frame of the nucleotide sequence encoding the regulatory protein or a component or subunit thereof. In another embodiment, the premature stop codon or nonsense mutation is at least 15 nucleotides, preferably at least 25 nucleotides, at least 50 nucleotides, at least 75 nucleotides or at least 100 nucleotides from the native stop codon in the open reading frame of the nucleotide 20 sequence encoding the regulatory protein or a component or subunit thereof. In another embodiment, the premature stop codon in the open reading frame of the nucleotide sequences. In another embodiment, the premature stop codon in the open reading frame of the nucleotide sequence encoding the regulatory protein or a component or subunit thereof is in the context of UGAA, UGAC, UGAG, UGAU, UAGA, UAGC, UAGG, UAGU, 25 UAAA, UAAC, UAAG or UAAU. In yet another embodiment, the nucleotide sequence encoding the regulatory protein or a component or subunit thereof, contains or is engineered to contain two, three, four or more stop codons. In another embodiment, the premature stop codon in the open reading frame of the nucleotide sequences encoding the regulatory protein or a component or subunit thereof is UAG or UGA.

30 In one embodiment, the invention provides a nucleic acid sequence comprising a nucleotide sequence encoding a regulatory protein with a premature stop codon or nonsense mutation. In accordance with this embodiment, the nucleic acid sequence can encode a naturally-occurring gene with a premature stop codon or nonsense mutation or the nucleic acid sequence can be engineered to contain a premature stop codon

or nonsense mutation using techniques well-known in the art. In this case, the expression of the full-length regulatory protein regulates the expression of the reporter gene which is detected by techniques well-known in the art or described herein.

In another embodiment, the invention provides a first nucleic acid sequence and a second nucleic acid sequence, wherein the first nucleic acid sequence comprises a nucleotide sequence encoding a first fusion protein comprising (or alternatively, consisting of) a DNA binding domain and a first protein, and the second nucleic acid sequence comprises a nucleotide sequence encoding a second fusion protein comprising (or alternatively, consisting of) an activation domain and a second protein. In accordance with 5 this embodiment, the nucleotide sequence encoding the first or second fusion protein may contain or be engineered to contain a premature stop codon or nonsense mutation. The first fusion protein and second fusion protein interact and produce a regulatory protein when the premature stop codon or nonsense mutation is suppressed by a compound. Thus, the production of a functional regulatory protein is dependent on suppression of a premature 10 stop codon or nonsense mutation. In this case, the production of the functional regulatory protein regulates the expression of the reporter gene which is detected by techniques well-known in the art or described herein.

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In one embodiment of the invention, the protein that regulates expression of a gene contains domains which are associated with various activities related to transcriptional regulation, including, but not limited to, binding and activation. In one embodiment, a binding domain of a regulatory protein is one that recognizes and specifically associates with a sequence of at least two nucleotides of a nucleic acid. Nucleic acids that can be recognized by a binding domain of a protein, include, but are not limited to, DNA and RNA both single and multiple stranded. In a more specific embodiment, a 20 binding domain can adopt one of a number of conformations or motifs, known in the art, including but not limited to, zinc finger, leucine zipper, helix turn helix and helix loop helix. In a more preferred embodiment, the binding domain protein is one that specifically recognizes a region of a nucleic acid. Such recognition can occur through a number of interactions, including, but not limited to, covalent, hydrophobic and van der Waals. In 25 another embodiment, an activation domain is one that modulates, regulates, enhances, suppresses or controls the expression of a gene. In such an embodiment, the activation domain can modulate, regulate, enhance, suppress or control the expression of a gene by interacting, either directly or indirectly, with other compounds or proteins that are required 30 or involved in gene expression. In one embodiment, such domains can be expressed as

proteins that are fused with other proteins suitable for the described assays. For example, in one embodiment, the activation domain of a regulatory protein is expressed as part of a protein or polypeptide encoded by a nucleic acid, and the binding domain of a regulatory protein is expressed as a part of a protein or polypeptide encoded by another nucleic acid.

5 In a more specific embodiment, such binding and regulatory domains are expressed as fusion proteins with other proteins with properties that are suitable to the assay. In an example of an embodiment suitable to the described assays, the binding and regulatory domains are expressed on separate fusion proteins with proteins that interact with each other. For example, the binding domain can be expressed as a chimeric protein that is fused
10 to another protein known to associate with another protein that is expressed from a separate nucleic acid and fused to the activation domain. In such an embodiment, a regulatory complex is formed by the association between the binding domain and the activation domain expressed as parts of the described fusion proteins. Interaction between the two domains can be mediated or initiated by a number of means, preferably through inter or
15 intra molecular associations between the parts of the described fusion proteins that are known to interact with one another. Examples of proteins or complexes that contain domains that bind to nucleic acids in addition to possessing regulatory functions include, but are not limited to, GAL4, glucocorticoid and estrogen receptors (GR and ER), Xfin protein, GCN4, and the transcription factor Max in complex with oncogene Myc.

20 The invention relates to the identification of compounds that modulate premature translation termination or nonsense-mediated mRNA decay, using, in some instances, a reporter based assay. The invention provides for the identification of compounds that modulated premature translation termination via a nonsense stop codon in a nucleic acids. Such nucleic acids include, but are not limited to, DNA and RNA. In a more
25 certain embodiment, the nucleic acid is RNA. In another embodiment, the nucleic acid is single stranded. In other embodiments, the nucleic acids are single stranded. In yet other embodiments, the nucleic acids are more than single stranded, e.g., double, triple or quadruple stranded.

5.1.3. Stop Codons

30 The present invention provides for methods for screening and identifying compounds that modulate premature translation termination and/or nonsense-mediated mRNA decay. A reporter gene may be engineered to contain a premature stop codon or may naturally contain a premature stop codon. Alternatively, a protein, polypeptide or peptide that regulates (directly or indirectly) the expression of a reporter gene may be

engineered to contain or may naturally contain a premature stop codon. The premature stop codon may any one of the stop codons known in the art including UAG, UAA and UGA.

The stop codons are UAG, UAA, and UGA, *i.e.*, signals to the ribosome to terminate protein synthesis, presumably through protein release factors. Even though the
5 use of these stop codons is widespread, they are not universal. For example, UGA specifies tryptophan in the mitochondria of mammals, yeast, *Neurospora crassa*, *Drosophila*, protozoa, and plants (see, *e.g.*, Breitenberger & RajBhandary, 1985, Trends Biochem Sci 10:481). Other examples include the use of UGA for tryptophan in *Mycoplasma* and, in ciliated protozoa, the use of UAA and UAG for glutamine (see, *e.g.*, Jukes et al., 1987, Cold
10 Spring Harb Symp Quant Biol. 52:769-776), the use of UGA for cysteine in the ciliate *Euplotes aediculatus* (see, *e.g.*, Kervestin et al., 2001, EMBO Rep 2001 Aug;2(8):680-684), the use of UGA for tryptophan in *Blepharisma americanum* and the use of UAR for glutamine in *Tetrahymena*, and three spirotrichs, *Stylonychia lemnae*, *S. mytilus*, and *Oxytricha trifallax* (see, *e.g.*, Lozupone et al., 2001, Curr Biol 11(2):65-74). It has been
15 proposed that the ancestral mitochondrion was bearing the universal genetic code and subsequently reassigned the UGA codon to tryptophan independently, at least in the lineage of ciliates, kinetoplastids, rhodophytes, prymnesiophytes, and fungi (see, *e.g.*, Inagaki et al., 1998, J Mol Evol 47(4):378-384).

The readthrough of stop codons also occurs in positive-sense ssRNA viruses
20 by a variety of naturally occurring suppressor tRNAs. Such naturally-occurring suppressor tRNAs include, but are not limited to, cytoplasmic tRNATyr, which reads through the UAG stop codon; cytoplasmic tRNAsGln, which read through UAG and UAA; cytoplasmic tRNAsLeu, which read through UAG; chloroplast and cytoplasmic tRNAsTrp, which read through UGA; chloroplast and cytoplasmic tRNAsCys, which read through UGA;
25 cytoplasmic tRNAsArg, which read through UGA (see, *e.g.*, Beier & Grimm, 2001, Nucl Acids Res 29(23):4767-4782 for a review); and the use of selenocysteine to suppress UGA in *E. coli* (see, *e.g.*, Baron & Böck, 1995, The selenocysteine inserting tRNA species: structure and function. In Söll,D. and RajBhandary,U.L. (eds), tRNA: Structure, Biosynthesis and Function, ASM Press, Washington, DC, pp. 529 544). The mechanism is
30 thought to involve unconventional base interactions and/or codon context effects.

As described above, the stop codons are not necessarily universal, with consideration variation amongst organelles (*e.g.*, mitochondria and chloroplasts), viruses (*e.g.*, single strand viruses), and protozoa (*e.g.*, ciliated protozoa) as to whether the codons UAG, UAA, and UGA signal translation termination or encode amino acids. Even though a

single release factor most probably recognizes all of the stop codons in eucaryotes, it appears that all of the stop codons are not suppressed in a similar manner. For example, in the yeast *Saccharomyces pombe*, nonsense suppression has to be strictly codon specific (see, e.g., Hottinger et al., 1984, EMBO J 3:423-428). In another example, significant 5 differences were found in the degree of suppression amongst three UAG codons and two UAA codons in different mRNA contexts in *Escherichia coli* and in human 293 cells, although data suggested that the context effects of nonsense suppression operated differently in *E. coli* and human cells (see, e.g., Martin et al., 1989, Mol Gen Genet 217(2 10 3):411-8). Since unconventional base interactions and/or codon context effects have been implicated in nonsense suppression, it is conceivable that compounds involved in nonsense suppression of one stop codon may not necessarily be involved in nonsense suppression of another stop codon. In other words, compounds involved in suppressing UAG codons may not necessarily be involved in suppressing UGA codons.

In a specific embodiment, a reporter gene or a gene encoding a protein, 15 polypeptide or peptide that regulates the expression of a reporter gene contains or is engineered to contain the premature stop codon UAG. In another embodiment, a reporter gene or a gene encoding a protein, polypeptide or peptide that regulates the expression of a reporter gene contains or is engineered to contain the premature stop codon UGA. In yet another embodiment, a reporter gene or a gene encoding a protein, polypeptide or peptide 20 that regulates the expression of a reporter gene contains or is engineered to contain a premature stop codon in the context of UGAA, UGAC, UGAG, UGAU, UAGA, UAGC, UAGG, UAGU, UAAA, UAAC, UAAG or UAAU.

In a particular embodiment, a reporter gene or a gene encoding a protein, 25 polypeptide or peptide that regulates the expression of a reporter gene contains or is engineered to contain two, three, four or more stop codons. In accordance with this embodiment, the stop codons are preferably at least 10 nucleotides, at least 15 nucleotides, at least 20 nucleotides, at least 25 nucleotides, at least 30 nucleotides, at least 35 nucleotides, at least 40 nucleotides, at least 45 nucleotides, at least 50 nucleotides, at least 75 nucleotides or at least 100 nucleotides apart from each other. Further, in accordance 30 with this embodiment, at least one of the stop codons is preferably UAG or UGA.

In a specific embodiment, a reporter gene or a gene encoding a protein, polypeptide or peptide that regulates the expression of a reporter gene contains or is engineered to contain a premature stop codon at least 15 nucleotides, preferably at least 20 nucleotides, at least 25 nucleotides, at least 30 nucleotides, at least 35 nucleotides, at least

40 nucleotides, at least 45 nucleotides, at least 50 nucleotides or at least 75 nucleotides from the start codon in the coding sequence. In another embodiment, a reporter gene or a gene encoding a protein, polypeptide or peptide that regulates the expression of a reporter gene contains or is engineered to contain a premature stop codon at least 15 nucleotides,
5 preferably at least 25 nucleotides, at least 50 nucleotides, at least 75 nucleotides, at least 100 nucleotides, at least 125 nucleotides, at least 150, at least 175 nucleotides or at least 200 nucleotides from the native stop codon in the coding sequence of the full-length reporter gene product or protein, polypeptide or peptide. In another embodiment, a reporter gene or a gene encoding a protein, polypeptide or peptide that regulates the expression of a reporter
10 gene contains or is engineered to contain a premature stop codon at least 15 nucleotides (preferably at least 20 nucleotides, at least 25 nucleotides, at least 30 nucleotides, at least 35 nucleotides, at least 40 nucleotides, at least 45 nucleotides, at least 50 nucleotides or at least 75 nucleotides) from the start codon in the coding sequence and at least 15 nucleotides (preferably at least 25 nucleotides, at least 50 nucleotides, at least 75 nucleotides, at least
15 100 nucleotides, at least 125 nucleotides, at least 150, at least 175 nucleotides or at least 200 nucleotides) from the native stop codon in the coding sequence of the full-length reporter gene product or protein, polypeptide or peptide. In accordance with these embodiments, the premature stop codon is preferably UAG or UGA.

The premature translation stop codon can be produced by *in vitro* mutagenesis
20 techniques such as, but not limited to, polymerase chain reaction ("PCR"), linker insertion, oligonucleotide-mediated mutagenesis, and random chemical mutagenesis.

5.1.4. Vectors

The nucleotide sequence encoding for a protein, polypeptide or peptide (*e.g.*, a reporter gene, or a protein, polypeptide or peptide that regulates the expression of a
25 reporter gene) can be inserted into an appropriate expression vector, *i.e.*, a vector which contains the necessary elements for the transcription and translation of the inserted protein-coding sequence. The necessary transcriptional and translational elements can also be supplied by the protein, polypeptide or peptide. The regulatory regions and enhancer elements can be of a variety of origins, both natural and synthetic. In a specific
30 embodiment, a reporter gene is operably linked to regulatory element that is responsive to a regulatory protein whose expression is dependent upon the suppression of a premature stop codon.

A variety of host-vector systems may be utilized to express a protein, polypeptide or peptide. These include, but are not limited to, mammalian cell systems

infected with virus (*e.g.*, vaccinia virus, adenovirus, *etc.*); insect cell systems infected with virus (*e.g.*, baculovirus); microorganisms such as yeast containing yeast vectors, or bacteria transformed with bacteriophage, DNA, plasmid DNA, or cosmid DNA; and stable cell lines generated by transformation using a selectable marker. The expression elements of vectors
5 vary in their strengths and specificities. Depending on the host-vector system utilized, any one of a number of suitable transcription and translation elements may be used.

Any of the methods previously described for the insertion of DNA fragments into a vector may be used to construct expression vectors containing a chimeric nucleic acid consisting of appropriate transcriptional/translational control signals and the protein coding sequences. These methods may include *in vitro* recombinant DNA and synthetic techniques
10 and *in vivo* recombinants (genetic recombination). Expression of a first nucleic acid sequence encoding a protein, polypeptide or peptide, such as a reporter gene, may be regulated by a second nucleic acid sequence so that the first nucleic acid sequence is expressed in a host transformed with the second nucleic acid sequence. For example,
15 expression of a nucleic acid sequence encoding a protein, polypeptide or peptide, such as a reporter gene, may be controlled by any promoter/enhancer element known in the art, such as a constitutive promoter, a tissue-specific promoter, or an inducible promoter. Specific examples of promoters which may be used to control gene expression include, but are not limited to, the SV40 early promoter region (Bernoist & Chambon, 1981, *Nature* 290:304-
20 310), the promoter contained in the 3' long terminal repeat of Rous sarcoma virus (Yamamoto et al., 1980, *Cell* 22:787-797), the herpes thymidine kinase promoter (Wagner et al., 1981, *Proc. Natl. Acad. Sci. U.S.A.* 78:1441-1445), the regulatory sequences of the metallothionein gene (Brinster et al., 1982, *Nature* 296:39-42); prokaryotic expression vectors such as the β -lactamase promoter (Villa-Kamaroff et al., 1978, *Proc. Natl. Acad.
25 Sci. U.S.A.* 75:3727-3731), or the *tac* promoter (DeBoer et al., 1983, *Proc. Natl. Acad. Sci. U.S.A.* 80:21-25); see also "Useful proteins from recombinant bacteria" in *Scientific American*, 1980, 242:74-94; plant expression vectors comprising the nopaline synthetase promoter region (Herrera-Estrella et al., *Nature* 303:209-213) or the cauliflower mosaic virus 35S RNA promoter (Gardner, et al., 1981, *Nucl. Acids Res.* 9:2871), and the promoter
30 of the photosynthetic enzyme ribulose biphosphate carboxylase (Herrera-Estrella et al., 1984, *Nature* 310:115-120); promoter elements from yeast or other fungi such as the Gal 4 promoter, the ADC (alcohol dehydrogenase) promoter, PGK (phosphoglycerol kinase) promoter, alkaline phosphatase promoter, and the following animal transcriptional control regions, which exhibit tissue specificity and have been utilized in transgenic animals:

elastase I gene control region which is active in pancreatic acinar cells (Swift et al., 1984, Cell 38:639-646; Ornitz et al., 1986, Cold Spring Harbor Symp. Quant. Biol. 50:399-409; MacDonald, 1987, Hepatology 7:425-515); insulin gene control region which is active in pancreatic beta cells (Hanahan, 1985, Nature 315:115-122), immunoglobulin gene control
5 region which is active in lymphoid cells (Grosschedl et al., 1984, Cell 38:647-658; Adames et al., 1985, Nature 318:533-538; Alexander et al., 1987, Mol. Cell. Biol. 7:1436-1444), mouse mammary tumor virus control region which is active in testicular, breast, lymphoid and mast cells (Leder et al., 1986, Cell 45:485-495), albumin gene control region which is active in liver (Pinkert et al., 1987, Genes and Devel. 1:268-276), alpha-fetoprotein gene
10 control region which is active in liver (Krumlauf et al., 1985, Mol. Cell. Biol. 5:1639-1648; Hammer et al., 1987, Science 235:53-58; alpha 1-antitrypsin gene control region which is active in the liver (Kelsey et al., 1987, Genes and Devel. 1:161-171), beta-globin gene control region which is active in myeloid cells (Mogram et al., 1985, Nature 315:338-340; Kollias et al., 1986, Cell 46:89-94; myelin basic protein gene control region which is active
15 in oligodendrocyte cells in the brain (Readhead et al., 1987, Cell 48:703-712); myosin light chain-2 gene control region which is active in skeletal muscle (Sani, 1985, Nature 314:283-286), and gonadotropin releasing hormone gene control region which is active in the hypothalamus (Mason et al., 1986, Science 234:1372-1378).

In a specific embodiment, a vector is used that comprises a promoter
20 operably linked to a reporter gene, one or more origins of replication, and, optionally, one or more selectable markers (e.g., an antibiotic resistance gene). In a preferred embodiment, the vectors are CMV vectors, T7 vectors, lac vectors, pCEP4 vectors, 5.0/F vectors, or vectors with a tetracycline-regulated promoter (e.g., pcDNATM5/FRT/TO from Invitrogen). Some vectors may be obtained commercially. Non-limiting examples of useful vectors are
25 described in Appendix 5 of Current Protocols in Molecular Biology, 1988, ed. Ausubel et al., Greene Publish. Assoc. & Wiley Interscience, which is incorporated herein by reference; and the catalogs of commercial suppliers such as Clontech Laboratories, Stratagene Inc., and Invitrogen, Inc.

Expression vectors containing a construct of the present invention can be
30 identified by the following general approaches: (a) nucleic acid hybridization, (b) presence or absence of "marker" nucleic acid functions, (c) expression of inserted sequences, and (d) sequencing. In the first approach, the presence of a particular nucleic acid sequence inserted in an expression vector can be detected by nucleic acid hybridization using probes comprising sequences that are homologous to the inserted nucleic acid sequence. In the

second approach, the recombinant vector/host system can be identified and selected based upon the presence or absence of certain "marker" nucleic acid functions (*e.g.*, thymidine kinase activity, resistance to antibiotics, transformation phenotype, occlusion body formation in baculovirus, *etc.*) caused by the insertion of the nucleic acid sequence of interest in the vector. For example, if the nucleic acid sequence of interest is inserted within the marker nucleic acid sequence of the vector, recombinants containing the insert can be identified by the absence of the marker nucleic acid function. In the third approach, recombinant expression vectors can be identified by assaying the product expressed by the recombinant. Such assays can be based, for example, on the physical or functional properties of the particular nucleic acid sequence.

In a preferred embodiment, nucleic acid sequences encoding proteins, polypeptides or peptides are cloned into stable cell line expression vectors. In a preferred embodiment, the stable cell line expression vector contains a site specific genomic integration site. In another preferred embodiment, the reporter gene construct is cloned into an episomal mammalian expression vector.

5.1.5. Transfection

Once a vector encoding the appropriate gene has been synthesized, a host cell is transformed or transfected with the vector of interest. The use of stable transformants is preferred. In a preferred embodiment, the host cell is a mammalian cell. In a more preferred embodiment, the host cell is a human cell. In another embodiment, the host cells are primary cells isolated from a tissue or other biological sample of interest. Host cells that can be used in the methods of the present invention include, but are not limited to, hybridomas, pre-B cells, 293 cells, 293T cells, HeLa cells, HepG2 cells, K562 cells, 3T3 cells. In another preferred embodiment, the host cells are derived from tissue specific to the nucleic acid sequence encoding a protein, polypeptide or peptide. In another preferred embodiment, the host cells are immortalized cell lines derived from a source, *e.g.*, a tissue. Other host cells that can be used in the present invention include, but are not limited to, bacterial cells, yeast cells, virally-infected cells, or plant cells.

Preferred mammalian host cells include but are not limited to those derived from humans, monkeys and rodents, (see, for example, Kriegler M. in "Gene Transfer and Expression: A Laboratory Manual", New York, Freeman & Co. 1990), such as monkey kidney cell line transformed by SV40 (COS-7, ATCC Accession No. CRL 1651); human embryonic kidney cell lines (293, 293-EBNA, or 293 cells subcloned for growth in suspension culture, Graham et al., *J. Gen. Virol.*, 36:59, 1977; baby hamster kidney cells

(BHK, ATCC Accession No. CCL 10); chinese hamster ovary-cells-DHFR (CHO, Urlaub and Chasin. Proc. Natl. Acad. Sci. 77; 4216, 1980); mouse sertoli cells (Mather, Biol. Reprod. 23:243-251, 1980); mouse fibroblast cells (NIH-3T3), monkey kidney cells (CVI ATCC Accession No. CCL 70); african green monkey kidney cells (VERO-76, ATCC Accession No. CRL-1587); human cervical carcinoma cells (HELA, ATCC Accession No. CCL 2); canine kidney cells (MDCK, ATCC Accession No. CCL 34); buffalo rat liver cells (BRL 3A, ATCC Accession No. CRL 1442); human lung cells (W138, ATCC Accession No. CCL 75); human liver cells (Hep G2, HB 8065); and mouse mammary tumor cells (MMT 060562, ATCC Accession No. CCL51).

Other useful eukaryotic host-vector system may include yeast and insect systems. In yeast, a number of vectors containing constitutive or inducible promoters may be used with *Saccharomyces cerevisiae* (baker's yeast), *Schizosaccharomyces pombe* (fission yeast), *Pichia pastoris*, and *Hansenula polymorpha* (methylo-tropic yeasts). For a review see, Current Protocols in Molecular Biology, Vol. 2, 1988, Ed. Ausubel et al., Greene Publish. Assoc. & Wiley Interscience, Ch. 13; Grant et al., 1987, Expression and Secretion Vectors for Yeast, in Methods in Enzymology, Eds. Wu & Grossman, 1987, Acad. Press, N.Y., Vol. 153, pp. 516-544; Glover, 1986, DNA Cloning, Vol. II, IRL Press, Wash., D.C., Ch. 3; and Bitter, 1987, Heterologous Gene Expression in Yeast, Methods in Enzymology, Eds. Berger & Kimmel, Acad. Press, N.Y., Vol. 152, pp. 673-684; and The Molecular Biology of the Yeast *Saccharomyces*, 1982, Eds. Strathern et al., Cold Spring Harbor Press, Vols. I and II.

Standard methods of introducing a nucleic acid sequence of interest into host cells can be used. Transformation may be by any known method for introducing polynucleotides into a host cell, including, for example packaging the polynucleotide in a virus and transducing a host cell with the virus, and by direct uptake of the polynucleotide. The transformation procedure used depends upon the host to be transformed. Mammalian transformations (*i.e.*, transfections) by direct uptake may be conducted using the calcium phosphate precipitation method of Graham & Van der Eb, 1978, Virol. 52:546, or the various known modifications thereof. Other methods for introducing recombinant polynucleotides into cells, particularly into mammalian cells, include dextran-mediated transfection, calcium phosphate mediated transfection, polybrene mediated transfection, protoplast fusion, electroporation, encapsulation of the polynucleotide(s) in liposomes, and direct microinjection of the polynucleotides into nuclei. Such methods are well-known to one of skill in the art.

In a preferred embodiment, stable cell lines containing the constructs of interest are generated for high throughput screening. Such stable cells lines may be generated by introducing a construct comprising a selectable marker, allowing the cells to grow for 1-2 days in an enriched medium, and then growing the cells on a selective medium. The selectable marker in the recombinant plasmid confers resistance to the selection and allows cells to stably integrate the plasmid into their chromosomes and grow to form foci which in turn can be cloned and expanded into cell lines.

A number of selection systems may be used, including but not limited to the herpes simplex virus thymidine kinase (Wigler, et al., 1977, Cell 11:223), hypoxanthine-guanine phosphoribosyltransferase (Szybalska & Szybalski, 1962, Proc. Natl. Acad. Sci. USA 48:2026), and adenine phosphoribosyltransferase (Lowy, et al., 1980, Cell 22:817) genes can be employed in tk-, hgprt- or aprt- cells, respectively. Also, anti-metabolite resistance can be used as the basis of selection for dhfr, which confers resistance to methotrexate (Wigler, et al., 1980, Natl. Acad. Sci. USA 77:3567; O'Hare, et al., 1981, Proc. Natl. Acad. Sci. USA 78:1527); gpt, which confers resistance to mycophenolic acid (Mulligan & Berg, 1981, Proc. Natl. Acad. Sci. USA 78:2072); neo, which confers resistance to the aminoglycoside G-418 (Colberre-Garapin, et al., 1981, J. Mol. Biol. 150:1); and hygro, which confers resistance to hygromycin (Santerre, et al., 1984, Gene 30:147) genes.

20 5.1.6. Cell-Free Extracts

The invention provides for the translation of a nucleic acid sequence encoding a protein, polypeptide or peptide (with or without a premature translation stop codon) in a cell-free system. Techniques for practicing the specific aspect of this invention will employ, unless otherwise indicated, conventional techniques of molecular biology, microbiology, and recombinant DNA manipulation and production, which are routinely practiced by one of skill in the art. See, e.g., Sambrook, 1989, Molecular Cloning, A Laboratory Manual, Second Edition; DNA Cloning, Volumes I and II (Glover, Ed. 1985); and Transcription and Translation (Hames & Higgins, Eds. 1984).

Any technique well-known to one of skill in the art may be used to generate cell-free extracts for translation. For example, the cell-free extracts can be generated by centrifuging cells and clarifying the supernatant. In one embodiment, the cells are incubated on ice during the preparation of the cell-free extract. In another embodiment, the cells are incubated on ice at least 12 hours, at least 24 hours, at least two days, at least five days, at least one week, at least longer than one week. In a more specific embodiment, the

cells are incubated on ice at least long enough so as to improve the translation activity of the cell extract in comparison to cell extracts that are not incubated on ice. In yet another embodiment, the cells are incubated at a temperature between about 0 °C and 10 °C. In a preferred embodiment, the cells are incubated at about 4 °C.

5 In another preferred embodiment, the cells are centrifuged at a low speed to isolate the cell-free extract for *in vitro* translation reactions. In a preferred embodiment, the cell extract is the supernatant from cells that are centrifuged at about 2 x g to 20,000 x g. In a more preferred embodiment, the cell extract is the supernatant from cells that are centrifuged at about 5 x g to 15,000 x g. In an even more preferred embodiment, the cell extract is the supernatant from cells that are centrifuged at about 10,000 x g. Alternatively, 10 in a preferred embodiment, the cell-free extract is about the S1 to S50 extract. In a more preferred embodiment, the cell extract is about the S5 to S25 extract. In an even more preferred embodiment, the cell extract is about the S10 extract.

15 The cell-free translation extract may be isolated from cells of any species origin. In another embodiment, the cell-free translation extract is isolated from yeast, cultured mouse or rat cells, Chinese hamster ovary (CHO) cells, *Xenopus* oocytes, reticulocytes, wheat germ, or rye embryo (see, e.g., Krieg & Melton, 1984, Nature 308:203 and Dignam *et al.*, 1990 Methods Enzymol. 182:194-203). Alternatively, the cell-free translation extract, e.g., rabbit reticulocyte lysates and wheat germ extract, can be purchased 20 from, e.g., Promega, (Madison, WI). In another embodiment, the cell-free translation extract is prepared as described in International Patent Publication No. WO 01/44516 and U.S. Patent No. 4,668,625 to Roberts, the disclosures of which are incorporated by reference in their entireties. In a preferred embodiment, the cell-free extract is an extract isolated from human cells. In a more preferred embodiment, the human cells are HeLa 25 cells. It is preferred that the endogenous expression of the genes with the premature translation codons is minimal, and preferably absent, in the cells from which the cell-free translation extract is prepared.

Systems for the *in vitro* transcription of RNAs with the gene of interest cloned in an expression vectors using promoters such as, but not limited to, Sp6, T3, or T7 30 promoters (see, e.g., expression vectors from Invitrogen, Carlesbad, CA; Promega, Madison, WI; and Stratagene, La Jolla, CA), and the subsequent transcription of the gene with the appropriate polymerase are well-known to one of skill in the art (see, e.g., Contreras *et al.*, 1982, Nucl. Acids. Res. 10:6353). In another embodiment, the gene encoding the premature stop codon can be PCR-amplified with the appropriate primers,

with the sequence of a promoter, such as but not limited to, Sp6, T3, or T7 promoters, incorporated into the upstream primer, so that the resulting amplified PCR product can be *in vitro* transcribed with the appropriate polymerase.

Alternatively, a coupled transcription-translation system can be used for the
5 expression of a gene encoding a premature stop codon in a cell free extract, such as the TnT® Coupled Transcription/Translation System (Promega, Madison, WI) or the system described in U.S. Patent No. 5,895,753 to Mierendorf *et al.*, which is incorporated by reference in its entirety.

5.2. Compounds

10 Libraries screened using the methods of the present invention can comprise a variety of types of compounds. Examples of libraries that can be screened in accordance with the methods of the invention include, but are not limited to, peptoids; random biooligomers; diversomers such as hydantoins, benzodiazepines and dipeptides; vinylogous polypeptides; nonpeptidal peptidomimetics; oligocarbamates; peptidyl phosphonates; 15 peptide nucleic acid libraries; antibody libraries; carbohydrate libraries; and small molecule libraries (preferably, small organic molecule libraries). In some embodiments, the compounds in the libraries screened are nucleic acid or peptide molecules. In a non-limiting example, peptide molecules can exist in a phage display library. In other embodiments, the types of compounds include, but are not limited to, peptide analogs including peptides comprising non-naturally occurring amino acids, *e.g.*, D-amino acids, phosphorous analogs of amino acids, such as α -amino phosphoric acids and α -amino phosphoric acids, or amino acids having non-peptide linkages, nucleic acid analogs such as phosphorothioates and PNAs, hormones, antigens, synthetic or naturally occurring drugs, opiates, dopamine, serotonin, catecholamines, thrombin, acetylcholine, prostaglandins, organic molecules, 20 pheromones, adenosine, sucrose, glucose, lactose and galactose. Libraries of polypeptides or proteins can also be used in the assays of the invention. In some embodiments, the compounds are nucleic acid or peptide molecules. In a non-limiting example, peptide molecules can exist in a phage display library.

In a preferred embodiment, the combinatorial libraries are small organic
30 molecule libraries, such as, but not limited to, benzodiazepines, isoprenoids, thiazolidinones, metathiazanones, pyrrolidines, morpholino compounds, and benzodiazepines. In another embodiment, the combinatorial libraries comprise peptoids; random bio-oligomers; benzodiazepines; diversomers such as hydantoins, benzodiazepines and dipeptides; vinylogous polypeptides; nonpeptidal peptidomimetics; oligocarbamates;

peptidyl phosphonates; peptide nucleic acid libraries; antibody libraries; or carbohydrate libraries. Combinatorial libraries are themselves commercially available (see, e.g., ComGenex, Princeton, New Jersey; Asinex, Moscow, Ru, Tripos, Inc., St. Louis, Missouri; ChemStar, Ltd, Moscow, Russia; 3D Pharmaceuticals, Exton, Pennsylvania; Martek Biosciences, Columbia, Maryland; etc.).

In a preferred embodiment, the library is preselected so that the compounds of the library are more amenable for cellular uptake. For example, compounds are selected based on specific parameters such as, but not limited to, size, lipophilicity, hydrophilicity, and hydrogen bonding, which enhance the likelihood of compounds getting into the cells.

- 10 5 In another embodiment, the compounds are analyzed by three-dimensional or four-dimensional computer computation programs.

In one embodiment, the combinatorial compound library for the methods of the present invention may be synthesized. There is a great interest in synthetic methods directed toward the creation of large collections of small organic compounds, or libraries, 15 which could be screened for pharmacological, biological or other activity. The synthetic methods applied to create vast combinatorial libraries are performed in solution or in the solid phase, *i.e.*, on a solid support. Solid-phase synthesis makes it easier to conduct multi-step reactions and to drive reactions to completion with high yields because excess reagents can be easily added and washed away after each reaction step. Solid-phase 20 combinatorial synthesis also tends to improve isolation, purification and screening. However, the more traditional solution phase chemistry supports a wider variety of organic reactions than solid-phase chemistry.

Combinatorial compound libraries of the present invention may be synthesized using the apparatus described in U.S. Patent No. 6,190,619 to Kilcoin *et al.*, 25 which is hereby incorporated by reference in its entirety. U.S. Patent No. 6,190,619 discloses a synthesis apparatus capable of holding a plurality of reaction vessels for parallel synthesis of multiple discrete compounds or for combinatorial libraries of compounds.

In one embodiment, the combinatorial compound library can be synthesized in solution. The method disclosed in U.S. Patent No. 6,194,612 to Boger *et al.*, which is 30 hereby incorporated by reference in its entirety, features compounds useful as templates for solution phase synthesis of combinatorial libraries. The template is designed to permit reaction products to be easily purified from unreacted reactants using liquid/liquid or solid/liquid extractions. The compounds produced by combinatorial synthesis using the template will preferably be small organic molecules. Some compounds in the library may

mimic the effects of non-peptides or peptides. In contrast to solid phase synthesis of combinatorial compound libraries, liquid phase synthesis does not require the use of specialized protocols for monitoring the individual steps of a multistep solid phase synthesis (Egner *et al.*, 1995, J. Org. Chem. 60:2652; Anderson *et al.*, 1995, J. Org. Chem. 60:2650; 5 Fitch *et al.*, 1994, J. Org. Chem. 59:7955; Look *et al.*, 1994, J. Org. Chem. 49:7588; Metzger *et al.*, 1993, Angew. Chem., Int. Ed. Engl. 32:894; Youngquist *et al.*, 1994, Rapid Commun. Mass Spect. 8:77; Chu *et al.*, 1995, J. Am. Chem. Soc. 117:5419; Brummel *et al.*, 1994, Science 264:399; Stevanovic *et al.*, 1993, Bioorg. Med. Chem. Lett. 3:431).

Combinatorial compound libraries useful for the methods of the present 10 invention can be synthesized on solid supports. In one embodiment, a split synthesis method, a protocol of separating and mixing solid supports during the synthesis, is used to synthesize a library of compounds on solid supports (see e.g., Lam *et al.*, 1997, Chem. Rev. 97:41-448; Ohlmeyer *et al.*, 1993, Proc. Natl. Acad. Sci. USA 90:10922-10926 and references cited therein). Each solid support in the final library has substantially one type of 15 compound attached to its surface. Other methods for synthesizing combinatorial libraries on solid supports, wherein one product is attached to each support, will be known to those of skill in the art (see, e.g., Nefzi *et al.*, 1997, Chem. Rev. 97:449-472).

As used herein, the term "solid support" is not limited to a specific type of 20 solid support. Rather a large number of supports are available and are known to one skilled in the art. Solid supports include silica gels, resins, derivatized plastic films, glass beads, cotton, plastic beads, polystyrene beads, alumina gels, and polysaccharides. A suitable solid support may be selected on the basis of desired end use and suitability for various synthetic 25 protocols. For example, for peptide synthesis, a solid support can be a resin such as p-methylbenzhydrylamine (pMBHA) resin (Peptides International, Louisville, KY), polystyrenes (e.g., PAM-resin obtained from Bachem Inc., Peninsula Laboratories, etc.), including chloromethylpolystyrene, hydroxymethylpolystyrene and aminomethylpolystyrene, poly (dimethylacrylamide)-grafted styrene co-divinyl-benzene (e.g., POLYHIPE resin, obtained from Aminotech, Canada), polyamide resin (obtained 30 from Peninsula Laboratories), polystyrene resin grafted with polyethylene glycol (e.g., TENTAGEL or ARGOGEL, Bayer, Tubingen, Germany) polydimethylacrylamide resin (obtained from Milligen/Bioscience, California), or Sepharose (Pharmacia, Sweden).

In some embodiments of the present invention, compounds can be attached to solid supports via linkers. Linkers can be integral and part of the solid support, or they may be nonintegral that are either synthesized on the solid support or attached thereto after

synthesis. Linkers are useful not only for providing points of compound attachment to the solid support, but also for allowing different groups of molecules to be cleaved from the solid support under different conditions, depending on the nature of the linker. For example, linkers can be, *inter alia*, electrophilically cleaved, nucleophilically cleaved, 5 photocleavable, enzymatically cleaved, cleaved by metals, cleaved under reductive conditions or cleaved under oxidative conditions. In a preferred embodiment, the compounds are cleaved from the solid support prior to high throughput screening of the compounds.

In certain embodiments of the invention, the compound is a small molecule.

10

5.3. Reporter Gene-Based Screening Assays

Various *in vitro* assays can be used to identify and verify the ability of a compound to modulate premature translation termination and/or nonsense-mediated mRNA decay. Multiple *in vitro* assays can be performed simultaneously or sequentially to assess 15 the affect of a compound on premature translation termination and/or nonsense-mediated mRNA decay. In a preferred embodiment, the *in vitro* assays described herein are performed in a high throughput format (*e.g.*, in microtiter plates).

5.3.1. Cell-Based Assays

After a vector containing the reporter gene construct and/or a vector(s) 20 containing a nucleic acid sequence comprising a regulatory protein, a component or a subunit thereof is transformed or transfected into a host cell and a compound library is synthesized or purchased or both, the cells are used to screen the library to identify compounds that modulate premature translation termination and/or nonsense-mediated mRNA decay. The reporter gene-based assays may be conducted by contacting a 25 compound or a member of a library of compounds with a cell (*e.g.*, a genetically engineered cell) containing a reporter gene construct comprising a reporter gene containing within the open reading frame of the reporter gene a premature stop codon or nonsense mutation; and measuring the expression and/or activity of the reporter gene. The reporter gene cell-based assays may also be conducted by: (a) contacting a compound with a cell containing a first 30 nucleic acid sequence and a second nucleic acid sequence, wherein the first nucleic acid sequence comprises a regulatory element operably linked to a reporter gene and the second nucleic acid sequence comprises a nucleotide sequence with a premature stop codon or nonsense mutation that encodes a regulatory protein that binds to the regulatory element of

the first nucleic acid sequence and regulates the expression of the reporter gene; and (b) measuring the expression and/or activity of the reporter gene.

The reporter gene cell-based assays may also be conducted by: (a) contacting a compound with a cell containing a first nucleic acid sequence, a second nucleic acid sequence and a third nucleic acid sequence, wherein (i) the first nucleic acid sequence comprises a nucleotide sequence encoding a first fusion protein comprising a DNA binding domain and a first protein, the nucleotide sequence of the DNA binding domain or the first protein containing a premature stop codon or nonsense mutation, (ii) the second nucleic acid sequence comprises a nucleotide sequence encoding a second fusion protein comprising an activation domain and a second protein, the second protein interacting with the first protein to produce a regulatory protein, and (iii) the third nucleic acid sequence comprises a regulatory element operably linked to a reporter gene, the expression of the reporter gene being regulated by the binding of the regulatory protein to the regulatory element: and (b) measuring the expression and/or activity of the reporter gene. Further, the reporter gene cell-based assays may also be conducted by: (a) contacting a compound with a cell containing a first nucleic acid sequence, a second nucleic acid sequence and a third nucleic acid sequence, wherein (i) the first nucleic acid sequence comprises a nucleotide sequence encoding a first fusion protein comprising a DNA binding domain and a first protein, (ii) the second nucleic acid sequence comprises a nucleotide sequence encoding a second fusion protein comprising an activation domain and a second protein, the nucleotide sequence of the activation domain or the second protein containing a premature stop codon, and the second protein interacting with the first protein to produce a premature stop codon or nonsense mutation, and (iii) the third nucleic acid sequence comprises a regulatory element operably linked to a reporter gene, the expression of the reporter gene being regulated by the binding of the regulatory protein to the regulatory element: and (b) measuring the expression and/or activity of the reporter gene.

The alteration in reporter gene expression and/or activity in the reporter gene cell-based assays relative to a previously determined reference range, or to the expression or activity of the reporter gene in the absence of the compound or the presence of an appropriate control (*e.g.*, a negative control such as phosphate buffered saline) indicates that a particular compound modulates premature translation termination and/or nonsense-mediated mRNA decay. In particular, a decrease in reporter gene expression or activity relative to a previously determined reference range, or to the expression in the absence of the compound or the presence of an appropriate control (*e.g.*, a negative control) may,

depending upon the parameters of the reporter gene assay, indicate that a particular compound reduces or suppresses premature translation termination and/or nonsense-mediated mRNA decay. In contrast, an increase in reporter gene expression or activity relative to a previously determined reference range, or to the expression in the absence of 5 the compound or the presence of an appropriate control (e.g., a negative control) may, depending upon the parameters of the reporter gene-based assay, indicate that a particular compound enhances premature translation termination and/or nonsense-mediated mRNA decay.

The step of contacting a compound or a member of a library of compounds 10 with cell in the reporter gene-based assays described herein is preferably conducted under physiologic conditions. In specific embodiment, a compound or a member of a library of compounds is added to the cells in the presence of an aqueous solution. In accordance with this embodiment, the aqueous solution may comprise a buffer and a combination of salts, preferably approximating or mimicking physiologic conditions. Alternatively, the aqueous 15 solution may comprise a buffer, a combination of salts, and a detergent or a surfactant. Examples of salts which may be used in the aqueous solution include, but not limited to, KCl, NaCl, and/or MgCl₂. The optimal concentration of each salt used in the aqueous solution is dependent on the cells and compounds used and can be determined using routine experimentation. The step of contacting a compound or a member of a library of 20 compounds with a cell containing a reporter gene construct and in some circumstances, a nucleic acid sequence encoding a regulatory protein, may be performed for at least 0.2 hours, 0.25 hours, 0.5 hours, 1 hour, 2 hours, 3 hours, 4 hours, 5 hours, 6 hours, 8 hours, 10 hours, 12 hours, 18 hours, at least 1 day, at least 2 days or at least 3 days.

The expression of a reporter gene and/or activity of the protein encoded by 25 the reporter gene in the cell-based reporter-gene assays may be detected by any technique well-known to one of skill in the art. The expression of a reporter gene can be readily detected, e.g., by quantifying the protein and/or RNA encoded by said gene. Compounds that modulate premature translation termination and/or nonsense-mediated mRNA decay may be identified by changes in the gene encoding the premature translation stop codon, 30 i.e., there is readthrough of the premature translation stop codon and a longer gene product is detected. If a gene encoding a naturally-occurring premature translation stop codon is used, a longer gene product in the presence of a compound that modulates premature translation termination and/or nonsense-mediated mRNA decay can be detected by any

method in the art that permits the detection of the longer polypeptide, such as, but not limited to, immunological methods.

Many methods standard in the art can be thus employed, including, but not limited to, immunoassays to detect and/or visualize gene expression (e.g., Western blot, 5 immunoprecipitation followed by sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE), immunocytochemistry, radioimmunoassays, ELISA (enzyme linked immunosorbent assay), "sandwich" immunoassays, immunoprecipitation assays, precipitin reactions, gel diffusion precipitin reactions, immunodiffusion assays, agglutination assays, complement-fixation assays, immunoradiometric assays, fluorescent 10 immunoassays, protein A immunoassays, or an epitope tag using an antibody that is specific to the polypeptide encoded by the gene of interest) and/or hybridization assays to detect gene expression by detecting and/or visualizing respectively mRNA encoding a gene (e.g., Northern assays, dot blots, *in situ* hybridization, etc), etc. Preferably, the antibody is specific to the C-terminal portion of the polypeptide used in an immunoassay. Such assays 15 are routine and well known in the art (see, e.g., Ausubel et al, eds, 1994, Current Protocols in Molecular Biology, Vol. 1, John Wiley & Sons, Inc., New York, which is incorporated by reference herein in its entirety). Exemplary immunoassays are described briefly below (but are not intended by way of limitation).

Immunoprecipitation protocols generally comprise lysing a population of 20 cells in a lysis buffer such as RIPA buffer (1% NP-40 or Triton X-100, 1% sodium deoxycholate, 0.1% SDS, 0.15 M NaCl, 0.01 M sodium phosphate at pH 7.2, 1% Trasylol) supplemented with protein phosphatase and/or protease inhibitors (e.g., EDTA, PMSF, aprotinin, sodium vanadate), adding the antibody which recognizes the antigen to the cell lysate, incubating for a period of time (e.g., 1 to 4 hours) at 40° C, adding protein A and/or 25 protein G sepharose beads to the cell lysate, incubating for about an hour or more at 40° C, washing the beads in lysis buffer and resuspending the beads in SDS/sample buffer. The ability of the antibody to immunoprecipitate a particular antigen can be assessed by, e.g., western blot analysis. One of skill in the art would be knowledgeable as to the parameters that can be modified to increase the binding of the antibody to an antigen and decrease the 30 background (e.g., pre-clearing the cell lysate with sepharose beads). For further discussion regarding immunoprecipitation protocols see, e.g., Ausubel et al, eds, 1994, Current Protocols in Molecular Biology, Vol. 1, John Wiley & Sons, Inc., New York at 10.16.1.

Western blot analysis generally comprises preparing protein samples, electrophoresis of the protein samples in a polyacrylamide gel (e.g., 8%- 20% SDS-PAGE

depending on the molecular weight of the antigen), transferring the protein sample from the polyacrylamide gel to a membrane such as nitrocellulose, PVDF or nylon, blocking the membrane in blocking solution (*e.g.*, PBS with 3% BSA or non-fat milk), washing the membrane in washing buffer (*e.g.*, PBS-Tween 20), blocking the membrane with primary

- 5 antibody (the antibody which recognizes the antigen) diluted in blocking buffer, washing the membrane in washing buffer, blocking the membrane with a secondary antibody (which recognizes the primary antibody, *e.g.*, an anti-human antibody) conjugated to an enzymatic substrate (*e.g.*, horseradish peroxidase or alkaline phosphatase) or radioactive molecule (*e.g.*, ^{32}P or ^{125}I) diluted in blocking buffer, washing the membrane in wash buffer, and
10 detecting the presence of the antigen. One of skill in the art would be knowledgeable as to the parameters that can be modified to increase the signal detected and to reduce the background noise. For further discussion regarding western blot protocols see, *e.g.*, Ausubel et al, eds, 1994, Current Protocols in Molecular Biology, Vol. 1, John Wiley & Sons, Inc., New York at 10.8.1.

- 15 ELISAs comprise preparing antigen, coating the well of a 96 well microtiter plate with the antigen, adding a primary antibody (which recognizes the antigen) conjugated to a detectable compound such as an enzymatic substrate (*e.g.*, horseradish peroxidase or alkaline phosphatase) to the well and incubating for a period of time, and detecting the presence of the antigen. In ELISAs the antibody of interest does not have to be conjugated
20 to a detectable compound; instead, a second antibody (which recognizes the primary antibody) conjugated to a detectable compound may be added to the well. Further, instead of coating the well with the antigen, the antibody may be coated to the well. In this case, a second antibody conjugated to a detectable compound may be added following the addition of the antigen of interest to the coated well. One of skill in the art would be knowledgeable
25 as to the parameters that can be modified to increase the signal detected as well as other variations of ELISAs known in the art. For further discussion regarding ELISAs see, *e.g.*, Ausubel et al, eds, 1994, Current Protocols in Molecular Biology, Vol. 1, John Wiley & Sons, Inc., New York at 11.2.1.

- 30 Methods for detecting the activity of a protein encoded by a reporter gene will vary with the reporter gene used. Assays for the various reporter genes are well-known to one of skill in the art. For example, as described in Section 5.1.1., luciferase, beta-galactosidase ("beta-gal"), beta-glucuronidase ("GUS"), beta-lactamase, chloramphenicol acetyltransferase ("CAT"), and alkaline phosphatase ("AP") are enzymes that can be analyzed in the presence of a substrate and could be amenable to high throughput screening.

For example, the reaction products of luciferase, beta-galactosidase (“beta-gal”), and alkaline phosphatase (“AP”) are assayed by changes in light imaging (e.g., luciferase), spectrophotometric absorbance (e.g., beta-gal), or fluorescence (e.g., AP). Assays for changes in light output, absorbance, and/or fluorescence are easily adapted for high throughput screening. For example, beta-gal activity can be measured with a microplate reader. Green fluorescent protein (“GFP”) activity can be measured by changes in fluorescence. For example, in the case of mutant GFPs that fluoresce at 488 nm, standard fluorescence activated cell sorting (“FACS”) equipment can be used to separate cells based upon GFP activity.

Changes in mRNA stability of the gene encoding the premature translation stop codon can be measured. As discussed above, nonsense-mediated mRNA decay alters the stability of an mRNA with a premature translation stop codon so that such mRNA is targeted for rapid decay instead of translation. In the presence of a compound that modulates premature translation termination and/or nonsense-mediated mRNA decay, the stability of the mRNA with the premature translation stop codon is likely altered, *i.e.*, stabilized. Methods of measuring changes in steady state levels of mRNA are well-known to one of skill in the art. Such methods include, but are not limited to, Northern blots, dot blots, solution hybridization, RNase protection assays, and S1 nuclease protection assays, wherein the steady state levels of the mRNA of interest are measured with an appropriately labeled nucleic acid probe. Alternatively, methods such as semi-quantitative polymerase chain reaction (“PCR”) can be used to measure changes in steady state levels of the mRNA of interest using the appropriate primers for amplification.

Alterations in the expression of a reporter gene may be determined by comparing the level of expression and/or activity of the reporter gene to a negative control (e.g., PBS or another agent that is known to have no effect on the expression of the reporter gene) and optionally, a positive control (e.g., an agent that is known to have an effect on the expression of the reporter gene, preferably an agent that effects premature translation termination and/or nonsense-mediated mRNA decay). Alternatively, alterations in the expression and/or activity of a reporter gene may be determined by comparing the level of expression and/or activity of the reporter gene to a previously determined reference range.

5.3.2. Cell-Free Extracts

After a vector containing the reporter gene construct and/or a vector(s) containing a nucleic acid sequence comprising a regulatory protein, a component or a subunit thereof is produced, a cell-free translation extract is generated or purchased, and a

compound library is synthesized or purchased or both, the cell-free translation extract and nucleic acid sequences are used to screen the library to identify compounds that modulate premature translation termination and/or nonsense-mediated mRNA decay. The reporter gene-based assays may be conducted in a cell-free manner by contacting a compound with a

- 5 cell-free extract and a reporter gene construct comprising a reporter gene containing within the open reading frame of the reporter gene a premature stop codon or nonsense mutation, and measuring the expression and/or activity of said reporter gene. The reporter gene cell-free assays may also be conducted by contacting a compound with a cell-free extract and an *in vitro* transcribed RNA of a reporter gene, wherein the RNA product contains a premature
10 stop codon or a nonsense mutation and measuring the expression and or activity of the protein encoded by the RNA product. Techniques for *in vitro* transcription are well-known to one of skill in the art or described herein (see, e.g. the Example in section 7). The reporter gene cell-free assays may also be conducted by: (a) contacting a compound with a cell-free extract, a first nucleic acid sequence and a second nucleic acid sequence, wherein
15 the first nucleic acid sequence comprises a regulatory element operably linked to a reporter gene and the second nucleic acid sequence comprises a nucleotide sequence with a premature stop codon or nonsense mutation that encodes a regulatory protein that binds to the regulatory element of the first nucleic acid sequence and regulates the expression of the reporter gene; and (b) measuring the expression and/or activity of the reporter gene.

20 The reporter gene cell-free assays may also be conducted by: (a) contacting a compound with a cell-free extract, a first nucleic acid sequence, a second nucleic acid sequence and a third nucleic acid sequence, wherein (i) the first nucleic acid sequence comprises a nucleotide sequence encoding a first fusion protein comprising a DNA binding domain and a first protein, the nucleotide sequence of the DNA binding domain or the first
25 protein containing a premature stop codon or nonsense mutation, (ii) the second nucleic acid sequence comprises a nucleotide sequence encoding a second fusion protein comprising an activation domain and a second protein, the second protein interacting with the first protein to produce a regulatory protein, and (iii) the third nucleic acid sequence comprises a regulatory element operably linked to a reporter gene, the expression of the reporter gene
30 being regulated by the binding of the regulatory protein to the regulatory element: and (b) measuring the expression and/or activity of the reporter gene. Further, the reporter gene cell-free assays may also be conducted by: (a) contacting a compound with a cell-free extract, a first nucleic acid sequence, a second nucleic acid sequence and a third nucleic acid sequence, wherein (i) the first nucleic acid sequence comprises a nucleotide sequence

encoding a first fusion protein comprising a DNA binding domain and a first protein, (ii) the second nucleic acid sequence comprises a nucleotide sequence encoding a second fusion protein comprising an activation domain and a second protein, the nucleotide sequence of the activation domain or the second protein containing a premature stop codon or nonsense mutation, and the second protein interacting with the first protein to produce a regulatory protein, and (iii) the third nucleic acid sequence comprises a regulatory element operably linked to a reporter gene, the expression of the reporter gene being regulated by the binding of the regulatory protein to the regulatory element: and (b) measuring the expression and/or activity of the reporter gene.

In the cell-free reporter gene assays described herein, the alteration in reporter gene expression or activity relative to a previously determined reference range, or to the expression or activity of the reporter gene in the absence of the compound or the presence of an appropriate control (*e.g.*, a negative control) indicates that a particular compound modulates premature translation termination and/or nonsense-mediated mRNA decay. In particular, a decrease in reporter gene expression or activity relative to a previously determined reference range, or to the expression in the absence of the compound or the presence of an appropriate control (*e.g.*, a negative control) may, depending upon the parameters of the reporter gene assay, indicate that a particular compound reduces or suppresses premature translation termination and/or nonsense-mediated mRNA decay. In contrast, an increase in reporter gene expression or activity relative to a previously determined reference range, or to the expression in the absence of the compound or the presence of an appropriate control (*e.g.*, a negative control) may, depending upon the parameters of the reporter gene-based assay, indicate that a particular compound enhances premature translation termination and/or nonsense-mediated mRNA decay.

In accordance with the invention, the step of contacting a compound with a cell-free extract and a nucleic acid sequence in the reporter gene-based assays described herein is preferably conducted in an aqueous solution comprising a buffer and a combination of salts (such as KCl, NaCl and/or MgCl₂). The optimal concentration of each salt used in the aqueous solution is dependent on, *e.g.*, the protein, polypeptide or peptide encoded by the nucleic acid sequence (*e.g.*, the regulatory protein) and the compounds used, and can be determined using routine experimentation. In a specific embodiment, the aqueous solution approximates or mimics physiologic conditions. In another specific embodiment, the aqueous solution further comprises a detergent or a surfactant.

The cell-free reporter gene assays of the present invention can be performed using different incubation times. The cell-free extract and the nucleic acid sequence(s) (e.g., a reporter gene) can be incubated together before the addition of a compound or a member of a library of compounds. In certain embodiments, the cell-free extract are incubated with a nucleic acid sequence(s) (e.g., a reporter gene) before the addition of a compound or a member of a library of compounds for at least 0.2 hours, 0.25 hours, 0.5 hours, 1 hour, 2 hours, 3 hours, 4 hours, 5 hours, 6 hours, 8 hours, 10 hours, 12 hours, 18 hours, or at least 1 day. In other embodiments, the cell-free extract, or the nucleic acid sequence(s) (e.g., a reporter gene) is incubated with a compound or a member of a library of compounds before the addition of the nucleic acid sequence(s) (e.g., a reporter gene), or the cell-free extract, respectively. In certain embodiments, a compound or a member of a library of compounds is incubated with a nucleic acid sequence(s) (e.g., a reporter gene) or cell-free extract before the addition of the remaining component, *i.e.*, cell-free extract, or a nucleic acid sequence(s) (e.g., a reporter gene), respectively, for at least 0.2 hours, 0.25 hours, 0.5 hours, 1 hour, 2 hours, 3 hours, 4 hours, 5 hours, 6 hours, 8 hours, 10 hours, 12 hours, 18 hours, or at least 1 day. Once the reaction vessel comprises the components, *i.e.*, a compound or a member of a library of compounds, the cell-free extract and the nucleic acid sequence(s) (e.g., a reporter gene), the reaction may be further incubated for at least 0.2 hours, 0.25 hours, 0.5 hours, 1 hour, 2 hours, 3 hours, 4 hours, 5 hours, 6 hours, 8 hours, 10 hours, 12 hours, 18 hours, or at least 1 day.

The progress of the reaction in the cell-free reporter gene-based assays can be measured continuously. Alternatively, time-points may be taken at different times of the reaction to monitor the progress of the reaction in the cell-free reporter gene-based assays.

The activity of a compound in the cell-free extract can be determined by assaying the activity of a reporter protein encoded by a reporter gene, or alternatively, by quantifying the expression of the reporter gene by, for example, labeling the *in vitro* translated protein (e.g., with ^{35}S -labeled methionine), northern blot analysis, RT-PCR or by immunological methods, such as western blot analysis or immunoprecipitation. Such methods are well-known to one of skill in the art. Examples of assays which can be used to measure the expression and/or activity of a reporter gene are described in Section 5.3.1 *supra*.

5.4. Characterization of the Structure of Compounds

If the library comprises arrays or microarrays of compounds, wherein each compound has an address or identifier, the compound can be deconvoluted, *e.g.*, by cross-

referencing the positive sample to original compound list that was applied to the individual test assays.

If the library is a peptide or nucleic acid library, the sequence of the compound can be determined by direct sequencing of the peptide or nucleic acid. Such methods are well known to one of skill in the art.

A number of physico-chemical techniques can be used for the *de novo* characterization of compounds that mediate premature translation termination and/or nonsense-mediated mRNA decay.

5.4.1. Mass Spectrometry

The invention provides, in part, for mass spectrometry methods to identify or characterize the compounds of the invention. Any mass spectrometric method can be used, for example, those employing an ionizer, ion analyzer and detector.

A number of techniques can be used in order to ionize a sample for investigative or characterization purposes. Such techniques form the charged particles required for analysis. Examples of ionization methods include, but are not limited to, electron impact, chemical ionization, electrospray ionization, fast atom bombardment and matrix assisted laser desorption ionization. The technique used for ionization will depend on the type of analyte being examined and the conditions necessary for acquisition. For example, electron impact and chemical ionization would be preferred with a relatively small volatile sample with a mass of 1 to 1000 daltons; electrospray ionization would be preferred with peptides, proteins and non-volatile samples with a mass of up to 200,000 daltons, fast atom bombardment would be preferred with carbohydrates, organometallics, peptides and nonvolatile compounds and matrix assisted laser desorption ionization would be preferred when examining peptides, proteins and nucleotides.

A number of ion analysis techniques can be used, in particular those where molecular ions and fragment ions are accelerated by manipulation of charged particles through the mass spectrometer. Such analyzers include, but are not limited to, quadropole, sector (magnetic and/or electrostatic), time of flight (TOF), and ion cyclotron resonance (ICR). The technique used for analysis would depend on the sample and the conditions for acquisition. For example, one might prefer quadropole when desiring a unit mass resolution, fast scan time, and low cost; one might prefer a sector analyzer when desiring high resolution and an exact mass; one might prefer time of flight when desiring no limitation for m/z maximum and a high throughput; and one might prefer ion cyclotron

resonance when desiring very high resolution an exact mass and also to perform ion chemistry.

Any ionizer method can be combined with any ion analyzer technique. There are many types of detectors that may be used as part of the mass spectrophotometric methods of the invention, in particular those that produce an electronic signal when struck by an ion. Calibration would be necessarily performed by introducing a well known compound into the instrument and adjusting the circuits so that the compound's molecular ion and fragment ions are reported accurately.

Mass spectrometry (*e.g.*, electrospray ionization ("ESI") and matrix-assisted laser desorption-ionization ("MALDI"), Fourier-transform ion cyclotron resonance ("FT-ICR") can be used both for high-throughput screening of compounds that bind to a target RNA and elucidating the structure of the compound.

MALDI uses a pulsed laser for desorption of the ions and a time-of-flight analyzer, and has been used for the detection of noncovalent tRNA:amino-acyl-tRNA synthetase complexes (Gruic-Sovulj *et al.*, 1997, *J. Biol. Chem.* 272:32084-32091). However, covalent cross-linking between the target nucleic acid and the compound is required for detection, since a non-covalently bound complex may dissociate during the MALDI process.

ESI mass spectrometry ("ESI-MS") has been of greater utility for studying non-covalent molecular interactions because, unlike the MALDI process, ESI-MS generates molecular ions with little to no fragmentation (Xavier *et al.*, 2000, *Trends Biotechnol.* 18(8):349-356). ESI-MS has been used to study the complexes formed by HIV Tat peptide and protein with the TAR RNA (Sannes-Lowery *et al.*, 1997, *Anal. Chem.* 69:5130-5135).

Fourier-transform ion cyclotron resonance ("FT-ICR") mass spectrometry provides high-resolution spectra, isotope-resolved precursor ion selection, and accurate mass assignments (Xavier *et al.*, 2000, *Trends Biotechnol.* 18(8):349-356). FT-ICR has been used to study the interaction of aminoglycoside antibiotics with cognate and non-cognate RNAs (Hofstadler *et al.*, 1999, *Anal. Chem.* 71:3436-3440; Griffey *et al.*, 1999, *Proc. Natl. Acad. Sci. USA* 96:10129-10133). As true for all of the mass spectrometry methods discussed herein, FT-ICR does not require labeling of the target RNA or a compound.

An advantage of mass spectroscopy is not only the elucidation of the structure of a compound, but also the determination of the structure of the compound bound

to a target RNA. Such information can enable the discovery of a consensus structure of a compound that specifically binds to a target RNA.

5.4.2. NMR Spectroscopy

The invention provides, in part, NMR spectroscopic techniques that may be used, for example, to characterize and identify small and large molecules of the invention. NMR methods are advantageous in understanding characteristics of the compounds of the invention because it allows rapid acquisition of single and multi-dimensional structural data about a compound in solution. Moreover, the NMR technique is a non-destructive technique that also provides dynamic information relating to a compound's behavior in complex or in association with other molecules of interest. There are a variety of techniques that can be used to examine compounds of the invention using NMR methods. In particular, any type of NMR spectrometer can be used, including, but not limited to, those of low, medium and high magnetic field. In a preferred embodiment, the NMR spectrometer that is used has a high magnetic field, in particular, if the compound has a high molecular weight, such as, those greater than 1000 daltons.

Any technique known in the art can be used to acquire data on the compounds and also to produce spectra for interpretation, including, but not limited to, those that measure through bond correlations and through space correlations. Both single and multi-dimensional spectra can be produced. In another embodiment, the technique that is used is homonuclear. In yet another embodiment, the technique is heteronuclear. In one embodiment of the invention, correlation spectroscopy, e.g., COSY or TOCSY, methods are used to measure through bond correlations. In another embodiment of the invention, nuclear overhauser effect spectroscopy methods, e.g., NOESY, are used to measure through space correlations. In yet another embodiment of the invention, multi-dimensional methods are used to identify relationships between heterologous nucleii, e.g., heteronuclear single quantum coherence (HSQC) and heteronuclear multiple quantum coherence (HMQC).

In another embodiment of the invention, NMR methods are used to characterize compounds that are associated with other molecules. For example, complexed target nucleic acids can be examined by qualitatively determining changes in chemical shift, specifically from distances measured using relaxation effects, and NMR-based approaches have been used in the identification of small molecule binders of protein drug targets (Xavier *et al.*, 2000, Trends Biotechnol. 18(8):349-356). The determination of structure-activity relationships ("SAR") by NMR is the first method for NMR described in which small molecules that bind adjacent subsites are identified by two-dimensional ^1H - ^{15}N spectra

of the target protein (Shuker *et al.*, 1996, *Science* 274:1531-1534). The signal from the bound molecule is monitored by employing line broadening, transferred NOEs and pulsed field gradient diffusion measurements (Moore, 1999, *Curr. Opin. Biotechnol.* 10:54-58). A strategy for lead generation by NMR using a library of small molecules has been recently 5 described (Fejzo *et al.*, 1999, *Chem. Biol.* 6:755-769).

Other examples of NMR methods that can be used for the invention include, but are not limited to, one-dimensional, two-dimensional, three dimension, four dimensional NMR methods as well as correlation spectroscopy ("COSY"), and nuclear Overhauser effect ("NOE") spectroscopy. Such methods of structure determination of compounds are 10 well known to one of skill in the art.

Similar to mass spectroscopy, an advantage of NMR is the not only the elucidation of the structure of a compound, but also the determination of the structure of the compound bound to the target RNA. Such information can enable the discovery of a consensus structure of a compound that specifically binds to a target RNA.

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5.4.3. X-ray Crystallography

X-ray crystallography can be used to elucidate the structure of a compound. For a review of x-ray crystallography see, *e.g.*, Blundell *et al.* 2002, *Nat Rev Drug Discov* 1(1):45-54. The first step in x-ray crystallography is the formation of crystals. The formation of crystals begins with the preparation of highly purified and soluble samples.

20 The conditions for crystallization is then determined by optimizing several solution variables known to induce nucleation, such as pH, ionic strength, temperature, and specific concentrations of organic additives, salts and detergent. Techniques for automating the crystallization process have been developed to automate the production of high-quality protein crystals. Once crystals have been formed, the crystals are harvested and prepared 25 for data collection. The crystals are then analyzed by diffraction (such as multi-circle diffractometers, high-speed CCD detectors, and detector off-set). Generally, multiple crystals must be screened for structure determinations.

A number of methods can be used to acquire a diffraction pattern so that a compound can be characterized. In one embodiment, an X-ray source is provided, for 30 example, by a rotating anode generator producing an X-ray beam of a characteristic wavelength. There are a number of sources of X-ray radiation that may be used in the methods of the invention, including low and high intensity radiation. In one example, the tunable X-ray radiation is produced by a Synchrotron. In another embodiment, the primary X-ray beam is monochromated by either crystal monochromators or focusing mirrors and

the beam is passed through a helium flushed collimator. In a preferred embodiment, the crystal is mounted on a pin on a goniometer head, that is mounted to a goniometer which allows to position the crystal in different orientations in the beam. The diffracted X-rays can be recorded using a number of techniques, including, but not limited to image plates, 5 multiwire detectors or CCD cameras. In other embodiments, flash cooling, for example, of protein crystals, to cryogenic temperatures (~100 K) offers many advantages, the most significant of which is the elimination of radiation damage.

5.4.4. Vibrational Spectroscopy

10 Vibrational spectroscopy (e.g., but not limited to, infrared (IR) spectroscopy or Raman spectroscopy) can be used for elucidating the structure of a compound.

Infrared spectroscopy measures the frequencies of infrared light (wavelengths from 100 to 10,000 nm) absorbed by the compound as a result of excitation of 15 vibrational modes according to quantum mechanical selection rules which require that absorption of light cause a change in the electric dipole moment of the molecule. The infrared spectrum of any molecule is a unique pattern of absorption wavelengths of varying intensity that can be considered as a molecular fingerprint to identify or characterize any compound.

Infrared spectra can be measured in a scanning mode by measuring the 20 absorption of individual frequencies of light, produced by a grating which separates frequencies from a mixed-frequency infrared light source, by the compound relative to a standard intensity (double-beam instrument) or pre-measured ('blank') intensity (single-beam instrument). In a preferred embodiment, infrared spectra are measured in a pulsed mode ("FT-IR") where a mixed beam, produced by an interferometer, of all infrared 25 light frequencies is passed through or reflected off the compound. The resulting interferogram, which may or may not be added with the resulting interferograms from subsequent pulses to increase the signal strength while averaging random noise in the electronic signal, is mathematically transformed into a spectrum using Fourier Transform or Fast Fourier Transform algorithms.

Raman spectroscopy measures the difference in frequency due to absorption 30 of infrared frequencies of scattered visible or ultraviolet light relative to the incident beam. The incident monochromatic light beam, usually a single laser frequency, is not truly absorbed by the compound but interacts with the electric field transiently. Most of the light scattered off the sample will be unchanged (Rayleigh scattering) but a portion of the scatter light will have frequencies that are the sum or difference of the incident and molecular

vibrational frequencies. The selection rules for Raman (inelastic) scattering require a change in polarizability of the molecule. While some vibrational transitions are observable in both infrared and Raman spectrometry, must are observable only with one or the other technique. The Raman spectrum of any molecule is a unique pattern of absorption wavelengths of varying intensity that can be considered as a molecular fingerprint to identify any compound.

Raman spectra are measured by submitting monochromatic light to the sample, either passed through or preferably reflected off, filtering the Rayleigh scattered light, and detecting the frequency of the Raman scattered light. An improved Raman spectrometer is described in US Patent No. 5,786,893 to Fink et al., which is hereby incorporated by reference.

Vibrational microscopy can be measured in a spatially resolved fashion to address single beads by integration of a visible microscope and spectrometer. A microscopic infrared spectrometer is described in U.S. Patent No. 5,581,085 to Reffner et al., which is hereby incorporated by reference in its entirety. An instrument that simultaneously performs a microscopic infrared and microscopic Raman analysis on a sample is described in U.S. Patent No. 5,841,139 to Sostek et al., which is hereby incorporated by reference in its entirety.

In one embodiment of the method, compounds are synthesized on polystyrene beads doped with chemically modified styrene monomers such that each resulting bead has a characteristic pattern of absorption lines in the vibrational (IR or Raman) spectrum, by methods including but not limited to those described by Fenniri et al., 2000, J. Am. Chem. Soc. 123:8151-8152. Using methods of split-pool synthesis familiar to one of skill in the art, the library of compounds is prepared so that the spectroscopic pattern of the bead identifies one of the components of the compound on the bead. Beads that have been separated according to their ability to bind target RNA can be identified by their vibrational spectrum. In one embodiment of the method, appropriate sorting and binning of the beads during synthesis then allows identification of one or more further components of the compound on any one bead. In another embodiment of the method, partial identification of the compound on a bead is possible through use of the spectroscopic pattern of the bead with or without the aid of further sorting during synthesis, followed by partial resynthesis of the possible compounds aided by doped beads and appropriate sorting during synthesis.

In another embodiment, the IR or Raman spectra of compounds are examined while the compound is still on a bead, preferably, or after cleavage from bead,

using methods including but not limited to photochemical, acid, or heat treatment. The compound can be identified by comparison of the IR or Raman spectral pattern to spectra previously acquired for each compound in the combinatorial library.

5 **5.5. Naturally Occurring Genes with Premature Stop Codons:
Examples of Disorders and Diseases**

The invention provides for naturally occurring genes with premature stop codons to ascertain the effects of compounds on premature translation termination and/or nonsense-mediated mRNA decay. In general, the expression of the gene product, in particular, a full-length gene product, is indicative of the effect of the compounds on premature translation termination and/or nonsense-mediated mRNA decay.

10 In a preferred embodiment, the naturally occurring genes with premature stop codons are genes that cause diseases which are due, in part, to the lack of expression of the gene resulting from the premature stop codon. Such diseases include, but are not limited to, cystic fibrosis, muscular dystrophy, heart disease (e.g., familial hypercholesterolemia),
15 p53-associated cancers (e.g., lung, breast, colon, pancreatic, non-Hodgkin's lymphoma, ovarian, and esophageal cancer), colorectal carcinomas, neurofibromatosis, retinoblastoma, Wilm's tumor, retinitis pigmentosa, collagen disorders (e.g., osteogenesis imperfecta and cirrhosis), Tay Sachs disease, blood disorders (e.g., hemophilia, von Willebrand disease, b-Thalassemia), kidney stones, ataxia-telangiectasia, lysosomal storage diseases, and tuberous
20 sclerosis. Genes involved in the etiology of these diseases are discussed below.

25 The recognition of translation termination signals is not necessarily limited to a simple trinucleotide stop codon, but is instead recognized by the sequences surrounding the stop codon in addition to the stop codon itself (see, e.g., Manuvakhova *et al.*, 2000, RNA 6(7):1044-1055, which is hereby incorporated by reference in its entirety). Thus, any genes containing particular tetranucleotide sequences at the stop codon, such as, but not limited to, UGAC, UAGU, UAGC, UAGG, UAGA, UGAA, UGAG, UGAU, UAAC, UAAU, UAAG, and UAAA, are candidates of naturally occurring genes with premature stop codons that are useful in the present invention. Human disease genes that contain these particular sequence motifs are sorted by chromosome is presented as an Example in Section
30 8.

5.5.1. Cystic Fibrosis

Cystic fibrosis is caused by mutations in the cystic fibrosis conductance regulator ("CFTR") gene. Such mutations vary between populations and depend on a multitude of factors such as, but not limited to, ethnic background and geographic location.

Nonsense mutations in the CFTR gene are expected to produce little or no CFTR chloride channels. Several nonsense mutations in the CFTR gene have been identified (see, e.g., Tzetis *et al.*, 2001, *Hum Genet.* 109(6):592-601. Strandvik *et al.*, 2001, *Genet Test.* 5(3):235-42; Feldmann *et al.*, 2001, *Hum Mutat.* 17(4):356; Wilschanski *et al.*, 2000, *Am J Respir Crit Care Med.* 161(3 Pt 1):860-5; Castaldo *et al.*, 1999, *Hum Mutat.* 14(3):272; Mitre *et al.*, 1999, *Hum Mutat.* 14(2):182; Mickle *et al.*, 1998, *Hum Mol Genet.* 7(4):729-35; Casals *et al.*, 1997, *Hum Genet.* 101(3):365-70; Mitre *et al.*, 1996, *Hum Mutat.* 8(4):392-3; Bonizzato *et al.*, 1995, *Hum Genet.* 1995 Apr;95(4):397-402; Greil *et al.*, 1995, *Wien Klin Wochenschr.* 107(15):464-9; Zielenski *et al.*, 1995, *Hum Mutat.* 5(1):43-7; Dork *et al.*, 1994, *Hum Genet.* 94(5):533-42; Balassopoulou *et al.*, 1994, *Hum Mol Genet.* 3(10):1887-8; Ghanem *et al.*, 1994, 21(2):434-6; Will *et al.*, *J Clin Invest.* 1994 Apr;93(4):1852-9; Hull *et al.*, 1994, *Genomics.* 1994 Jan 15;19(2):362-4; Dork *et al.*, 1994, *Hum Genet.* 93(1):67-73; Rolfini & Cabrini, 1993, *J Clin Invest.* 92(6):2683-7; Will *et al.*, 1993, *J Med Genet.* 30(10):833-7; Bienvenu *et al.*, 1993, *J Med Genet.* 30(7):621-2; Cheadle *et al.*, 1993, *Hum Mol Genet.* 2(7):1067-8; Casals *et al.*, 1993, *Hum Genet.* 91(1):66-70; Reiss *et al.*, 1993, *Hum Genet.* 91(1):78-9; Chevalier-Porst *et al.*, 1992, *Hum Mol Genet.* 1(8):647-8; Hamosh *et al.*, 1992, *Hum Mol Genet.* 1(7):542-4; Gasparini *et al.*, 1992, *J Med Genet.* 29(8):558-62; Fanen *et al.*, 1992, *Genomics.* 13(3):770-6; Jones *et al.*, 1992, *Hum Mol Genet.* 1(1):11-7; Ronchetto *et al.*, 1992, *Genomics.* 12(2):417-8.; Macek *et al.*, 1992, *Hum Mutat.* 1(6):501-2; Shoshani *et al.*, 1992, *Am J Hum Genet.* 50(1):222-8; Schloesser *et al.*, 1991, *J Med Genet.* 28(12):878-80; Hamosh *et al.*, 1991, *J Clin Invest.* 88(6):1880-5; Bal *et al.*, 1991, *J Med Genet.* 28(10):715-7; Dork *et al.*, 1991, *Hum Genet.* 87(4):441-6; Beaudet *et al.*, 1991, *Am J Hum Genet.* 48(6):1213; Gasparini *et al.*, 1991, *Genomics.* 10(1):193-200; Cutting *et al.*, 1990, *N Engl J Med.* 1990, 323(24):1685-9; and Kerem *et al.*, 1990, *Proc Natl Acad Sci U S A.* 87(21):8447-51, the disclosures of which are hereby incorporated by reference in their entireties). Any CFTR gene encoding a premature translation codon including, but not limited to, the nonsense mutations described in the references cited above, can be used in the present invention to identify compounds that mediate premature translation termination and/or nonsense-mediated mRNA decay.

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5.5.2. Muscular Dystrophy

Muscular dystrophy is a genetic disease characterized by severe, progressive muscle wasting and weakness. Duchenne muscular dystrophy and Becker muscular dystrophy are generally caused by nonsense mutations of the dystrophin gene (see, e.g., Kerr *et al.*, 2001, *Hum Genet.* 109(4):402-7 and Wagner *et al.*, 2001, *Ann Neurol.*

49(6):706-11). Nonsense mutations in other genes have also been implicated in other types of muscular dystrophy, such as, but not limited to, collagen genes in Ullrich congenital muscular dystrophy (see, e.g., Demir *et al.*, 2002, Am J Hum Genet. 70(6):1446-58), the emerin gene and lamins genes in Emery-Dreifuss muscular dystrophy (see, e.g., Holt *et al.*, 5 2001, Biochem Biophys Res Commun. 287(5):1129-33; Becane *et al.*, 2000, Pacing Clin Electrophysiol. 23(11 Pt 1):1661-6; and Bonne *et al.*, 2000, Ann Neurol. 48(2):170-80.), the dysferlin gene in Miyoshi myopathy (see, e.g., Nakagawa *et al.*, 2001, J Neurol Sci. 184(1):15-9), the plectin gene in late onset muscular dystrophy (see, e.g., Bauer *et al.*, 2001, Am J Pathol. 158(2):617-25), the delta-sarcoglycan gene in recessive limb-girdle muscular 10 dystrophy (see, e.g., Duggan *et al.*, 1997, Neurogenetics. 1(1):49-58), the laminina2-chain gene in congenital muscular dystrophy (see, e.g., Mendell *et al.*, 1998, Hum Mutat. 12(2):135), the plectin gene in late-onset muscular dystrophy (see, e.g., Rouan *et al.*, 2000, J Invest Dermatol. 114(2):381-7 and Kunz *et al.*, 2000, J Invest Dermatol. 114(2):376-80), the myophosphorylase gene in McArdle's disease (see, e.g., Bruno *et al.*, 1999, Neuromuscul Disord. 9(1):34-7), and the collagen VI in Bethlem myopathy (see, e.g., Lamande *et al.*, 15 1998, Hum Mol Genet. 1998 Jun;7(6):981-9).

Several nonsense mutations in the dystrophin gene have been identified (see, e.g., Kerr *et al.*, 2001, Hum Genet. 109(4):402-7; Mendell *et al.*, 2001, Neurology 57(4):645-50; Fajkusova *et al.*, 2001, Neuromuscul Disord. 11(2):133-8; Ginjaar *et al.*, 20 2000, Eur J Hum Genet. 8(10):793-6; Lu *et al.*, 2000, J Cell Biol. 148(5):985-96; Tuffery-Giraud *et al.*, 1999, Hum Mutat. 14(5):359-68; Fajkusova *et al.*, 1998, J Neurogenet. 12(3):183-9; Tuffery *et al.*, 1998, Hum Genet. 102(3):334-42; Shiga *et al.*, 1997, J Clin Invest. 100(9):2204-10; Winnard *et al.*, 1995, Am J Hum Genet. 56(1):158-66; Prior *et al.*, 1994, Am J Med Genet. 50(1):68-73; Prior *et al.*, 1993, Hum Mol Genet. 25 2(3):311-3; Prior *et al.*, 1993, Hum Mutat. 2(3):192-5; Nigro *et al.*, 1992, Hum Mol Genet. 1(7):517-20; Worton, 1992, J Inherit Metab Dis. 15(4):539-50; and Bulman *et al.*, 1991, Genomics. 10(2):457-60; the disclosures of which are hereby incorporated by reference in their entireties). Any gene encoding a premature translation codon implicated in muscular dystrophy including, but not limited to, the nonsense mutations described in the references cited above, can be used in the present invention to identify compounds that mediate 30 premature translation termination and/or nonsense-mediated mRNA decay.

5.5.3. Familial Hypercholesterolemia

Hypercholesterolemia, or high blood cholesterol, results from either the overproduction or the underutilization of low density lipoprotein ("LDL").

Hypercholesterolemia is caused by either the genetic disease familial hypercholesterolemia or the consumption of a high cholesterol diet. Nonsense mutations in the LDL receptor gene have been implicated in familial hypercholesterolemia. Several nonsense mutations in the LDL receptor gene have been identified (see, e.g., Lind *et al.*, 2002, Atherosclerosis 163(2):399-407; Salazar *et al.*, 2002, Hum Mutat. 19(4):462-3; Kuhrova *et al.*, 2002, Hum Mutat. 19(1):80; Zakharova *et al.*, 2001, Bioorg Khim. 27(5):393-6; Kuhrova *et al.*, 2001, Hum Mutat. 18(3):253; Genschel *et al.*, 2001, Hum Mutat. 17(4):354; Weiss *et al.*, 2000, J Inherit Metab Dis. 23(8):778-90; Mozas *et al.*, 2000, Hum Mutat. 15(5):483-4; Shin *et al.*, 2000, Clin Genet. 57(3):225-9; Graham *et al.*, 1999, Atherosclerosis 147(2):309-16; Hattori *et al.*, 1999, Hum Mutat. 14(1):87; Cenarro *et al.*, 1998, Hum Mutat. 11(5):413; Rodninen *et al.*, 1999, Hum Mutat. 13(3):186-96; Hirayama *et al.*, 1998, J Hum Genet. 43(4):250-4; Lind *et al.*, 1998, J Intern Med. 244(1):19-25; Thiart *et al.*, 1997, Mol Cell Probes 11(6):457-8; Maruyama *et al.*, 1995, Arterioscler Thromb Vasc Biol. 15(10):1713-8; Koivisto *et al.*, 1995, Am J Hum Genet. 57(4):789-97; Lombardi *et al.*, 1995, J Lipid Res. 36(4):860-7; Leren *et al.*, 1993, Hum Genet. 92(1):6-10; Landsberger *et al.*, 1992, Am J Hum Genet. 50(2):427-33; Loux *et al.*, 1992, Hum Mutat. 1992;1(4):325-32; Motulsky, 1989, Arteriosclerosis. 9(1 Suppl):I3-7; Lehrman *et al.*, 1987, J Biol Chem. 262(1):401-10; and Lehrman *et al.*, 1985, Cell 41(3):735-43; the disclosures of which are hereby incorporated by reference in their entireties). Any LDL receptor gene encoding a premature translation codon including, but not limited to, the nonsense mutations described in the references cited above, can be used in the present invention to identify compounds that mediate premature translation termination and/or nonsense-mediated mRNA decay.

5.5.4. p53-associated Cancers

Mutant forms of the p53 protein, which is thought to act as a negative regulator of cell proliferation, transformation, and tumorigenesis, have been implicated as a common genetic change characteristic of human cancer (see, e.g., Levine *et al.*, 1991, Nature 351:453-456 and Hollstein *et al.*, 1991, Science 253:49-53). p53 mutations have been implicated in cancers such as, but not limited to, lung cancer, breast cancer, colon cancer, pancreatic cancer, non-Hodgkin's lymphoma, ovarian cancer, and esophageal cancer.

Nonsense mutations have been identified in the p53 gene and have been implicated in cancer. Several nonsense mutations in the p53 gene have been identified (see, e.g., Masuda *et al.*, 2000, Tokai J Exp Clin Med. 25(2):69-77; Oh *et al.*, 2000, Mol Cells 10(3):275-80; Li *et al.*, 2000, Lab Invest. 80(4):493-9; Yang *et al.*, 1999, Zhonghua Zhong

Liu Za Zhi 21(2):114-8; Finkelstein *et al.*, 1998, Mol Diagn. 3(1):37-41; Kajiyama *et al.*, 1998, Dis Esophagus. 11(4):279-83; Kawamura *et al.*, 1999, Leuk Res. 23(2):115-26; Radig *et al.*, 1998, Hum Pathol. 29(11):1310-6; Schuyer *et al.*, 1998, Int J Cancer 76(3):299-303; Wang-Gohrke *et al.*, 1998, Oncol Rep. 5(1):65-8; Fulop *et al.*, 1998, J Reprod Med. 43(2):119-27; Ninomiya *et al.*, 1997, J Dermatol Sci. 14(3):173-8; Hsieh *et al.*, 1996, Cancer Lett. 100(1-2):107-13; Rall *et al.*, 1996, Pancreas. 12(1):10-7; Fukutomi *et al.*, 1995, Nippon Rinsho. 53(11):2764-8; Frebourg *et al.*, 1995, Am J Hum Genet. 56(3):608-15; Dove *et al.*, 1995, Cancer Surv. 25:335-55; Adamson *et al.*, 1995, Br J Haematol. 89(1):61-6; Grayson *et al.*, 1994, Am J Pediatr Hematol Oncol. 16(4):341-7; Lepelley *et al.*, 1994, Leukemia. 8(8):1342-9; McIntyre *et al.*, 1994, J Clin Oncol. 12(5):925-30; Horio *et al.*, 1994, Oncogene. 9(4):1231-5; Nakamura *et al.*, 1992, Jpn J Cancer Res. 83(12):1293-8; Davidoff *et al.*, 1992, Oncogene. 7(1):127-33; and Ishioka *et al.*, 1991, Biochem Biophys Res Commun. 177(3):901-6; the disclosures of which are hereby incorporated by reference in their entireties). Any p53 gene encoding a premature translation codon including, but not limited to, the nonsense mutations described in the references cited above, can be used in the present invention to identify compounds that mediate premature translation termination and/or nonsense-mediated mRNA decay.

5.5.5. Colorectal Carcinomas

Molecular genetic abnormalities resulting in colorectal carcinoma involve tumor-suppressor genes that undergo inactivation (such as, but not limited to, *apc*, *mcc*, *dcc*, *p53*, and possibly genes on chromosomes 8p, 1p, and 22q) and dominant-acting oncogenes (such, but not limited to, *ras*, *src*, and *myc*) (see, e.g., Hamilton, 1992, Cancer 70(5 Suppl):1216-21). Nonsense mutations in the adenomatous polyposis coli ("APC") gene and mismatch repair genes (such as, but not limited to, *mlh1* and *msh2*) have also been described. Nonsense mutations have been implicated in colorectal carcinomas (see, e.g., Viel *et al.*, 1997, Genes Chromosomes Cancer. 18(1):8-18; Akiyama *et al.*, 1996, Cancer 78(12):2478-84; Itoh & Imai , 1996, Hokkaido Igaku Zasshi 71(1):9-14; Kolodner *et al.*, 1994, Genomics. 24(3):516-26; Ohue *et al.*, 1994, Cancer Res. 54(17):4798-804; and Yin *et al.*, 1993, Gastroenterology. 104(6):1633-9; the disclosures of which are hereby incorporated by reference in their entireties). Any gene encoding a premature translation codon implicated in colorectal carcinoma including, but not limited to, the nonsense mutations described in the references cited above, can be used in the present invention to identify compounds that mediate premature translation termination and/or nonsense-mediated mRNA decay.

5.5.6. Neurofibromatosis

Neurofibromatosis is an inherited disorder, which is commonly caused caused by mutations in the NF1 and NF2 tumor suppressor genes. It is characterized by multiple intracranial tumors including schwannomas, meningiomas, and ependymomas.

- 5 Nonsense mutations in the NF1 and NF2 genes have been described. Nonsense mutations have been implicated in neurofibromatosis (see, e.g., Lamszus *et al.*, 2001, Int J Cancer 91(6):803-8; Sestini *et al.*, 2000, Hum Genet. 107(4):366-71; Fukasawa *et al.*, 2000, Jpn J Cancer Res. 91(12):1241-9; Park *et al.*, 2000, J Hum Genet. 45(2):84-5; Ueki *et al.*, 1999, Cancer Res. 59(23):5995-8; , 1999, Hokkaido Igaku Zasshi. 74(5):377-86; Buske *et al.*,
- 10 1999, Am J Med Genet. 86(4):328-30; Harada *et al.*, 1999, Surg Neurol. 51(5):528-35; Krkljus *et al.*, 1998, Hum Mutat. 11(5):411; Klose *et al.*, 1999, Am J Med Genet. 83(1):6-12; Park & Pivnick, 1998, J Med Genet. 35(10):813-20; Bahauau *et al.*, 1998, Am J Med Genet. 75(3):265-72; Bijlsma *et al.*, 1997, J Med Genet. 34(11):934-6; MacCollin *et al.*, 1996, Ann Neurol. 40(3):440-5; Upadhyaya *et al.*, 1996, Am J Med Genet. 67(4):421-3;
- 15 Robinson *et al.*, 1995, Hum Genet. 96(1):95-8.; Legius *et al.*, 1995, J Med Genet. 32(4):316-9; von Deimling *et al.*, 1995, Brain Pathol. 5(1):11-4; Dublin *et al.*, 1995, Hum Mutat. 5(1):81-5; Legius *et al.*, 1994, Genes Chromosomes Cancer. 10(4):250-5; Purandare *et al.*, 1994, Hum Mol Genet. 3(7):1109-15; Shen & Upadhyaya, 1993, Hum Genet. 92(4):410-2; and Estivill *et al.*, 1991, Hum Genet. 88(2):185-8; the disclosures of which are hereby incorporated by reference in their entireties). Any gene encoding a premature translation codon implicated in neurofibromatosis including, but not limited to, the nonsense mutations described in the references cited above, can be used in the present invention to identify compounds that mediate premature translation termination and/or nonsense-mediated mRNA decay.
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25 5.5.7. Retinoblastoma

The retinoblastoma gene plays important roles in the genesis of human cancers. Several pieces of evidence have shown that the retinoblastoma protein has dual roles in gating cell cycle progression and promoting cellular differentiation (see, e.g., Lee & Lee, 1997, Gan To Kagaku Ryoho 24(11):1368-80 for a review). Nonsense mutations in

- 30 the RB1 gene have been described. Nonsense mutations have been implicated in retinoblastoma (see, e.g., Klutz *et al.*, 2002, Am J Hum Genet. 71(1):174-9; Alonso *et al.*, 2001, Hum Mutat. 17(5):412-22; Wong *et al.*, 2000, Cancer Res. 60(21):6171-7; Harbour , 1998, Ophthalmology 105(8):1442-7; Fulop *et al.*, 1998, J Reprod Med. 43(2):119-27; Onadim *et al.*, 1997, Br J Cancer 76(11):1405-9; Lohmann *et al.*, 1997, Ophthalmologe

94(4):263-7; Cowell & Cragg, 1996, Eur J Cancer. 32A(10):1749-52; Lohmann et al., 1996,
Am J Hum Genet. 58(5):940-9; Shapiro et al., 1995, Cancer Res. 55(24):6200-9; Huang et
al., 1993, Cancer Res. 53(8):1889-94; and Cheng & Haas, 1990, Mol Cell Biol.
10(10):5502-9; the disclosures of which are hereby incorporated by reference in their
5 entireties). Any gene encoding a premature translation codon implicated in retinoblastoma
including, but not limited to, the nonsense mutations described in the references cited
above, can be used in the present invention to identify compounds that mediate premature
translation termination and/or nonsense-mediated mRNA decay.

5.5.8. Wilm's Tumor

10 Wilm's tumor, or nephroblastoma, is an embryonal malignancy of the kidney
that affects children. Nonsense mutations in the WT1 gene have been implicated in Wilm's
tumor. Several nonsense mutations in the WT1 have been identified (see, e.g., Nakadate et
al., 1999, Genes Chromosomes Cancer 25(1):26-32; Diller et al., 1998, J Clin Oncol.
16(11):3634-40; Schumacher et al., 1997, Proc Natl Acad Sci U S A. 94(8):3972-7; Coppes
15 et al., 1993, Proc Natl Acad Sci U S A. 90(4):1416-9; and Little et al., 1992, Proc Natl
Acad Sci U S A. 89(11):4791-5; the disclosures of which are hereby incorporated by
reference in their entireties). Any WT1 gene encoding a premature translation codon
including, but not limited to, the nonsense mutations described in the references cited
above, can be used in the present invention to identify compounds that mediate premature
20 translation termination and/or nonsense-mediated mRNA decay.

5.5.9. Retinitis Pigmentosa

Retinitis pigmentosa is a genetic disease in which affected individuals
develop progressive degeneration of the rod and cone photoreceptors. Retinitis pigmentosa
cannot be explained by a single genetic defect but instead the hereditary aberration
25 responsible for triggering the onset of the disease is localized in different genes and at
different sites within these genes (reviewed in, e.g., Kohler et al., 1997, Klin Monatsbl
Augenheilkd 211(2):84-93). Nonsense mutations have been implicated in retinitis
pigmentosa (see, e.g., Ching et al., 2002, Neurology 58(11):1673-4; Zhang et al., 2002,
Zhonghua Yi Xue Yi Chuan Xue Za Zhi. 19(3):194-7; Zhang et al., 2002, Hum Mol Genet.
30 1;11(9):993-1003; Dietrich et al., 2002, Br J Ophthalmol. 86(3):328-32; Grayson et al.,
2002, J Med Genet. 39(1):62-7; Liu et al., 2001, Zhonghua Yi Xue Za Zhi 81(2):71-2;
Damji et al., 2001, Can J Ophthalmol. 36(5):252-9; Berson et al., 2001, Invest Ophthalmol
Vis Sci. 42(10):2217-24; Chan et al., 2001, Br J Ophthalmol. 85(9):1046-8; Baum et al.,

2001, *Hum Mutat.* 17(5):436; Mashima *et al.*, 2001, *Ophthalmic Genet.* 22(1):43-7;
Zwaenepoel *et al.*, 2001, *Hum Mutat.* 2001;17(1):34-41; Bork *et al.*, 2001, *Am J Hum Genet.* 68(1):26-37; Sharon *et al.*, 2000, *Invest Ophthalmol Vis Sci.* 41(9):2712-21; Dreyer *et al.*, 2000, *Eur J Hum Genet.* 8(7):500-6; Liu *et al.*, 2000, *Hum Mutat.* 15(6):584; Wang *et al.*, 1999, *Exp Eye Res.* 69(4):; Bowne *et al.*, 1999, *Hum Mol Genet.* 8(11):2121-8;
Guillonneau *et al.*, 1999, *Hum Mol Genet.* 8(8):1541-6; Dryja *et al.*, 1999, *Invest Ophthalmol Vis Sci.* 40(8):1859-65; Sullivan *et al.*, 1999, *Nat Genet.* 22(3):255-9; Pierce *et al.*, 1999, *Nat Genet.* 22(3):248-54; Janecke *et al.*, 1999, *Hum Mutat.* 13(2):133-40; Cuevas *et al.*, 1998, *Mol Cell Probes* 12(6):417-20; Schwahn *et al.*, 1998, *Nat Genet.* 19(4):327-32;
10 Buraczynska *et al.*, 1997, *Am J Hum Genet.* 61(6):1287-92; Meindl *et al.*, 1996, *Nat Genet.* 13(1):35-42; Keen *et al.*, 1996, *Hum Mutat.* 8(4):297-303; Dryja *et al.*, 1995, *Proc Natl Acad Sci U S A.* 92(22):10177-81; Apfelstedt-Sylla *et al.*, 1995, *Br J Ophthalmol.* 79(1):28-34; Bayes *et al.*, 1995, *Hum Mutat.* 5(3):228-34; Shastry, 1994, *Am J Med Genet.* 52(4):467-74; Gal *et al.*, 1994, *Nat Genet.* 7(1):64-8; Sargan *et al.*, 1994, *Gene Ther.* 1
15 Suppl 1:S89; McLaughlin *et al.*, 1993, *Nat Genet.* 4(2):130-4; Rosenfeld *et al.*, 1992, *Nat Genet.* 1(3):209-13; the disclosures of which are hereby incorporated by reference in their entireties). Any gene encoding a premature translation codon implicated in retinitis pigmentosa including, but not limited to, the nonsense mutations described in the references cited above, can be used in the present invention to identify compounds that mediate
20 premature translation termination and/or nonsense-mediated mRNA decay.

5.5.10. Osteogenesis Imperfecta

Osteogenesis imperfecta is a heterogeneous disorder of type I collagen resulting in varying degrees of severity and results from mutations the genes that encode the proalpha chains of type I collagen. Nonsense mutations have been implicated in the genes
25 that encode the proalpha chains of type I collagen (“COLA1” genes) (see, e.g., Slayton *et al.*, 2000, *Matrix Biol.* 19(1):1-9; Bateman *et al.*, 1999, *Hum Mutat.* 13(4):311-7; and Willing *et al.*, 1996, *Am J Hum Genet.* 59(4):799-809; the disclosures of which are hereby incorporated by reference in their entireties). Any COLA1 gene encoding a premature translation codon including, but not limited to, the nonsense mutations described in the
30 references cited above, can be used in the present invention to identify compounds that mediate premature translation termination and/or nonsense-mediated mRNA decay.

5.5.11. Cirrhosis

Cirrhosis generally refers to a chronic liver disease that is marked by replacement of normal tissue with fibrous tissue. The multidrug resistance 3 gene has been implicated in cirrhosis, and nonsense mutations have been identified in this gene (see, e.g., Jacquemin *et al.*, 2001, *Gastroenterology*. 2001 May;120(6):1448-58; the disclosure of which is hereby incorporated by reference in its entirety). Any gene involved in cirrhosis encoding a premature translation codon including, but not limited to, the nonsense mutations described in the reference cited above, can be used in the present invention to identify compounds that mediate premature translation termination and/or nonsense-mediated mRNA decay.

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5.5.12. Tay Sachs Disease

Tay Sachs disease is an autosomal recessive disorder affecting the central nervous system. The disorder results from mutations in the gene encoding the alpha-subunit of beta-hexosaminidase A, a lysosomal enzyme composed of alpha and beta polypeptides. Several nonsense mutations have been implicated in Tay Sachs disease (see, e.g., Rajavel & Neufeld, 2001, *Mol Cell Biol*. 21(16):5512-9; Myerowitz, 1997, *Hum Mutat*. 9(3):195-208; Akli *et al.*, 1993, *Hum Genet*. 90(6):614-20; Mules *et al.*, 1992, *Am J Hum Genet*. 50(4):834-41; and Akli *et al.*, 1991, *Genomics*. 11(1):124-34; the disclosures of which are hereby incorporated by reference in their entireties). Any hexosaminidase gene encoding a premature translation codon including, but not limited to, the nonsense mutations described in the references cited above, can be used in the present invention to identify compounds that mediate premature translation termination and/or nonsense-mediated mRNA decay.

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5.5.13. Blood Disorders

Hemophilia is caused by a deficiency in blood coagulation factors. Affected individuals are at risk for spontaneous bleeding into organs and treatment usually consists of administration of clotting factors. Hemophilia A is caused by a deficiency of blood coagulation factor VIII and hemophilia B is caused by a deficiency in blood coagulation factor IX. Nonsense mutations in the genes encoding coagulation factors have been implicated in hemophilia (see, e.g., Dansako *et al.*, 2001, *Ann Hematol*. 80(5):292-4; Moller-Morlang *et al.*, 1999, *Hum Mutat*. 13(6):504; Kamiya *et al.*, 1998, *Rinsho Ketsueki* 39(5):402-4; Freson *et al.*, 1998, *Hum Mutat*. 11(6):470-9; Kamiya *et al.*, 1995, *Int J Hematol*. 62(3):175-81; Walter *et al.*, 1994, *Thromb Haemost*. 72(1):74-7; Figueiredo, 1993, *Braz J Med Biol Res*. 26(9):919-31; Reiner & Thompson, 1992, *Hum Genet*. 89(1):88-94; Koeberl *et al.*, 1990, *Hum Genet*. 84(5):387-90; Driscoll *et al.*, 1989, *Blood*.

74(2):737-42; Chen *et al.*, 1989, Am J Hum Genet. 44(4):567-9; Mikami *et al.*, 1988, Jinrui Idengaku Zasshi. 33(4):409-15; Gitschier *et al.*, 1988, Blood 72(3):1022-8; and Sommer *et al.*, 1987, Mayo Clin Proc. 62(5):387-404; the disclosures of which are hereby incorporated by reference in their entireties). Any gene encoding a premature translation codon

5 implicated in hemophilia including, but not limited to, the nonsense mutations described in the references cited above, can be used in the present invention to identify compounds that mediate premature translation termination and/or nonsense-mediated mRNA decay.

Von Willebrand disease is a single-locus disorder resulting from a deficiency of von Willebrand factor: a multimeric multifunctional protein involved in platelet adhesion and platelet-to-platelet cohesion in high shear stress vessels, and in protecting from proteolysis and directing circulating factor VIII to the site of injury (reviewed in Rodeghiero, 2002, Haemophilia. 8(3):292-300). Nonsense mutations have implicated in von Willebrand disease (see, e.g., Rodeghiero, 2002, Haemophilia. 8(3):292-300; Enayat *et al.*, 2001, Blood 98(3):674-80; Surdhar *et al.*, 2001, Blood 98(1):248-50; Casana *et al.*,

15 2000, Br J Haematol. 111(2):552-5; Baronciani *et al.*, 2000, Thromb Haemost.

84(4):536-40; Fellowes *et al.*, 2000, Blood 96(2):773-5; Waseem *et al.*, 1999, Thromb Haemost. 81(6):900-5; Mohlke *et al.*, 1999, Int J Clin Lab Res. 29(1):1-7; Rieger *et al.*, 1998, Thromb Haemost. 80(2):332-7; Kenny *et al.*, 1998, Blood 92(1):175-83; Mazurier *et al.*, 1998, Ann Genet. 41(1):34-43; Hagiwara *et al.*, 1996, Thromb Haemost. 76(2):253-7;

20 Mazurier & Meyer, 1996, Baillieres Clin Haematol. 9(2):229-41; Schneppenheim *et al.*, 1994, Hum Genet. 94(6):640-52; Zhang *et al.*, 1994, Genomics 21(1):188-93; Ginsburg & Sadler, 1993, Thromb Haemost. 69(2):177-84; Eikenboom *et al.*, 1992, Thromb Haemost. 68(4):448-54; Zhang *et al.*, 1992, Am J Hum Genet. 51(4):850-8; Zhang *et al.*, 1992, Hum Mol Genet. 1(1):61-2; and Mancuso *et al.*, 1991, Biochemistry 30(1):253-69; the

25 disclosures of which are hereby incorporated by reference in their entireties). Any gene encoding a premature translation codon implicated in von Willebrand disease including, but not limited to, the nonsense mutations described in the references cited above, can be used in the present invention to identify compounds that mediate premature translation termination and/or nonsense-mediated mRNA decay.

30 β thalassemia is caused by a deficiency in beta globin polypeptides which in turn causes a deficiency in hemoglobin production. Nonsense mutations have been implicated in b thalassemia (see, e.g., El-Latif *et al.*, 2002, Hemoglobin 26(1):33-40; Sanguansermsri *et al.*, 2001, Hemoglobin 25(1):19-27; Romao 2000, Blood 96(8):2895-901; Perea *et al.*, 1999, Hemoglobin 23(3):231-7; Rhodes *et al.*, 1999, Am J

Med Sci. 317(5):341-5; Fonseca *et al.*, 1998, Hemoglobin 22(3):197-207; Gasperini *et al.*, 1998, Am J Hematol. 1998 Jan;57(1):43-7; Galanello *et al.*, 1997, Br J Haematol. 99(2):433-6; Pistidda *et al.*, 1997, Eur J Haematol. 58(5):320-5; Oner *et al.*, 1997, Br J Haematol. 96(2):229-34; Yasunaga *et al.*, 1995, Intern Med. 34(12):1198-200; Molina *et al.*, 1994, Sangre (Barc) 39(4):253-6; Chang *et al.*, 1994, Int J Hematol. 59(4):267-72; Gilman *et al.*, 1994, Am J Hematol. 45(3):265-7; Chan *et al.*, 1993, Prenat Diagn. 13(10):977-82; George *et al.*, 1993, Med J Malaysia 48(3):325-9; Divoky *et al.*, 1993, Br J Haematol. 83(3):523-4; Fioretti *et al.*, 1993, Hemoglobin 17(1):9-17; Rosatelli *et al.*, 1992, Am J Hum Genet. 50(2):422-6; Moi *et al.*, 1992, Blood 79(2):512-6; Loudianos *et al.*, 1992, Hemoglobin 16(6):503-9; Fukumaki, 1991, Rinsho Ketsueki 32(6):587-91; Cao *et al.*, 1991, Am J Pediatr Hematol Oncol. 13(2):179-88; Galanello *et al.*, 1990, Clin Genet. 38(5):327-31; Liu, 1990, Zhongguo Yi Xue Ke Xue Yuan Xue Bao 12(2):90-5; Aulehla-Scholz *et al.*, 1990, Hum Genet. 84(2):195-7; Cao *et al.*, 1990, Ann N Y Acad Sci. 612:215-25; Sanguansermsri *et al.*, 1990, Hemoglobin 14(2):157-68; Galanello *et al.*, 1989, Blood 74(2):823-7; Rosatelli *et al.*, 1989, Blood 73(2):601-5; Galanello *et al.*, 1989, Prog Clin Biol Res. 316B:113-21; Galanello *et al.*, 1988, Am J Hematol. 29(2):63-6; Chan *et al.*, 1988, Blood 72(4):1420-3; Atweh *et al.*, 1988, J Clin Invest. 82(2):557-61; Masala *et al.*, 1988, Hemoglobin 12(5-6):661-71; Pirastu *et al.*, 1987, Proc Natl Acad Sci U S A 84(9):2882-5; Kazazian *et al.*, 1986, Am J Hum Genet. 38(6):860-7; Cao *et al.*, 1986, Prenat Diagn. 6(3):159-67; Cao *et al.*, 1985, Ann N Y Acad Sci. 1985;445:380-92; Pirastu *et al.*, 1984, Science 223(4639):929-30; Pirastu *et al.*, 1983, N Engl J Med. 309(5):284-7; Trecartin *et al.*, 1981, J Clin Invest. 68(4):1012-7; and Liebhaber *et al.*, 1981, Trans Assoc Am Physicians 94:88-96; the disclosures of which are hereby incorporated by reference in their entireties). Any gene encoding a premature translation codon implicated in b thalassemia including, but not limited to, the nonsense mutations described in the references cited above, can be used in the present invention to identify compounds that mediate premature translation termination and/or nonsense-mediated mRNA decay.

5.5.14. Kidney Stones

Kidney stones (nephrolithiasis), which affect 12% of males and 5% of females in the western world, are familial in 45% of patients and are most commonly associated with hypercalciuria (see, e.g., Lloyd *et al.*, Nature 1996 Feb 1;379(6564):445-9). Mutations of the renal-specific chloride channel gene are associated with hypercalciuric nephrolithiasis (kidney stones). Nonsense mutations have been implicated in kidney stones (see, e.g., Hoopes *et al.*, 1998, Kidney Int. 54(3):698-705; Lloyd *et al.*, 1997, Hum Mol - 74 -

Genet. 6(8):1233-9; Lloyd *et al.*, 1996, Nature 379(6564):445-9; and Pras *et al.*, 1995, Am J Hum Genet. 56(6):1297-303; the disclosures of which are hereby incorporated by reference in their entireties). Any gene encoding a premature translation codon implicated in kidney stones including, but not limited to, the nonsense mutations described in the references cited above, can be used in the present invention to identify compounds that mediate premature translation termination and/or nonsense-mediated mRNA decay.

5.5.15. Ataxia-Telangiectasia

Ataxia-telangiectasia is characterized by increased sensitivity to ionizing radiation, increased incidence of cancer, and neurodegeneration and is generally caused by mutations in the ataxia-telangiectasia gene (see, e.g., Barlow *et al.*, 1999, Proc Natl Acad Sci USA 96(17):9915-9). Nonsense mutations have been implicated in ataxia-telangiectasia (see, e.g., Camacho *et al.*, 2002, Blood 99(1):238-44; Pitts *et al.*, 2001, Hum Mol Genet. 10(11):1155-62; Laake *et al.*, 2000, Hum Mutat. 16(3):232-46; Li & Swift, 2000, Am J Med Genet. 92(3):170-7; Teraoka *et al.*, 1999, Am J Hum Genet. 64(6):1617-31; and Stoppa-Lyonnet *et al.*, 1998, Blood 91(10):3920-6; the disclosures of which are hereby incorporated by reference in their entireties). Any gene encoding a premature translation codon implicated in ataxia-telangiectasia including, but not limited to, the nonsense mutations described in the references cited above, can be used in the present invention to identify compounds that mediate premature translation termination and/or nonsense-mediated mRNA decay.

5.5.16. Lysosomal Storage Diseases

There are more than 40 individually recognized lysosomal storage disorders. Each disorder results from a deficiency in the activity of a specific enzyme, which impedes the lysosome from carrying out its normal degradative role. These include but are not limited to the diseases listed subsequently. Aspartylglucosaminuria is caused by a deficiency of N-aspartyl-beta-glucosaminidase (Fisher *et al.*, 1990, FEBS Lett. 269:440-444); cholesterol ester storage disease (Wolman disease) is caused by mutations in the LIPA gene (Fujiyama *et al.*, 1996, Hum. Mutat. 8:377-380); mutations in the CTNS gene are associated with cystinosis (Town *et al.*, 1998, Nature Genet. 18:319-324); mutations in a galactosidase A are associated with Fabry disease (Eng *et al.*, 1993, Pediat. Res. 33:128A; Sakuraba *et al.*, 1990, Am. J. Hum. Genet. 47:784-789; Davies *et al.*, 1993, Hum. Molec. Genet. 2:1051-1053; Miyamura *et al.*, 1996, J. Clin. Invest. 98:1809-1817); fucosidosis is caused by mutations in the FUCA1 gene (Kretz *et al.*, 1989, J. Molec. Neurosci. 1:177-180;

Yang et al., 1992, Biochem. Biophys. Res. Commun. 189:1063-1068; Seo et al., 1993, Hum. Molec. Genet. 2:1205-1208); mucolipidosis type I results from mutations in the NEU1 gene (Bonten et al., 1996, Genes Dev. 10:3156-3169); mucolipidosis type IV results from mutations in the MCOLN1 gene (Bargal et al., 2000, Nature Genet. 26:120-123; Sun et al., 2000, Hum. Molec. Genet. 9:2471-2478); Mucopolysaccharidosis type I (Hurler syndrome) is caused by mutations in the IDUA gene (Scott et al., 1992, Genomics 13:1311-1313; Bach et al., 1993, Am. J. Hum. Genet. 53:330-338); Mucopolysaccharidosis type II (Hunter syndrome) is caused by mutations in the IDS gene (Sukegawa et al., 1992, Biochem. Biophys. Res. Commun. 183:809-813; Bunge et al., 1992, Hum. Molec. Genet. 1:335-339; Flomen et al., 1992, Genomics 13:543-550); mucopolysaccharidosis type 25IIIB (Sanfilippo syndrome type A) is caused by mutations in the SGSH gene (Yogalingam et al., 2001, Hum. Mutat. 18:264-281); mucopolysaccharidosis type IIIB (Sanfilippo syndrome) is caused by mutations in the NAGLU gene (Zhao et al., 1996, Proc. Nat. Acad. Sci. 93:6101-6105; Zhao et al., 1995, Am. J. Hum. Genet. 57:A185); mucopolysaccharidosis type IIID is caused by mutations in the glucosamine-6-sulfatase (G6S) gene (Robertson et al., 1988, Hum. Genet. 79:175-178); mucopolysaccharidosis type IVA (Morquio syndrome) is caused by mutations in the GALNS gene (Tomatsu et al., 1995, Am. J. Hum. Genet. 57:556-563; Tomatsu et al., 1995, Hum. Mutat. 6:195-196); mucopolysaccharidosis type VI (Maroteaux-Lamy syndrome) is caused by mutations in the ARSB gene (Litjens et al., 1992, Hum. Mutat. 1:397-402; Isbrandt et al., 1996, Hum. Mutat. 7:361-363); mucopolysaccharidosis type VII (Sly syndrome) is caused by mutations in the beta-glucuronidase (GUSB) gene (Yamada et al., 1995, Hum. Molec. Genet. 4:651-655); mutations in CLN1 (PPT1) cause infantile neuronal ceroid lipofuscinosis (Das et al., 1998, J. Clin. Invest. 102:361-370; Mitchison et al., 1998, Hum. Molec. Genet. 7:291-297); late infantile type ceroid lipofuscinosis is caused by mutations in the CLN2 gene (Sleat et al., 1997, Science 277:1802-1805); juvenile neuronal ceroid lipofuscinosis (Batten disease) is caused by mutations in the CLN3 gene (Mole et al., 1999, Hum. Mutat. 14: 199-215); late infantile neuronal ceroid lipofuscinosis, Finnish variant, is caused by mutations in the CLN5 gene (Savukoski et al., 1998, Nature Genet. 19:286-288); late-infantile form of neuronal ceroid lipofuscinosis is caused by mutations in the CLN6 gene (Gao et al., 2002, Am. J. Hum. Genet. 70:324-335); Niemann-Pick disease is caused by mutations in the ASM gene (Takahashi et al., 1992, J. Biol. Chem. 267:12552-12558; types A and B) and the NPC1 gene (Millat et al., 2001, Am. J. Hum. Genet. 68:1373-1385; type C); Kanzaki disease is caused by mutations in the NAGA gene (Keulemans et al., 1996, J. Med. Genet. 33:458-464); Gaucher disease is caused by mutations in the GBA gene (Stone, et al., 1999, Europ.

J. Hum. Genet. 7:505-509); Glycogen storage disease II is the prototypic lysosomal storage disease and is caused by mutations in the GAA gene(Becker et al., 1998, Am. J. Hum. Genet. 62:991-994); Krabbe disease is caused by mutations in the GALC gene (Sakai et al., 1994, Biochem. Biophys. Res. Commun. 198:485-491); Tay-Sachs disease is caused by mutations in the HEXA gene (Akli et al., 1991, Genomics 11:124-134; Mules et al., 1992, Am. J. Hum. Genet. 50: 834-841;Triggs-Raine et al., 1991, Am. J. Hum. Genet. 49:1041-1054; Drucker et al., 1993, Hum. Mutat. 2:415-417; Shore et al., 1992, Hum. Mutat. 1:486-490); mutations in the GM2Agene causes Tay-Sachs variant AB (Schepers et al., 1996, Am. J. Hum. Genet. 59:1048-1056; Chen et al., 1999, Am. J. Hum. Genet. 65:77-87); mutations in the HEXB gene cause Sandhoff disease (Zhang et al., 1994, Hum Mol Genet 3:139-145); alphamannosidosis type II is caused by mutations in the MAN2B1 gene (Gotoda et al., 1998, Am. J. Hum. Genet. 63:1015-1024; Autio et al., 1973, Acta Paediat. Scand. 62:555-565); metachromatic leukodystrophy is caused by mutations in the ARSA gene(Gieselmann et al., 1994, Hum. Mutat. 4:233-242). Any gene containing a premature translation codon implicated in lysosomal storage disease disorders including, but not limited to, the nonsense mutations and genes described in the references cited above, can be used in the present invention to identify compounds that mediate premature translation termination and/or nonsense-mediated mRNA decay.

5.5.17. Tuberous Sclerosis

Tuberous sclerosis complex (TSC) is a dominantly inherited disease characterized by the presence of hamartomata in multiple organ systems. The disease is caused by mutations in TSC1 (van Slegtenhorst *et al.*, 1997 Science 277:805-808; Sato *et al.*, 2002, J. Hum. Genet. 47:20-28) and/or TSC2 (Vrtel *et al.*, 1996, J. Med. Genet. 33:47-51; Wilson *et al.*, 1996, Hum. Molec. Genet. 5:249-256; Au *et al.*, 1998, Am. J.Hum. Genet. 62:286-294; Verhoef *et al.*, 1999, Europ. J. Pediat. 158:284-287;Carsillo *et al.*, 2000, Proc. Nat. Acad. Sci. 97:6085-6090). Any gene containing a premature translation codon implicated in tuberous sclerosis including, but not limited to, the nonsense mutations described in the references cited above, can be used in the present invention to identify compounds that mediate premature translation termination and/or nonsense-mediated mRNA decay.

5.6. Secondary Biological Screens or Assays

5.6.1. *In vitro* Assays

The compounds identified in the assays described supra (for convenience referred to herein as a "lead" compound) can be tested for biological activity using host cells containing or engineered to contain a gene of interest with a premature stop codon or nonsense mutation coupled to a functional readout system. For example, a phenotypic or physiological readout can be used to assess the premature translation termination and/or nonsense-mediated mRNA decay of the RNA product encoded by the gene of interest in the presence and absence of the lead compound.

In one embodiment, a phenotypic or physiological readout can be used to assess the premature translation termination and/or nonsense-mediated mRNA decay of an RNA product of interest in the presence and absence of the lead compound. In accordance with this embodiment, cell-based and cell-free assays described herein, or in International Publication No. WO 01/44516 (which is incorporated herein by reference in its entirety) may be used to assess the premature translation termination and/or nonsense-mediated mRNA decay of the RNA product of interest. Where the gene product of interest is involved in cell growth or viability, the *in vivo* effect of the lead compound can be assayed by measuring the cell growth or viability of the target cell. Such assays can be carried out with representative cells of cell types involved in a particular disease or disorder (e.g., leukocytes such as T cells, B cells, natural killer cells, macrophages, neutrophils and eosinophils). A lower level of proliferation or survival of the contacted cells indicates that the lead compound is effective to treat a condition in the patient characterized by uncontrolled cell growth. Alternatively, instead of culturing cells from a patient, a lead compound may be screened using cells of a tumor or malignant cell line or an endothelial cell line. Specific examples of cell culture models include, but are not limited to, for lung cancer, primary rat lung tumor cells (see, e.g., Swafford et al., 1997, Mol. Cell. Biol., 17:1366-1374) and large-cell undifferentiated cancer cell lines (see, e.g., Mabry et al., 1991, Cancer Cells, 3:53-58); colorectal cell lines for colon cancer (see, e.g., Park & Gazdar, 1996, J. Cell Biochem. Suppl. 24:131-141); multiple established cell lines for breast cancer (see, e.g., Hambly et al., 1997, Breast Cancer Res. Treat. 43:247-258; Gierthy et al., 1997, Chemosphere 34:1495-1505; and Prasad & Church, 1997, Biochem. Biophys. Res. Commun. 232:14-19); a number of well-characterized cell models for prostate cancer (see, e.g., Webber et al., 1996, Prostate, Part 1, 29:386-394; Part 2, 30:58-64; and Part 3, 30:136-142 and Boulikas, 1997, Anticancer Res. 17:1471-1505); for genitourinary cancers, continuous human bladder cancer cell lines (see, e.g., Ribeiro et al., 1997, Int. J. Radiat. Biol. 72:11-20); organ cultures of transitional cell carcinomas (see, e.g., Booth et al., 1997,

Lab Invest. 76:843-857) and rat progression models (see, e.g., Vet et al., 1997, Biochim. Biophys Acta 1360:39-44); and established cell lines for leukemias and lymphomas (see, e.g., Drexler, 1994, Leuk. Res. 18:919-927 and Tohyama, 1997, Int. J. Hematol. 65:309-317).

5 Many assays well-known in the art can be used to assess the survival and/or growth of a patient cell or cell line following exposure to a lead compound; for example, cell proliferation can be assayed by measuring bromodeoxyuridine (BrdU) incorporation (see, e.g., Hoshino et al., 1986, Int. J. Cancer 38:369 and Campana et al., 1988, J. Immunol. Meth. 107:79) or (3H)-thymidine incorporation (see, e.g., Chen, 1996, Oncogene 13:1395-10 403 and Jeoung, 1995, J. Biol. Chem. 270:18367-73), by direct cell count, by detecting changes in transcription, translation or activity of known genes such as proto-oncogenes (e.g., fos, myc) or cell cycle markers (Rb, cdc2, cyclin A, D1, D2, D3, E, etc.). The levels 15 of such protein and mRNA and activity can be determined by any method well known in the art. For example, protein can be quantitated by known immunodiagnostic methods such as western blotting or immunoprecipitation using commercially available antibodies. mRNA can be quantitated using methods that are well known and routine in the art, for example, using northern analysis, RNase protection, the polymerase chain reaction in connection with reverse transcription ("RT-PCR"). Cell viability can be assessed by using trypan-blue staining or other cell death or viability markers known in the art. In a specific embodiment, 20 the level of cellular ATP is measured to determine cell viability. Differentiation can be assessed, for example, visually based on changes in morphology.

The lead compound can also be assessed for its ability to inhibit cell transformation (or progression to malignant phenotype) in vitro. In this embodiment, cells with a transformed cell phenotype are contacted with a lead compound, and examined for 25 change in characteristics associated with a transformed phenotype (a set of in vitro characteristics associated with a tumorigenic ability in vivo), for example, but not limited to, colony formation in soft agar, a more rounded cell morphology, looser substratum attachment, loss of contact inhibition, loss of anchorage dependence, release of proteases such as plasminogen activator, increased sugar transport, decreased serum requirement, or 30 expression of fetal antigens, etc. (see, e.g., Luria et al., 1978, General Virology, 3d Ed., John Wiley & Sons, New York, pp. 436-446).

Loss of invasiveness or decreased adhesion can also be assessed to demonstrate the anti-cancer effects of a lead compound. For example, an aspect of the formation of a metastatic cancer is the ability of a precancerous or cancerous cell to detach

from primary site of disease and establish a novel colony of growth at a secondary site. The ability of a cell to invade peripheral sites reflects its potential for a cancerous state. Loss of invasiveness can be measured by a variety of techniques known in the art including, for example, induction of E-cadherin-mediated cell-cell adhesion. Such E-cadherin-mediated adhesion can result in phenotypic reversion and loss of invasiveness (see, e.g., Hordijk et al., 1997, Science 278:1464-66).

Loss of invasiveness can further be examined by inhibition of cell migration. A variety of 2-dimensional and 3-dimensional cellular matrices are commercially available (Calbiochem-Novabiochem Corp. San Diego, CA). Cell migration across or into a matrix can be examined using microscopy, time-lapsed photography or videography, or by any method in the art allowing measurement of cellular migration. In a related embodiment, loss of invasiveness is examined by response to hepatocyte growth factor ("HGF"). HGF-induced cell scattering is correlated with invasiveness of cells such as Madin-Darby canine kidney ("MDCK") cells. This assay identifies a cell population that has lost cell scattering activity in response to HGF (see, e.g., Hordijk et al., 1997, Science 278:1464-66).

Alternatively, loss of invasiveness can be measured by cell migration through a chemotaxis chamber (Neuroprobe/ Precision Biochemicals Inc. Vancouver, BC). In such assay, a chemo-attractant agent is incubated on one side of the chamber (e.g., the bottom chamber) and cells are plated on a filter separating the opposite side (e.g., the top chamber). In order for cells to pass from the top chamber to the bottom chamber, the cells must actively migrate through small pores in the filter. Checkerboard analysis of the number of cells that have migrated can then be correlated with invasiveness (see e.g., Ohnishi, 1993, Biochem. Biophys. Res. Commun. 193:518-25).

A lead compound can also be assessed for its ability to alter the expression of a secondary protein (as determined, e.g. by western blot analysis) or RNA, whose expression and/or activation is regulated directly or indirectly by the gene product of a gene of interest containing a premature stop codon or a nonsense mutation (as determined, e.g., by RT-PCR or northern blot analysis) in cultured cells *in vitro* using methods which are well known in the art. Further, chemical footprinting analysis can be conducted as described herein (see, e.g., Example 7) or also well-known in the art.

5.6.2. Animal Models

Animal model systems can be used to demonstrate the safety and efficacy of the lead compounds identified in the nonsense suppression assays described above. The

lead compounds identified in the nonsense suppression assay can then be tested for biological activity using animal models for a disease, condition, or syndrome of interest. These include animals engineered to contain the target RNA element coupled to a functional readout system, such as a transgenic mouse.

- 5 There are a number of methods that can be used to conduct animal model studies. Briefly, a compound identified in accordance with the methods of the invention is introduced into an animal model so that the effect of the compound on the manifestation of disease can be determined. The prevention or reduction in the severity, duration or onset of a symptom associated with the disease or disorder of the animal model that is associated
10 with, characterized by or caused by premature translation termination and/or nonsense mediated mRNA decay would indicate that the compound administered to the animal model had a prophylactic or therapeutic effect. Any method can be used to introduce the compound into the animal model, including, but not limited to, injection, intravenous infusion, oral ingestion, or inhalation. In a preferred embodiment, transgenic hosts are
15 constructed so that the animal genome encodes a gene of interest with a premature translation termination sequence or stop codon. In such an embodiment, the gene, containing a premature translation termination sequence or stop codon, would not encode a full length peptide from a transcribed mRNA. The administration of a compound to the animal model, and the expression of a full length protein, polypeptide or peptide, for example,
20 corresponding to the gene containing a premature stop codon would indicate that the compound modulates premature translation termination. Any method known in the art, or described herein, can be used to determine if the stop codon was modulated by the compound. In another embodiment, an animal is transfected with a reporter construct comprising a regulatory element operably linked to a reporter gene so that the expression
25 the reporter gene is regulated by a regulatory protein or subunit thereof encoded by a nucleic acid sequence that contains a premature translation termination sequence or stop codon suppression. In such an embodiment, the animal can be cotransfected with a recombinant vector comprising the nucleic acid sequence encoding the regulatory protein with a premature stop codon. In another embodiment, the animal host genome encodes a
30 native gene containing a premature stop codon. In yet another embodiment of the invention, the animal host is a natural mutant, *i.e.*, natively encoding a gene with a premature stop codon. For example, the animal can be a model for cystic fibrosis wherein the animal genome contains a natural mutation that incorporates a premature stop codon or translation termination sequence.

Examples of animal models for cystic fibrosis include, but are not limited to, cftr(-/-) mice (see, e.g., Freedman *et al.*, 2001, *Gastroenterology* 121(4):950-7), cftr(tm1HGU/tm1HGU) mice (see, e.g., Bernhard *et al.*, 2001, *Exp Lung Res* 27(4):349-66), CFTR-deficient mice with defective cAMP-mediated Cl(-) conductance (see, e.g., Stotland *et al.*, 2000, *Pediatr Pulmonol* 30(5):413-24), C57BL/6-Cftr(m1UNC)/Cftr(m1UNC) knockout mice (see, e.g., Stotland *et al.*, 2000, *Pediatr Pulmonol* 30(5):413-24), an animal model of the human airway, using bronchial xenografts grafted on rat tracheas and implanted into nude mice (see, e.g., Engelhardt *et al.*, 1992, *J. Clin. Invest.* 90: 2598-2607), a transgenic mouse model of cystic fibrosis (see, e.g., Clarke *et al.*, 1992, *Science* 257: 1125-1128; Colledge *et al.*, 1992, *Lancet* 340: 680 only; Dorin *et al.*, 1992, *Nature* 359: 211-215; Snouwaert *et al.*, 1992, *Science* 257: 1083-1088; Manson *et al.*, 1997, *EMBO J.* 16: 4238-4249).

Examples of animal models for muscular dystrophy include, but are not limited to, mouse, hamster, cat, dog, and *C. elegans*. Examples of mouse models for muscular dystrophy include, but are not limited to, the dy-/- mouse (see, e.g., Connolly *et al.*, 2002, *J Neuroimmunol* 127(1-2):80-7), a muscular dystrophy with myositis (mdm) mouse mutation (see, e.g., Garvey *et al.*, 2002, *Genomics* 79(2):146-9), the mdx mouse (see, e.g., Nakamura *et al.*, 2001, *Neuromuscul Disord* 11(3):251-9), the utrophin-dystrophin knockout (dko) mouse (see, e.g., Nakamura *et al.*, 2001, *Neuromuscul Disord* 11(3):251-9), the dy/dy mouse (see, e.g., Dubowitz *et al.*, 2000, *Neuromuscul Disord* 10(4-5):292-8), the mdx(Cv3) mouse model (see, e.g., Pillers *et al.*, 1999, *Laryngoscope* 109(8):1310-2), and the myotonic ADR-MDX mutant mice (see, e.g., Kramer *et al.*, 1998, *Neuromuscul Disord* 8(8):542-50). Examples of hamster models for muscular dystrophy include, but are not limited to, sarcoglycan-deficient hamsters (see, e.g., Nakamura *et al.*, 2001, *Am J Physiol Cell Physiol* 281(2):C690-9) and the BIO 14.6 dystrophic hamster (see, e.g., Schlenker & Burbach, 1991, *J Appl Physiol* 71(5):1655-62). An example of a feline model for muscular dystrophy includes, but is not limited to, the hypertrophic feline muscular dystrophy model (see, e.g., Gaschen & Burgunder, 2001, *Acta Neuropathol (Berl)* 101(6):591-600). Canine models for muscular dystrophy include, but are not limited to, golden retriever muscular dystrophy (see, e.g., Fletcher *et al.*, 2001, *Neuromuscul Disord* 11(3):239-43) and canine X-linked muscular dystrophy (see, e.g., Valentine *et al.*, 1992, *Am J Med Genet* 42(3):352-6). Examples of *C. elegans* models for muscular dystrophy are described in Chamberlain & Benian, 2000, *Curr Biol* 10(21):R795-7 and Culette & Sattelle, 2000, *Hum Mol Genet* 9(6):869-77. Also, a mouse model for Duchenne type muscular

dystrophy has been used to show that treatment with anabolic steroids increases myofiber damage (see, e.g., Krahn *et al.*, 1994, *J. Neurol. Sci.* 125: 138-146). A feline model for Duchenne type muscular dystrophy has also been described (see, e.g., Winand *et al.*, 1994, 4: 433-445).

5 Examples of animal models for familial hypercholesterolemia include, but are not limited to, mice lacking functional LDL receptor genes (see, e.g., Aji *et al.*, 1997, *Circulation* 95(2):430-7), Yoshida rats (see, e.g., Fantappie *et al.*, 1992, *Life Sci* 50(24):1913-24), the JCR:LA-cp rat (see, e.g., Richardson *et al.*, 1998, *Atherosclerosis* 138(1):135-46), swine (see, e.g., Hasler-Rapacz *et al.*, 1998, *Am J Med Genet* 76(5):379-10 86), the Watanabe heritable hyperlipidaemic rabbit (see, e.g., Tsutsumi *et al.*, 2000, *Arzneimittelforschung* 50(2):118-21; Harsch *et al.*, 1998, *Br J Pharmacol* 124(2):227-82; and Tanaka *et al.*, 1995, *Atherosclerosis* 114(1):73-82); and a family of rhesus monkeys with hypercholesterolemia due to deficiency of the LDL receptor (see, e.g., Scanu *et al.*, 1988, *J. Lipid Res.* 29: 1671-1681).

15 An example of an animal model for human cancer in general includes, but is not limited to, spontaneously occurring tumors of companion animals (see, e.g., Vail & MacEwen, 2000, *Cancer Invest* 18(8):781-92). Examples of animal models for lung cancer include, but are not limited to, lung cancer animal models described by Zhang & Roth (1994, *In Vivo* 8(5):755-69) and a transgenic mouse model with disrupted p53 function
20 (see, e.g., Morris *et al.*, 1998, *J La State Med Soc* 150(4):179-85). An example of an animal model for breast cancer includes, but is not limited to, a transgenic mouse that overexpresses cyclin D1 (see, e.g., Hosokawa *et al.*, 2001, *Transgenic Res* 10(5):471-8). An example of an animal model for colon cancer includes, but is not limited to, a TCRbeta and p53 double knockout mouse (see, e.g., Kado *et al.*, 2001, *Cancer Res* 61(6):2395-8).
25 Examples of animal models for pancreatic cancer include, but are not limited to, a metastatic model of Panc02 murine pancreatic adenocarcinoma (see, e.g., Wang *et al.*, 2001, *Int J Pancreatol* 29(1):37-46) and nu-nu mice generated in subcutaneous pancreatic tumours (see, e.g., Ghaneh *et al.*, 2001, *Gene Ther* 8(3):199-208). Examples of animal models for non-Hodgkin's lymphoma include, but are not limited to, a severe combined
30 immunodeficiency ("SCID") mouse (see, e.g., Bryant *et al.*, 2000, *Lab Invest* 80(4):553-73) and an IgHmu-HOX11 transgenic mouse (see, e.g., Hough *et al.*, 1998, *Proc Natl Acad Sci USA* 95(23):13853-8). An example of an animal model for esophageal cancer includes, but is not limited to, a mouse transgenic for the human papillomavirus type 16 E7 oncogene (see, e.g., Herber *et al.*, 1996, *J Virol* 70(3):1873-81). Examples of animal models for

colorectal carcinomas include, but are not limited to, Apc mouse models (see, e.g., Fodde & Smits, 2001, Trends Mol Med 7(8):369-73 and Kuraguchi *et al.*, 2000, Oncogene 19(50):5755-63). An example of an animal model for neurofibromatosis includes, but is not limited to, mutant NF1 mice (see, e.g., Cichowski *et al.*, 1996, Semin Cancer Biol 7(5):291-8). Examples of animal models for retinoblastoma include, but are not limited to, transgenic mice that express the simian virus 40 T antigen in the retina (see, e.g., Howes *et al.*, 1994, Invest Ophthalmol Vis Sci 35(2):342-51 and Windle *et al.*, 1990, Nature 343(6259):665-9) and inbred rats (see, e.g., Nishida *et al.*, 1981, Curr Eye Res 1(1):53-5 and Kobayashi *et al.*, 1982, Acta Neuropathol (Berl) 57(2-3):203-8). Examples of animal models for Wilm's tumor include, but are not limited to, a WT1 knockout mouse (see, e.g., Scharnhorst *et al.*, 1997, Cell Growth Differ 8(2):133-43), a rat subline with a high incidence of nephroblastoma (see, e.g., Mesfin & Breech, 1996, Lab Anim Sci 46(3):321-6), and a Wistar/Furth rat with Wilms' tumor (see, e.g., Murphy *et al.*, 1987, Anticancer Res 7(4B):717-9).

Examples of animal models for retinitis pigmentosa include, but are not limited to, the Royal College of Surgeons ("RCS") rat (see, e.g., Vollrath *et al.*, 2001, Proc Natl Acad Sci USA 98(22):12584-9 and Hanitzsch *et al.*, 1998, Acta Anat (Basel) 162(2-3):119-26), a rhodopsin knockout mouse (see, e.g., Jaissle *et al.*, 2001, Invest Ophthalmol Vis Sci 42(2):506-13), Wag/Rij rats (see, e.g., Lai *et al.*, 1980, Am J Pathol 98(1):281-4).

Examples of animal models for cirrhosis include, but are not limited to, CCl₄-exposed rats (see, e.g., Kloehn *et al.*, 2001, Horm Metab Res 33(7):394-401) and rodent models instigated by bacterial cell components or colitis (see, e.g., Vierling, 2001, Best Pract Res Clin Gastroenterol 15(4):591-610).

Examples of animal models for hemophilia include, but are not limited to, rodent models for hemophilia A (see, e.g., Reipert *et al.*, 2000, Thromb Haemost 84(5):826-32; Jarvis *et al.*, 1996, Thromb Haemost 75(2):318-25; and Bi *et al.*, 1995, Nat Genet 10(1):119-21), canine models for hemophilia A (see, e.g., Gallo-Penn *et al.*, 1999, Hum Gene Ther 10(11):1791-802 and Connelly *et al.*, 1998, Blood 91(9):3273-81), murine models for hemophilia B (see, e.g., Snyder *et al.*, 1999, Nat Med 5(1):64-70; Wang *et al.*, 1997, Proc Natl Acad Sci USA 94(21):11563-6; and Fang *et al.*, 1996, Gene Ther 3(3):217-22), canine models for hemophilia B (see, e.g., Mount *et al.*, 2002, Blood 99(8):2670-6; Snyder *et al.*, 1999, Nat Med 5(1):64-70; Fang *et al.*, 1996, Gene Ther 3(3):217-22); and Kay *et al.*, 1994, Proc Natl Acad Sci USA 91(6):2353-7), and a rhesus macaque model for hemophilia B (see, e.g., Lozier *et al.*, 1999, Blood 93(6):1875-81).

Examples of animal models for von Willebrand disease include, but are not limited to, an inbred mouse strain RIIS/J (see, e.g., Nichols *et al.*, 1994, 83(11):3225-31 and Sweeney *et al.*, 1990, 76(11):2258-65), rats injected with botrocetin (see, e.g., Sanders *et al.*, 1988, Lab Invest 59(4):443-52), and porcine models for von Willebrand disease (see, 5 e.g., Nichols *et al.*, 1995, Proc Natl Acad Sci USA 92(7):2455-9; Johnson & Bowie, 1992, J Lab Clin Med 120(4):553-8); and Brinkhous *et al.*, 1991, Mayo Clin Proc 66(7):733-42).

Examples of animal models for b-thalassemia include, but are not limited to, murine models with mutations in globin genes (see, e.g., Lewis *et al.*, 1998, Blood 91(6):2152-6; Raja *et al.*, 1994, Br J Haematol 86(1):156-62; Popp *et al.*, 1985, 445:432-44; 10 Skow *et al.*, 1983, Cell 34(3):1043-52). Ciavatta and co-workers created a mouse model of beta-zero-thalassemia by targeted deletion of both adult beta-like globin genes, beta(maj) and beta(min), in mouse embryonic stem cells (see, e.g., Ciavatta *et al.*, 1995, Proc Natl Acad Sci U S A. Sep 26;92(20):9259-63).

Examples of animal models for kidney stones include, but are not limited to, 15 genetic hypercalciuric rats (see, e.g., Bushinsky *et al.*, 1999, Kidney Int 55(1):234-43 and Bushinsky *et al.*, 1995, Kidney Int 48(6):1705-13), chemically treated rats (see, e.g., Grases *et al.*, 1998, Scand J Urol Nephrol 32(4):261-5; Burgess *et al.*, 1995, Urol Res 23(4):239- 42; Kumar *et al.*, 1991, J Urol 146(5):1384-9; Okada *et al.*, 1985, Hinyokika Kiyo 31(4):565-77; and Bluestone *et al.*, 1975, Lab Invest 33(3):273-9), hyperoxaluric rats (see, 20 e.g., Jones *et al.*, 1991, J Urol 145(4):868-74), pigs with unilateral retrograde flexible nephroscopy (see, e.g., Seifmah *et al.*, 2001, 57(4):832-6), and rabbits with an obstructed upper urinary tract (see, e.g., Itatani *et al.*, 1979, Invest Urol 17(3):234-40).

Examples of animal models for ataxia-telangiectasia include, but are not limited to, murine models of ataxia-telangiectasia (see, e.g., Barlow *et al.*, 1999, Proc Natl 25 Acad Sci USA 96(17):9915-9 and Inoue *et al.*, 1986, Cancer Res 46(8):3979-82). A mouse model was generated for ataxia-telangiectasia using gene targeting to generate mice that did not express the Atm protein (see, e.g., Elson *et al.*, 1996, Proc. Nat. Acad. Sci. 93: 13084- 13089).

Examples of animal models for lysosomal storage diseases include, but are 30 not limited to, mouse models for mucopolysaccharidosis type VII (see, e.g., Brooks *et al.*, 2002, Proc Natl Acad Sci U S A. 99(9):6216-21; Monroy *et al.*, 2002, Bone 30(2):352-9; Vogler *et al.*, 2001, Pediatr Dev Pathol. 4(5):421-33; Vogler *et al.*, 2001, Pediatr Res. 49(3):342-8; and Wolfe *et al.*, 2000, Mol Ther. 2(6):552-6), a mouse model for metachromatic leukodystrophy (see, e.g., Matzner *et al.*, 2002, Gene Ther. 9(1):53-63), a

mouse model of Sandhoff disease (see, e.g., Sango *et al.*, 2002, *Neuropathol Appl Neurobiol.* 28(1):23-34), mouse models for mucopolysaccharidosis type III A (see, e.g., Bhattacharyya *et al.*, 2001, *Glycobiology* 11(1):99-10 and Bhaumik *et al.*, 1999, *Glycobiology* 9(12):1389-96.), arylsulfatase A (ASA)-deficient mice (see, e.g., D'Hooge *et al.*, 1999, *Brain Res.* 847(2):352-6 and D'Hooge *et al.*, 1999, *Neurosci Lett.* 273(2):93-6);
5 mice with an aspartylglucosaminuria mutation (see, e.g., Jalanko *et al.*, 1998, *Hum Mol Genet.* 7(2):265-72); feline models of mucopolysaccharidosis type VI (see, e.g., Crawley *et al.*, 1998, *J Clin Invest.* 101(1):109-19 and Norrdin *et al.*, 1995, *Bone* 17(5):485-9); a feline model of Niemann-Pick disease type C (see, e.g., March *et al.*, 1997, *Acta Neuropathol (Berl.)* 94(2):164-72); acid sphingomyelinase-deficient mice (see, e.g., Otterbach & Stoffel, 10 1995, *Cell* 81(7):1053-6), and bovine mannosidosis (see, e.g., Jolly *et al.*, 1975, *Birth Defects Orig Artic Ser.* 11(6):273-8).

Examples of animal models for tuberous sclerosis ("TSC") include, but are not limited to, a mouse model of TSC1 (see, e.g., Kwiatkowski *et al.*, 2002, *Hum Mol Genet.* 11(5):525-34), a Tsc1 (TSC1 homologue) knockout mouse (see, e.g., Kobayashi *et al.*, 2001, *Proc Natl Acad Sci U S A.* 2001 Jul 17;98(15):8762-7), a TSC2 gene mutant(Eker) rat model (see, e.g., Hino 2000, *Nippon Rinsho* 58(6):1255-61; Mizuguchi *et al.*, 2000, *J Neuropathol Exp Neurol.* 59(3):188-9; and Hino *et al.*, 1999, *Prog Exp Tumor Res.* 35:95-108); and Tsc2(+/-) mice (see, e.g., Onda *et al.*, 1999, *J Clin Invest.* 20 104(6):687-95).

5.6.3. Toxicity

The toxicity and/or efficacy of a compound identified in accordance with the invention can be determined by standard pharmaceutical procedures in cell cultures or experimental animals, e.g., for determining the LD₅₀ (the dose lethal to 50% of the population) and the ED₅₀ (the dose therapeutically effective in 50% of the population). Cells and cell lines that can be used to assess the cytotoxicity of a compound identified in accordance with the invention include, but are not limited to, peripheral blood mononuclear cells (PBMCs), Caco-2 cells, and Huh7 cells. The dose ratio between toxic and therapeutic effects is the therapeutic index and it can be expressed as the ratio LD₅₀/ED₅₀. A compound identified in accordance with the invention that exhibits large therapeutic indices is preferred. While a compound identified in accordance with the invention that exhibits toxic side effects may be used, care should be taken to design a delivery system that targets such

agents to the site of affected tissue in order to minimize potential damage to uninfected cells and, thereby, reduce side effects.

The data obtained from the cell culture assays and animal studies can be used in formulating a range of dosage of a compound identified in accordance with the invention for use in humans. The dosage of such agents lies preferably within a range of circulating concentrations that include the ED₅₀ with little or no toxicity. The dosage may vary within this range depending upon the dosage form employed and the route of administration utilized. For any agent used in the method of the invention, the therapeutically effective dose can be estimated initially from cell culture assays. A dose may be formulated in animal models to achieve a circulating plasma concentration range that includes the IC₅₀ (*i.e.*, the concentration of the compound that achieves a half-maximal inhibition of symptoms) as determined in cell culture. Such information can be used to more accurately determine useful doses in humans. Levels in plasma may be measured, for example, by high performance liquid chromatography.

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5.7. Design of Congeners or Analogs

The compounds which display the desired biological activity can be used as lead compounds for the development or design of congeners or analogs having useful pharmacological activity. For example, once a lead compound is identified, molecular modeling techniques can be used to design variants of the compound that can be more effective. Examples of molecular modeling systems are the CHARM and QUANTA programs (Polygen Corporation, Waltham, MA). CHARM performs the energy minimization and molecular dynamics functions. QUANTA performs the construction, graphic modelling and analysis of molecular structure. QUANTA allows interactive construction, modification, visualization, and analysis of the behavior of molecules with each other.

A number of articles review computer modeling of drugs interactive with specific proteins, such as Rotivinen et al., 1988, Acta Pharmaceutical Fennica 97:159-166; Ripka, 1998, New Scientist 54-57; McKinlay & Rossmann, 1989, Annu. Rev. Pharmacol. Toxicol. 29:111-122; Perry & Davies, OSAR: Quantitative Structure-Activity Relationships in Drug Design pp. 189-193 (Alan R. Liss, Inc. 1989); Lewis & Dean, 1989, Proc. R. Soc. Lond. 236:125-140 and 141-162; Askew et al., 1989, J. Am. Chem. Soc. 111:1082-1090. Other computer programs that screen and graphically depict chemicals are available from companies such as BioDesign, Inc. (Pasadena, California), Allelix, Inc. (Mississauga, Ontario, Canada), and Hypercube, Inc. (Cambridge, Ontario). Although these

are primarily designed for application to drugs specific to particular proteins, they can be adapted to design of drugs specific to any identified region. The analogs and congeners can be tested for binding to translational machinery using assays well-known in the art or described herein for biologic activity. Alternatively, lead compounds with little or no 5 biologic activity, as ascertained in the screen, can also be used to design analogs and congeners of the compound that have biologic activity.

5.8. Use of Identified Compounds to Treat/Prevent a Disease or Disorder

The present invention provides methods of preventing, treating, managing or ameliorating a disorder associated with premature translation termination and/or nonsense-mediated mRNA decay, or one or more symptoms thereof, said methods comprising 10 administering to a subject in need thereof one or more compounds identified in accordance with the methods of the invention or a pharmaceutically acceptable salt thereof. Examples of diseases associated with, characterized by or caused by associated with premature translation termination and/or nonsense-mediated mRNA decay include, but are not limited 15 to, cystic fibrosis, muscular dystrophy, heart disease, lung cancer, breast cancer, colon cancer, pancreatic cancer, non-Hodgkin's lymphoma, ovarian cancer, esophageal cancer, colorectal carcinomas, neurofibromatosis, retinoblastoma, Wilm's tumor, retinitis pigmentosa, collagen disorders, cirrhosis, Tay-Sachs disease, blood disorders, kidney stones, ataxia-telangiectasia, lysosomal storage diseases, and tuberous sclerosis. See 20 Sections 5.5 and 8 for additional non-limiting examples of diseases and genetic disorders which can be prevented, treated, managed or ameliorated by administering one or more of the compounds identified in accordance with the methods of the invention or a pharmaceutically acceptable salt thereof. Genes that contain one or more nonsense mutations that are potentially involved in causing disease are presented in table form 25 according to chromosome location in Example 8 *infra*.

In a preferred embodiment, it is first determined that the patient is suffering from a disease associated with premature translation termination and/or nonsense-mediated mRNA decay before administering a compound identified in accordance with the invention or a combination therapy described herein. In a preferred embodiment, the DNA of the 30 patient can be sequenced or subject to Southern Blot, polymerase chain reaction (PCR), use of the Short Tandem Repeat (STR), or polymorphic length restriction fragments (RFLP) analysis to determine if a nonsense mutation is present in the DNA of the patient. Alternatively, it can be determined if altered levels of the protein with the nonsense

mutation are expressed in the patient by western blot or other immunoassays. Such methods are well known to one of skill in the art.

In one embodiment, the invention provides a method of preventing, treating, managing or ameliorating a disorder or one or more symptoms thereof, said method

- 5 comprising administering to a subject in need thereof a dose of a prophylactically or therapeutically effective amount of one or more compounds identified in accordance with the methods of the invention. In another embodiment, a compound identified in accordance with the methods of the invention is not administered to prevent, treat, or ameliorate a disorder or one or more symptoms thereof, if such compound has been used previously to
10 prevent, treat, manage or ameliorate said disorder. In a more specific embodiment of the invention, disorders that can be treated with the compounds of the invention, include, but are not limited to, disorders that are associated with, characterized by or caused by premature translation termination and/or nonsense mediated mRNA decay.

The invention also provides methods of preventing, treating, managing or
15 ameliorating a disorder associated with, characterized by or caused by premature translation termination and/or nonsense mediated mRNA decay, or one or more symptoms thereof, said methods comprising administering to a subject in need thereof one or more of the compounds identified utilizing the screening methods described herein or a pharmaceutically acceptable salt thereof, and one or more other therapies (e.g., prophylactic
20 or therapeutic agents). Preferably, the other therapies are currently being used, have been used or are known to be useful in the prevention, treatment, management or amelioration of said disorder or a symptom thereof. Non-limiting examples of such therapies are in Section 5.8.1 *infra*.

The therapies (e.g., prophylactic or therapeutic agents) or the combination
25 therapies of the invention can be administered sequentially or concurrently. In a specific embodiment, the combination therapies of the invention comprise a compound identified in accordance with the invention and at least one other therapy that has the same mechanism of action as said compound. In another specific embodiment, the combination therapies of the invention comprise a compound identified in accordance with the methods of the invention
30 and at least one other therapy (e.g., prophylactic or therapeutic agent) which has a different mechanism of action than said compound. The combination therapies of the present invention improve the prophylactic or therapeutic effect of a compound of the invention by functioning together with the compound to have an additive or synergistic effect. The

combination therapies of the present invention reduce the side effects associated with the therapies (e.g., prophylactic or therapeutic agents).

The prophylactic or therapeutic agents of the combination therapies can be administered to a subject in the same pharmaceutical composition. Alternatively, the prophylactic or therapeutic agents of the combination therapies can be administered concurrently to a subject in separate pharmaceutical compositions. The prophylactic or therapeutic agents may be administered to a subject by the same or different routes of administration.

In a specific embodiment, a pharmaceutical composition comprising one or more compounds identified in a screening assay described herein is administered to a subject, preferably a human, to prevent, treat, manage or ameliorate a disorder associated with, characterized by or caused by premature translation termination and/or nonsense mediated mRNA decay or one or more symptoms thereof. In accordance with the invention, the pharmaceutical composition may also comprise one or more other prophylactic or therapeutic agents. Preferably, such prophylactic or therapeutic agents are currently being used, have been used or are known to be useful in the prevention, treatment, management or amelioration of a disorder associated with, characterized by, or caused by premature translation termination or nonsense-mediated mRNA decay or one or more symptoms thereof.

A compound identified in accordance with the methods of the invention may be used as a first, second, third, fourth or fifth line of therapy for a disorder associated with, characterized by or caused by premature translation termination and/or nonsense-mediated mRNA decay. The invention provides methods for treating, managing or ameliorating a disorder associated with, characterized by or caused by premature translation termination and/or nonsense-mediated mRNA decay or one or more symptoms thereof in a subject refractory to conventional therapies for such disorder, said methods comprising administering to said subject a dose of a prophylactically or therapeutically effective amount of a compound identified in accordance with the methods of the invention. In particular, a disorder may be determined to be refractory to a therapy when at least some significant portion of the disorder is not resolved in response to the therapy. Such a determination can be made either *in vivo* or *in vitro* by any method known in the art for assaying the effectiveness of a therapy on a subject, using the art-accepted meanings of "refractory" in such a context. In a specific embodiment, a disorder is refractory where the number of symptoms of the disorder has not been significantly reduced, or has increased.

The invention provides methods for treating, managing or ameliorating one or more symptoms of a disorder associated with, characterized by or caused by premature translation termination and/or nonsense-mediated mRNA decay in a subject refractory to existing single agent therapies for such disorder, said methods comprising administering to

5 said subject a dose of a prophylactically or therapeutically effective amount of a compound identified in accordance with the methods of the invention and a dose of a prophylactically or therapeutically effective amount of one or more other therapies (e.g., prophylactic or therapeutic agents). The invention also provides methods for treating or managing a disorder associated with, characterized by or caused by premature translation termination

10 and/or nonsense-mediated mRNA decay by administering a compound identified in accordance with the methods of the invention in combination with any other therapy (e.g., radiation therapy, chemotherapy or surgery) to patients who have proven refractory to other therapies but are no longer on these therapies. The invention also provides methods for the treatment or management of a patient having a disorder associated with, characterized by or

15 caused by premature translation termination and/or nonsense-mediated mRNA decay and said patient is immunosuppressed by reason of having previously undergone other therapies. Further, the invention provides methods for preventing the recurrence of a disorder associated with, characterized by or caused by premature translation termination and/or nonsense-mediated mRNA decay such as, e.g., cancer in patients that have been undergone

20 therapy and have no disease activity by administering a compound identified in accordance with the methods of the invention.

5.8.1. Other Therapies

The present invention provides methods of preventing, treating, managing or ameliorating a disorder associated with, characterized by or caused by premature translation termination and/or nonsense-mediated mRNA decay, or one or more symptoms thereof, said methods comprising administering to a subject in need thereof one or more compounds identified in accordance with the methods of the invention or a pharmaceutically acceptable salt thereof, and one or more other therapies (e.g., prophylactic or therapeutic agents). Any therapy (e.g., chemotherapies, radiation therapies, hormonal therapies, and/or biological therapies/immunotherapies) which is known to be useful, or which has been used or is currently being used for the prevention, treatment, management or amelioration of disorders associated with, characterized by or caused by premature translation termination and/or nonsense-mediated mRNA decay or one or more symptoms thereof can be used in combination with a compound identified in accordance with the methods of the invention.

Examples of therapeutic or prophylactic agents which can be used in combination with a compound identified in accordance with the invention include, but are not limited to, peptides, polypeptides, fusion proteins, nucleic acid molecules, small molecules, mimetic agents, synthetic drugs, inorganic molecules, and organic molecules.

- 5 Proliferative disorders associated with, characterized by or caused by premature translation termination and/or nonsense-mediated mRNA decay can be prevented, treated, managed or ameliorated by administering to a subject in need thereof one or more of the compounds identified in accordance with the methods of the invention, and one or more other therapies for prevention, treatment, management or amelioration of said disorders or a symptom thereof. Examples of such therapies include, but are not limited to, angiogenesis inhibitors, topoisomerase inhibitors, immunomodulatory agents (such as chemotherapeutic agents) and radiation therapy. Angiogenesis inhibitors (*i.e.*, anti-angiogenic agents) include, but are not limited to, angiostatin (plasminogen fragment); antiangiogenic antithrombin III; angiozyme; ABT-627; Bay 12-9566; Benefin; Bevacizumab; BMS-275291; cartilage-derived inhibitor (CDI); CAI; CD59 complement fragment; CEP-7055; Col 3; combretastatin A-4; endostatin (collagen XVIII fragment); fibronectin fragment; Gro-beta; Halofuginone; Heparinases; Heparin hexasaccharide fragment; HMV833; human chorionic gonadotropin (hCG); IM-862; Interferon alpha/beta/gamma; Interferon inducible protein (IP-10); Interleukin-12; Kringle 5 (plasminogen fragment); Marimastat; Metalloproteinase inhibitors (TIMPs); 2-methoxyestradiol; MMI 270 (CGS 27023A); MoAb IMC-1C11; Neovastat; NM-3; Panzem; PI-88; Placental ribonuclease inhibitor; plasminogen activator inhibitor; platelet factor-4 (PF4); Prinomastat; Prolactin 16kD fragment; Proliferin-related protein (PRP); PTK 787/ZK 222594; retinoids; solimastat; squalamine; SS 3304; SU 5416; SU6668; SU11248; 25 tetrahydrocortisol-S; tetrathiomolybdate; thalidomide; thrombospondin-1 (TSP-1); TNP-470; transforming growth factor-beta; vasculostatin; vasostatin (calreticulin fragment); ZD6126; ZD 6474; farnesyl transferase inhibitors (FTI); and bisphosphonates. In a specific embodiment, anti-angiogenic agents do not include antibodies or fragments thereof that immunospecifically bind to integrin $\alpha_v\beta_3$.
- 30 Specific examples of prophylactic or therapeutic agents which can be used in accordance with the methods of the invention to prevent, treat, manage or ameliorate a proliferative disorder associated with, characterized by or caused by premature translation termination and/or nonsense-mediated mRNA decay, or a symptom thereof include, but not limited to: acivicin; aclarubicin; acodazole hydrochloride; acronine; adozelesin; aldesleukin;

altretamine; ambomycin; ametantrone acetate; aminoglutethimide; amsacrine; anastrozole;
anthramycin; asparaginase; asperlin; azacitidine; azetepa; azotomycin; batimastat;
benzodepa; bicalutamide; bisantrene hydrochloride; bisnafide dimesylate; bizelesin;
bleomycin sulfate; brequinar sodium; bropirimine; busulfan; cactinomycin; calusterone;
5 caracemide; carbetimer; carboplatin; carmustine; carubicin hydrochloride; carzelesin;
cedefingol; chlorambucil; cirolemycin; cisplatin; cladribine; crisnatol mesylate;
cyclophosphamide; cytarabine; dacarbazine; dactinomycin; daunorubicin hydrochloride;
decitabine; dexormaplatin; dezaguanine; dezaguanine mesylate; diaziquone; docetaxel;
doxorubicin; doxorubicin hydrochloride; droloxifene; droloxifene citrate; dromostanolone
10 propionate; duazomycin; edatrexate; eflornithine hydrochloride; elsamitruclin; enloplatin;
enpromate; epipropidine; epirubicin hydrochloride; erbulozole; esorubicin hydrochloride;
estramustine; estramustine phosphate sodium; etanidazole; etoposide; etoposide phosphate;
etoprime; fadrozole hydrochloride; fazarabine; fenretinide; floxuridine; fludarabine
phosphate; fluorouracil; flurocitabine; fosquidone; fostriecin sodium; gemcitabine;
15 gemcitabine hydrochloride; hydroxyurea; idarubicin hydrochloride; ifosfamide; ilmofosine;
interleukin II (including recombinant interleukin II, or rIL2), interferon alpha-2a; interferon
alpha-2b; interferon alpha-n1 ; interferon alpha-n3; interferon beta-I a; interferon gamma-I
b; iproplatin; irinotecan hydrochloride; lanreotide acetate; letrozole; leuprolide acetate;
liarozole hydrochloride; lometrexol sodium; lomustine; losoxantrone hydrochloride;
20 masoprocol; maytansine; mechlorethamine hydrochloride; megestrol acetate; melengestrol
acetate; melphalan; menogaril; mercaptapurine; methotrexate; methotrexate sodium;
metoprine; meturedepa; mitindomide; mitocarcin; mitocromin; mitogillin; mitomalcin;
mitomycin; mitosper; mitotane; mitoxantrone hydrochloride; mycophenolic acid;
nocodazole; nogalamycin; ormaplatin; oxisuran; paclitaxel; pegaspargase; peliomycin;
25 pentamustine; peplomycin sulfate; perfosfamide; pipobroman; pipsulfan; piroxantrone
hydrochloride; plicamycin; plomestane; porfimer sodium; porfiromycin; prednimustine;
procabazine hydrochloride; puromycin; puromycin hydrochloride; pyrazofurin; riboprine;
rogletimide; safingol; safingol hydrochloride; semustine; simtrazene; sparfosate sodium;
sparsomycin; spirogermanium hydrochloride; spiromustine; spiroplatin; streptonigrin;
30 streptozocin; sulofenur; talisomycin; tecogalan sodium; tegafur; teloxantrone hydrochloride;
temoporfin; teniposide; teroxirone; testolactone; thiamiprime; thioguanine; thiotepla;
tiazofurin; tirapazamine; toremifene citrate; trestolone acetate; triciribine phosphate;
trimetrexate; trimetrexate glucuronate; triptorelin; tubulozole hydrochloride; uracil mustard;
uredepa; vapreotide; verteporfin; vinblastine sulfate; vincristine sulfate; vindesine;

vindesine sulfate; vinepidine sulfate; vinglycinate sulfate; vinleurosine sulfate; vinorelbine tartrate; vinrosidine sulfate; vinzolidine sulfate; vorozole; zeniplatin; zinostatin; zorubicin hydrochloride. Other anti-cancer drugs include, but are not limited to: 20-epi-1,25 dihydroxyvitamin D3; 5-ethynyluracil; abiraterone; aclarubicin; acylfulvene; adecy penol; 5 adozelesin; aldesleukin; ALL-TK antagonists; altretamine; ambamustine; amidox; amifostine; aminolevulinic acid; amrubicin; amsacrine; anagrelide; anastrozole; andrographolide; angiogenesis inhibitors; antagonist D; antagonist G; antarelix; anti-dorsalizing morphogenetic protein-1; antiandrogen, prostatic carcinoma; antiestrogen; 10 antineoplaston; antisense oligonucleotides; aphidicolin glycinate; apoptosis gene modulators; apoptosis regulators; apurinic acid; ara-CDP-DL-PTBA; arginine deaminase; asulacrine; atamestane; atrimustine; axinastatin 1; axinastatin 2; axinastatin 3; azasetron; azatoxin; azatyrosine; baccatin III derivatives; balanol; batimastat; BCR/ABL antagonists; benzochlorins; benzoylstaurosporine; beta lactam derivatives; beta-alethine; betaclamycin B; betulinic acid; bFGF inhibitor; bicalutamide; bisantrene; bisaziridinylspermine; 15 bisnafide; bistratene A; bizelesin; breflate; bropirimine; budotitane; buthionine sulfoximine; calcipotriol; calphostin C; camptothecin derivatives; canarypox IL-2; capecitabine; carboxamide-amino-triazole; carboxyamidotriazole; CaRest M3; CARN 700; cartilage derived inhibitor; carzelesin; casein kinase inhibitors (ICOS); castanospermine; cecropin B; cetrorelix; chlorlins; chloroquinoxaline sulfonamide; cicaprost; cis-porphyrin; cladribine; 20 clomifene analogues; clotrimazole; collismycin A; collismycin B; combretastatin A4; combretastatin analogue; conagenin; crambescidin 816; crisnatol; cryptophycin 8; cryptophycin A derivatives; curacin A; cyclopentanthraquinones; cycloplatam; cypemycin; cytarabine ocfosfate; cytolytic factor; cytostatin; dacliximab; decitabine; dehydrodide mnin B; deslorelin; dexamethasone; dexifosfamide; dexrazoxane; dexverapamil; diaziquone; 25 didemnin B; didox; diethylnorspermine; dihydro-5-azacytidine; dihydrotaxol, 9-; dioxamycin; diphenyl spiromustine; docetaxel; docosanol; dolasetron; doxifluridine; droloxitene; dronabinol; duocarmycin SA; ebselen; ecomustine; edelfosine; edrecolomab; eflornithine; elemene; emitefur; epirubicin; epristeride; estramustine analogue; estrogen agonists; estrogen antagonists; etanidazole; etoposide phosphate; exemestane; fadrozole; 30 fazarabine; fenretinide; filgrastim; finasteride; flavopiridol; flezelastine; fluasterone; fludarabine; fluorodaunorubicin hydrochloride; forfenimex; formestane; fostriecin; fotemustine; gadolinium texaphyrin; gallium nitrate; galocitabine; ganirelix; gelatinase inhibitors; gemcitabine; glutathione inhibitors; hepsulfam; heregulin; hexamethylene bisacetamide; hypericin; ibandronic acid; idarubicin; idoxifene; idramantone; ilmofosine;

ilomastat; imidazoacridones; imiquimod; immunostimulant peptides; insulin-like growth factor-1 receptor inhibitor; interferon agonists; interferons; interleukins; iobenguane; iododoxorubicin; ipomeanol, 4-; iroplact; irsogladine; isobengazole; isohomohalicondrin B; itasetron; jasplakinolide; kahalalide F; lamellarin-N triacetate; lanreotide; leinamycin;

5 lenograstim; lentinan sulfate; leptolstatin; letrozole; leukemia inhibiting factor; leukocyte alpha interferon; leuprolide+estrogen+progesterone; leuprorelin; levamisole; liarozole; linear polyamine analogue; lipophilic disaccharide peptide; lipophilic platinum compounds; lissoclinamide 7; lobaplatin; lombricine; lometrexol; lonidamine; losoxantrone; lovastatin; loxoribine; lurtotecan; lutetium texaphyrin; lysofylline; lytic peptides; maitansine;

10 mannostatin A; marimastat; masoprolol; maspin; matrilysin inhibitors; matrix metalloproteinase inhibitors; menogaril; merbarone; meterelin; methioninase; metoclopramide; MIF inhibitor; mifepristone; miltefosine; mirimostim; mismatched double stranded RNA; mitoguazone; mitolactol; mitomycin analogues; mitonafide; mitotoxin fibroblast growth factor-saporin; mitoxantrone; mofarotene; molgramostim; monoclonal

15 antibody, human chorionic gonadotrophin; monophosphoryl lipid A+myobacterium cell wall sk; molidamol; multiple drug resistance gene inhibitor; multiple tumor suppressor 1-based therapy; mustard anticancer agent; mycaperoxide B; mycobacterial cell wall extract; myriaporone; N-acetylinaline; N-substituted benzamides; nafarelin; nagrestip; naloxone+pentazocine; napavin; naphterpin; nartograstim; nedaplatin; nemorubicin;

20 neridronic acid; neutral endopeptidase; nilutamide; nisamycin; nitric oxide modulators; nitroxide antioxidant; nitrullyn; O6-benzylguanine; octreotide; okicenone; oligonucleotides; onapristone; ondansetron; ondansetron; oracin; oral cytokine inducer; ormaplatin; osaterone; oxaliplatin; oxaunomycin; paclitaxel; paclitaxel analogues; paclitaxel derivatives; palauamine; palmitoylrhizoxin; pamidronic acid; panaxytriol; panomifene; parabactin;

25 pazelliptine; pegaspargase; peldesine; pentosan polysulfate sodium; pentostatin; pentrozole; perflubron; perfosfamide; perillyl alcohol; phenazinomycin; phenylacetate; phosphatase inhibitors; picibanil; pilocarpine hydrochloride; pirarubicin; piritrexim; placetin A; placetin B; plasminogen activator inhibitor; platinum complex; platinum compounds; platinum-triamine complex; porfimer sodium; porfiromycin; prednisone; propyl

30 bis-acridone; prostaglandin J2; proteasome inhibitors; protein A-based immune modulator; protein kinase C inhibitor; protein kinase C inhibitors, microalgal; protein tyrosine phosphatase inhibitors; purine nucleoside phosphorylase inhibitors; purpurins; pyrazoloacridine; pyridoxylated hemoglobin polyoxyethylene conjugate; raf antagonists; raltitrexed; ramosetron; ras farnesyl protein transferase inhibitors; ras inhibitors; ras-GAP

inhibitor; retelliptine demethylated; rhenium Re 186 etidronate; rhizoxin; ribozymes; RII
retinamide; rogletimide; rohitukine; romurtide; roquinimex; rubiginone B1; ruboxyl;
safingol; saintopin; SarCNU; sarcophytol A; sargramostim; Sdi 1 mimetics; semustine;
senescence derived inhibitor 1; sense oligonucleotides; signal transduction inhibitors; signal
5 transduction modulators; single chain antigen binding protein; sizofiran; sobuzoxane;
sodium borocaptate; sodium phenylacetate; solverol; somatomedin binding protein;
sönermin; sparfосic acid; spicamycin D; spiromustine; splenopentin; spongistatin 1;
squalamine; stem cell inhibitor; stem-cell division inhibitors; stipiamide; stromelysin
inhibitors; sulfinosine; superactive vasoactive intestinal peptide antagonist; suradista;
10 suramin; swainsonine; synthetic glycosaminoglycans; tallimustine; 5-fluorouracil;
leucovorin; tamoxifen methiodide; tauromustine; tazarotene; tecogalan sodium; tegafur;
tellurapyrylium; telomerase inhibitors; temoporfin; temozolomide; teniposide;
tetrachlorodecaoxide; tetrazomine; thaliblastine; thiocoraline; thrombopoietin;
thrombopoietin mimetic; thymalfasin; thymopoietin receptor agonist; thymotrinan; thyroid
15 stimulating hormone; tin ethyl etiopurpurin; tirapazamine; titanocene bichloride; topsentin;
toremifene; totipotent stem cell factor; translation inhibitors; tretinoin; triacetyluridine;
triciribine; trimetrexate; triptorelin; tropisetron; turosteride; tyrosine kinase inhibitors;
tyrphostins; UBC inhibitors; ubenimex; urogenital sinus-derived growth inhibitory factor;
urokinase receptor antagonists; vapreotide; variolin B; vector system, erythrocyte gene
20 therapy; thalidomide; velaresol; veramine; verdins; verteporfin; vinorelbine; vinxaltine;
vorozole; zanoterone; zeniplatin; zilascorb; and zinostatin stimalamer.

Specific examples of prophylactic or therapeutic agents which can be used in accordance with the methods of the invention to prevent, treat, manage and/or ameliorate a central nervous system disorders associated with, characterized by or caused by premature translation termination and/or nonsense-mediated mRNA decay, or a symptom thereof include, but are not limited to: Levodopa, L-DOPA, cocaine, α -methyl-tyrosine, reserpine, tetrabenazine, benzotropine, pargyline, fenodolpam mesylate, cabergoline, pramipexole dihydrochloride, ropinirole, amantadine hydrochloride, selegiline hydrochloride, carbidopa, pergolide mesylate, Sinemet CR, or Symmetrel.

30 Specific examples of prophylactic or therapeutic agents which can be used in accordance with the methods of the invention to prevent, treat, manage and/or ameliorate a metabolic disorders associated with, characterized by or caused by premature translation termination and/or nonsense-mediated mRNA decay, or a symptom thereof include, but are not limited to: a monoamine oxidase inhibitor (MAO), for example, but not limited to,

iproniazid, clorgyline, phenelzine and isocarboxazid; an acetylcholinesterase inhibitor, for example, but not limited to, physostigmine saliclate, physostigmine sulfate, physostigmine bromide, meostigmine bromide, neostigmine methylsulfate, ambenonium chloride, edrophonium chloride, tacrine, pralidoxime chloride, obidoxime chloride, trimedoxime 5 bromide, diacetyl monoxim, endrophonium, pyridostigmine, and demecarium; an anti-inflamatory agent, including, but not limited to, naproxen sodium, diclofenac sodium, diclofenac potassium, celecoxib, sulindac, oxaprozin, diflunisal, etodolac, meloxicam, ibuprofen, ketoprofen, nabumetone, refecoxib, methotrexate, leflunomide, sulfasalazine, gold salts, RHo-D Immune Globulin, mycophenylate mofetil, cyclosporine, azathioprine, 10 tacrolimus, basiliximab, daclizumab, salicylic acid, acetylsalicylic acid, methyl salicylate, diflunisal, salsalate, olsalazine, sulfasalazine, acetaminophen, indomethacin, sulindac, mefenamic acid, meclofenamate sodium, tolmetin, ketorolac, dichlofenac, flurbiprofen, oxaprozin, piroxicam, meloxicam, ampiroxicam, droxicam, pivoxicam, tenoxicam, phenylbutazone, oxyphenbutazone, antipyrine, aminopyrine, apazone, zileuton, 15 aurothioglucose, gold sodium thiomalate, auranofin, methotrexate, colchicine, allopurinol, probenecid, sulfapyrazone and benzboronate or betamethasone and other glucocorticoids; an antiemetic agent, for example, but not limited to, metoclopramide, domperidone, prochlorperazine, promethazine, chlorpromazine, trimethobenzamide, ondansetron, granisetron, hydroxyzine, acetylleucine monoethanolamine, alizapride, azasetron, 20 benzquinamide, bietanautine, bromopride, buclizine, clebopride, cyclizine, dimenhydrinate, diphenidol, dolasetron, meclizine, methallatal, metopimazine, nabilone, oxperndyl, pipamazine, scopolamine, sulpiride, tetrahydrocannabinol, thiethylperazine, thioproperazine, tropisetron, and mixtures thereof.

5.9. Compounds and Methods of Administering Compounds

25 Biologically active compounds identified using the methods of the invention or a pharmaceutically acceptable salt thereof can be administered to a patient, preferably a mammal, more preferably a human, suffering from a disorder associated with, characterized by or caused by premature translation termination and/or nonsense mediated mRNA decay. In a specific embodiment, a compound or a pharmaceutically acceptable salt thereof is administered to a patient, preferably a mammal, more preferably a human, as a preventative measure against a disorder associated with, characterized by or caused by premature 30 translation termination and/or nonsense-mediated mRNA decay.

When administered to a patient, the compound or a pharmaceutically acceptable salt thereof is preferably administered as component of a composition that

optionally comprises a pharmaceutically acceptable vehicle. The composition can be administered orally, or by any other convenient route, for example, by infusion or bolus injection, by absorption through epithelial or mucocutaneous linings (*e.g.*, oral mucosa, rectal, and intestinal mucosa, *etc.*) and may be administered together with another 5 biologically active agent. Administration can be systemic or local. Various delivery systems are known, *e.g.*, encapsulation in liposomes, microparticles, microcapsules, capsules, *etc.*, and can be used to administer the compound and pharmaceutically acceptable salts thereof.

Methods of administration include but are not limited to intradermal, 10 intramuscular, intraperitoneal, intravenous, subcutaneous, intranasal, epidural, oral, sublingual, intranasal, intracerebral, intravaginal, transdermal, rectally, by inhalation, or topically, particularly to the ears, nose, eyes, or skin. The mode of administration is left to the discretion of the practitioner. In most instances, administration will result in the release of the compound or a pharmaceutically acceptable salt thereof into the bloodstream.

15 In specific embodiments, it may be desirable to administer the compound or a pharmaceutically acceptable salt thereof locally. This may be achieved, for example, and not by way of limitation, by local infusion during surgery, topical application, *e.g.*, in conjunction with a wound dressing after surgery, by injection, by means of a catheter, by means of a suppository, or by means of an implant, said implant being of a porous, non-porous, 20 or gelatinous material, including membranes, such as sialastic membranes, or fibers.

In certain embodiments, it may be desirable to introduce the compound or a pharmaceutically acceptable salt thereof into the central nervous system by any suitable route, including intraventricular, intrathecal and epidural injection. Intraventricular injection may be facilitated by an intraventricular catheter, for example, attached to a 25 reservoir, such as an Ommaya reservoir.

Pulmonary administration can also be employed, *e.g.*, by use of an inhaler or nebulizer, and formulation with an aerosolizing agent, or via perfusion in a fluorocarbon or synthetic pulmonary surfactant. In certain embodiments, the compound and pharmaceutically acceptable salts thereof can be formulated as a suppository, with 30 traditional binders and vehicles such as triglycerides.

In another embodiment, the compound and pharmaceutically acceptable salts thereof can be delivered in a vesicle, in particular a liposome (see Langer, 1990, *Science* 249:1527-1533; Treat et al., in *Liposomes in the Therapy of Infectious Disease and Cancer*,

Lopez-Berestein and Fidler (eds.), Liss, New York, pp. 353-365 (1989); Lopez-Berestein, *ibid.*, pp. 317-327; see generally *ibid.*).

In yet another embodiment, the compound and pharmaceutically acceptable salts thereof can be delivered in a controlled release system (see, e.g., Goodson, in Medical Applications of Controlled Release, *supra*, vol. 2, pp. 115-138 (1984)). Other controlled-release systems discussed in the review by Langer, 1990, Science 249:1527-1533 may be used. In one embodiment, a pump may be used (see Langer, *supra*; Sefton, 1987, CRC Crit. Ref. Biomed. Eng. 14:201; Buchwald et al., 1980, Surgery 88:507; Saudek et al., 1989, N. Engl. J. Med. 321:574). In another embodiment, polymeric materials can be used (see Medical Applications of Controlled Release, Langer and Wise (eds.), CRC Pres., Boca Raton, Florida (1974); Controlled Drug Bioavailability, Drug Product Design and Performance, Smolen and Ball (eds.), Wiley, New York (1984); Ranger and Peppas, 1983, J. Macromol. Sci. Rev. Macromol. Chem. 23:61; see also Levy et al., 1985, Science 228:190; During et al., 1989, Ann. Neurol. 25:351; Howard et al., 1989, J. Neurosurg. 71:105). In yet another embodiment, a controlled-release system can be placed in proximity of a target RNA of the compound or a pharmaceutically acceptable salt thereof, thus requiring only a fraction of the systemic dose.

Compositions comprising the compound or a pharmaceutically acceptable salt thereof ("compound compositions") can additionally comprise a suitable amount of a pharmaceutically acceptable vehicle so as to provide the form for proper administration to the patient.

In a specific embodiment, the term "pharmaceutically acceptable" means approved by a regulatory agency of the Federal or a state government or listed in the U.S. Pharmacopeia or other generally recognized pharmacopeia for use in animals, mammals, and more particularly in humans. The term "vehicle" refers to a diluent, adjuvant, excipient, or carrier with which a compound of the invention is administered. Such pharmaceutical vehicles can be liquids, such as water and oils, including those of petroleum, animal, vegetable or synthetic origin, such as peanut oil, soybean oil, mineral oil, sesame oil and the like. The pharmaceutical vehicles can be saline, gum acacia, gelatin, starch paste, talc, keratin, colloidal silica, urea, and the like. In addition, auxiliary, stabilizing, thickening, lubricating and coloring agents may be used. When administered to a patient, the pharmaceutically acceptable vehicles are preferably sterile. Water is a preferred vehicle when the compound of the invention is administered intravenously. Saline solutions and aqueous dextrose and glycerol solutions can also be employed as liquid vehicles,

particularly for injectable solutions. Suitable pharmaceutical vehicles also include excipients such as starch, glucose, lactose, sucrose, gelatin, malt, rice, flour, chalk, silica gel, sodium stearate, glycerol monostearate, talc, sodium chloride, dried skim milk, glycerol, propylene, glycol, water, ethanol and the like. Compound compositions, if desired, can also contain minor amounts of wetting or emulsifying agents, or pH buffering agents.

Compound compositions can take the form of solutions, suspensions, emulsion, tablets, pills, pellets, capsules, capsules containing liquids, powders, sustained-release formulations, suppositories, emulsions, aerosols, sprays, suspensions, or any other form suitable for use. In one embodiment, the pharmaceutically acceptable vehicle is a capsule (see e.g., U.S. Patent No. 5,698,155). Other examples of suitable pharmaceutical vehicles are described in Remington's Pharmaceutical Sciences, Alfonso R. Gennaro, ed., Mack Publishing Co. Easton, PA, 19th ed., 1995, pp. 1447 to 1676, incorporated herein by reference.

In a preferred embodiment, the compound or a pharmaceutically acceptable salt thereof is formulated in accordance with routine procedures as a pharmaceutical composition adapted for oral administration to human beings. Compositions for oral delivery may be in the form of tablets, lozenges, aqueous or oily suspensions, granules, powders, emulsions, capsules, syrups, or elixirs, for example. Orally administered compositions may contain one or more agents, for example, sweetening agents such as fructose, aspartame or saccharin; flavoring agents such as peppermint, oil of wintergreen, or cherry; coloring agents; and preserving agents, to provide a pharmaceutically palatable preparation. Moreover, where in tablet or pill form, the compositions can be coated to delay disintegration and absorption in the gastrointestinal tract thereby providing a sustained action over an extended period of time. Selectively permeable membranes surrounding an osmotically active driving compound are also suitable for orally administered compositions. In these later platforms, fluid from the environment surrounding the capsule is imbibed by the driving compound, which swells to displace the agent or agent composition through an aperture. These delivery platforms can provide an essentially zero order delivery profile as opposed to the spiked profiles of immediate release formulations. A time delay material such as glycerol monostearate or glycerol stearate may also be used. Oral compositions can include standard vehicles such as mannitol, lactose, starch, magnesium stearate, sodium saccharine, cellulose, magnesium carbonate, and the like. Such vehicles are preferably of pharmaceutical grade. Typically, compositions for intravenous administration comprise

sterile isotonic aqueous buffer. Where necessary, the compositions may also include a solubilizing agent.

In another embodiment, the compound or a pharmaceutically acceptable salt thereof can be formulated for intravenous administration. Compositions for intravenous administration may optionally include a local anesthetic such as lignocaine to lessen pain at the site of the injection. Generally, the ingredients are supplied either separately or mixed together in unit dosage form, for example, as a dry lyophilized powder or water-free concentrate in a hermetically sealed container such as an ampoule or sachette indicating the quantity of active agent. Where the compound or a pharmaceutically acceptable salt thereof is to be administered by infusion, it can be dispensed, for example, with an infusion bottle containing sterile pharmaceutical grade water or saline. Where the compound or a pharmaceutically acceptable salt thereof is administered by injection, an ampoule of sterile water for injection or saline can be provided so that the ingredients may be mixed prior to administration.

The amount of a compound or a pharmaceutically acceptable salt thereof that will be effective in the treatment of a particular disease will depend on the nature of the disease, and can be determined by standard clinical techniques. In addition, *in vitro* or *in vivo* assays may optionally be employed to help identify optimal dosage ranges. The precise dose to be employed will also depend on the route of administration, and the seriousness of the disease, and should be decided according to the judgment of the practitioner and each patient's circumstances. However, suitable dosage ranges for oral administration are generally about 0.001 milligram to about 500 milligrams of a compound or a pharmaceutically acceptable salt thereof per kilogram body weight per day. In specific preferred embodiments of the invention, the oral dose is about 0.01 milligram to about 100 milligrams per kilogram body weight per day, more preferably about 0.1 milligram to about 75 milligrams per kilogram body weight per day, more preferably about 0.5 milligram to 5 milligrams per kilogram body weight per day. The dosage amounts described herein refer to total amounts administered; that is, if more than one compound is administered, or if a compound is administered with a therapeutic agent, then the preferred dosages correspond to the total amount administered. Oral compositions preferably contain about 10% to about 95% active ingredient by weight.

Suitable dosage ranges for intravenous (i.v.) administration are about 0.01 milligram to about 100 milligrams per kilogram body weight per day, about 0.1 milligram to about 35 milligrams per kilogram body weight per day, and about 1 milligram to about

10 milligrams per kilogram body weight per day. Suitable dosage ranges for intranasal administration are generally about 0.01 pg/kg body weight per day to about 1 mg/kg body weight per day. Suppositories generally contain about 0.01 milligram to about 50 milligrams of a compound of the invention per kilogram body weight per day and comprise 5 active ingredient in the range of about 0.5% to about 10% by weight.

Recommended dosages for intradermal, intramuscular, intraperitoneal, subcutaneous, epidural, sublingual, intracerebral, intravaginal, transdermal administration or administration by inhalation are in the range of about 0.001 milligram to about 200 milligrams per kilogram of body weight per day. Suitable doses for topical administration 10 are in the range of about 0.001 milligram to about 1 milligram, depending on the area of administration. Effective doses may be extrapolated from dose-response curves derived from *in vitro* or animal model test systems. Such animal models and systems are well known in the art.

The compound and pharmaceutically acceptable salts thereof are preferably assayed *in vitro* and *in vivo*, for the desired therapeutic or prophylactic activity, prior to use 15 in humans. For example, *in vitro* assays can be used to determine whether it is preferable to administer the compound, a pharmaceutically acceptable salt thereof, and/or another therapeutic agent. Animal model systems can be used to demonstrate safety and efficacy.

20 **6. EXAMPLE: PREPARATION OF EXTRACTS FROM HELACELLS FOR
INVITRO TRANSLATION REACTIONS**

This Example describes a method of preparing a cell extract to perform *in vitro* translation reactions to monitor nonsense suppression or to produce proteins *in vitro*. This method is different from other methods used to prepare translation extracts for several reasons. First, the centrifugation step is performed at low speed (12000Xg) compared to 25 most other protocols that use a 100,000Xg spin; and second, the cells are incubated on ice for several hours to weeks, which increases the activity of the extract significantly.

6.1. Preparation of Translation Extract from Hela Cells

HeLa S3 cells were grown to a density of 10^6 cells/ml in DMEM; 5%CO₂, 10% FBS, 1X P/S in a spinner flask. Cells were harvested by spinning at 1000Xg. 30 Cells were washed twice with phosphate buffered saline. The cell pellet sat on ice for 12 to 24 hours before proceeding. By letting the cells sit on ice, the activity of the extract is increased two-fold. The length of time on ice can range from 0 hours to 1 week. The cells were resuspended in 1.5 volumes (packed cell volume) of hypotonic buffer (10 mM HEPES

(KOH) pH 7.4; 15 mM KCl; 1.5 mM Mg(OAc)₂; 0.5 mM Pefabloc (Roche); 2 mM DTT). Cells were allowed to swell for 5 minutes on ice and dounce homogenized with 10 to 100 strokes using a tight-fitting pestle. The cells were spun for 10 minutes at 12000Xg at 4°C in a Sorvall SS-34 rotor. The supernatant was carefully collected with a Pasteur pipet without disturbing the lipid layer and transferred into Eppendorf tubes (50-200 mL aliquots) and immediately frozen in liquid nitrogen. Figure 1 shows the amount of wild-type luciferase produced in an *in vitro* translation reaction when the amount of wild-type luciferase RNA and the amount of HeLa cell extract are varied. Figure 2 shows the amount of wild-type luciferase produced in an *in vitro* translation reaction when the amount of luciferase RNA containing the nonsense mutation RGA and the amount of the cell are varied.

6.2 Incubating Cells on Ice Improves Translation Activity of Extract

As shown in Figure 3, incubating cells on ice prior to preparation of the translation extract improves the translation activity up to 20 fold. Further, in the presence of the aminoglycoside gentamicin, nonsense suppression activity (as measured by the amount of luciferase activity produced from a luciferase RNA containing a UGA premature termination codon) is increased 2 to 3 fold above untreated extracts (see Figure 4). These results demonstrate that extracts prepared by this method actively translate wild type RNA as well as mediate nonsense suppression.

7. EXAMPLE: IDENTIFICATION AND CHARACTERIZATION OF COMPOUND THAT PROMOTE NONSENSE SUPPRESSION AND/OR MODULATE TRANSLATION TERMINATION

7.1. Development of Assays for High Throughput Screens

Two assays were developed for use in high throughput screens to identify small molecules that promote nonsense suppression. Each assay utilized luciferase because it is a functional reporter gene assay (light is only produced if the protein is functional) and it is extremely sensitive (Light intensity is proportional to luciferase concentration in the nM range). The first assay was a cell-based luciferase reporter assay and the second was a biochemical assay consisting of rabbit reticulocyte lysate and a nonsense-containing luciferase reporter mRNA. In the cell-based assay, a luciferase reporter construct containing a UGA premature termination codon was stably transfected in 293T Human Embryonic Kidney cells. In the biochemical assay, mRNA containing a UGA premature termination codon was used as a reporter in an *in vitro* translation reaction using rabbit

reticulocyte lysate supplemented with tRNA, hemin, creatine kinase, amino acids, KOAc, Mg(OAc)₂, and creatine phosphate. Translation of the mRNA was initiated within a virus derived leader sequence, which significantly reduced the cost of the assay because capped RNA was not required. Synthetic mRNA was prepared *in vitro* using the T7 promoter and 5 the MegaScript in vitro transcription kit (Ambion). In both of the biochemical and cell-based assays, addition of gentamicin, a small molecule known to allow readthrough of premature termination codons, resulted in increased luciferase activity and was, therefore, used as an internal standard.

7.2. Screening of a Chemical Library Using the Nonsense Suppression Assays

10 The assays described above in Section 7.1 were used in two high throughput screens. Approximately eight hundred thousand compounds were screened in the cell-based and biochemical assays. From these initial screens two hundred hits were retested with both luciferase assays and seven compounds were subsequently selected for further investigation. These compounds fall into four classes of scaffolds. One class of compound is a nucleoside 15 analog; the second class is a quinazoline compound; the third class is an oxadiazole compound similar to diarylfuran antibiotics; and the final class is a unique scaffold harboring one or more phenyl, amide, or similar functional groups. Interestingly, none of the compounds are similar in structure to gentamicin. Compound A (molecular formula C₁₉H₂₁NO₄), a member of the fourth class and Compound B (molecular formula 20 C₁₉H₁₈N₂O₄), a compound synthesized independently of the screen, because of its potential RNA binding properties were the focus of subsequent attention.

7.3. Compound A and Compound B Increase In Vitro Nonsense Suppression at UGA Codons

Based on the results of the high throughput screen, Compound A was 25 characterized further with the *in vitro* luciferase nonsense suppression assay. To ensure that the observed nonsense suppression activity of the selected compounds was not limited to the rabbit reticulocyte assay system, HeLa cell extract was prepared and optimized (Lie & Macdonald, 1999, Development 126(22):4989-4996 and Lie & Macdonald, 2000, Biochem. Biophys. Res. Commun. 270(2):473-481). Figure 5 shows that Compound A and 30 Compound B exhibit greater nonsense suppression activity of the UGA codon than gentamicin in the HeLa cell translation extracts.

7.4. Characterization of Compounds That Increase Nonsense Suppression and Product Function Protein

Compound A and Compound B increase the level of nonsense suppression in the biochemical assay three to four fold over untreated extracts. To determine whether these compounds also function *in vivo*, a stable cell line harboring the UGA nonsense-containing luciferase gene was treated with each compound. Cells were grown in standard medium supplemented with 1% penicillin-streptomycin (P/S) and 10% fetal bovine serum (FBS) to 70% confluence and split 1:1 the day before treatment. On the following day, cells were trypsinized and 40,000 cells were added to each well of a 96-well tissue culture dish. Serial dilutions of each compound were prepared to generate a six-point dose response curve spanning 2 logs (30 μ M to 0.3 μ M). The final concentration of the DMSO solvent remained constant at 1% in each well. Cells treated with 1% DMSO served as the background standard, and cells treated with gentamicin served as a positive control. As shown in Figure 6, these two compounds are more potent and efficacious than gentamicin at these concentrations.

Cells were transiently transfected with plasmids harboring the UGA, UAA or UAG nonsense alleles of luciferase in each codon context (UGAA, UGAC, UGAG, UGAU, UAGA, UAGC, UAGG, UAGU, UAAA, UAAC, UAAG, and UAAU) then the cells were treated overnight with Compound A, and gentamicin. The following day, the level of suppression was determined by measuring the amount of luminescence produced. The fold suppression above control cells treated with solvent was calculated and is numerically reported. The results are presented in Table 2 and Figure 6B.

Table 2

| Context | Gentamicin 3mg/ml | Compound A 10 μ M |
|---------|----------------------|--------------------------|
| UAAA | 0.71 | 0.17 |
| UAAC | 2.32 | 0.67 |
| UAAG | 0.01 | 0.02 |
| UAAU | 0.92 | 0.33 |
| UAGA | 1.31 | 0.64 |
| UAGC | 2.16 | 3.05 |
| UAGG | 0.64 | 0.51 |
| UAGU | 0.54 | 0.31 |
| UGAA | 0.76 | 0.4 |
| UGAC | 1.91 | 2.96 |
| UGAG | 0.45 | 0.23 |
| UGAU | 6.74 | 1.67 |

7.5. Compound A Alters the Accessibility of the Chemical Modifying Agents to Specific Nucleotides in the 28S rRNA

Previous studies have demonstrated that gentamicin and other members of the aminoglycoside family that decrease the fidelity of translation bind to the A site of the 16S rRNA. By chemical footprinting, UV cross-linking and NMR, gentamicin has been shown to bind at the A site (comprised of nucleotides 1400-1410 and 1490-1500, *E. coli* numbering) of the rRNA at nucleotides 1406, 1407, 1494, and 1496 (Moazed & Noller, 5 1987, *Nature* 327(6121):389-394; Woodcock *et al.*, 1991, *EMBO J.* 10(10):3099-3103; and Schroeder *et al.*, 2000). These observations prompted us to determine whether similar experiments could provide information on the mechanism of action of Compound A. To do 10 this, ribosomes prepared from HeLa cells were incubated with the small molecules (at a concentration of 100 μ M), followed by treatment with chemical modifying agents (dimethyl sulfate [DMS] and kethoxal [KE]). Following chemical modification, rRNA was phenol-chloroform extracted, ethanol precipitated, analyzed in primer extension reactions using end-labeled oligonucleotides hybridizing to different regions of the three rRNAs and resolved on 6% polyacrylamide gels. The probes used for primer extension cover the entire 15 18S (7 oligonucleotide primers), 28S (24 oligonucleotide primers), and 5S (one primer) rRNAs. Controls in these experiments include DMSO (a control for changes in rRNA accessibility induced by DMSO), paromomycin (a marker for 18S rRNA binding), and anisomycin (a marker for 28S rRNA binding).

The results of these foot-printing experiments indicated that Compound A 20 alters the accessibility of the chemical modifying agents to specific nucleotides in the 28S rRNA. More specifically, the regions protected by Compound A include: (1) a conserved region in the vicinity of the peptidyl transferase center (domain V) implicated in peptide bond formation (see Figure 7A) and (2) a conserved region in domain II that may interact with the peptidyl transferase center based on binding of vernamycinin B to both these areas 25 (Vannuffel *et al.*, 1994, *Nucleic Acids Res.* 22(21):4449-4453; see Figure 7B).

7.6. Compound A Causes Readthrough of Premature Termination Codons in Cell-based Disease Models

To address the effects of the nonsense-suppressing compounds on mRNAs altered in specific inherited diseases, a bronchial epithelial cell line harboring a nonsense 30 codon at amino acid 1282 (W1282X) was treated with Compound A (20 μ M) and CFTR function was monitored as a cAMP-activated chloride channel using the SPQ assay (Yang *et al.*, 1993, *Hum Mol Genet.* 2(8):1253-1261 and Howard *et al.*, 1996, *Nat Med.* 2(4):467-469). These experiments showed that cAMP treatment of these cells resulted in an increase in SPQ fluorescence, consistent with stimulation of CFTR-mediated halide efflux

(Figure 8). No increase in fluorescence was observed when cells were not treated with compound or if the cells were not stimulated with cAMP. These results indicate that the full-length CFTR expressed from this nonsense-containing allele following compound treatment also functions as a cAMP-stimulated anion channel, thus demonstrating that 5 cystic fibrosis cell lines increase chloride channel activity when treated with Compound A.

7.7. Primary Cells from the mdx Nonsense-containing Mouse Express Full-length Dystrophin Protein When Treated with Compound A

The mutation in the mdx mouse that premature termination of the 427 kDa dystrophin polypeptide has been shown to be a C to T transition at position 3185 in exon 23 (Sicinski *et al.*, 1989, *Science*. 244(4912):1578-1580). Mouse primary skeletal muscle cultures derived from 1-day old mdx mice were prepared as described previously (Barton-Davis *et al.*, 1999, *J Clin Invest*. 104(4):375-381). Cells were cultured for 10 days in the presence of Compound A (20 μ M). Culture medium was replaced every four days and the presence of dystrophin in myoblast cultures was detected by immunostaining as 10 described previously (Barton-Davis *et al.*, 1999, *J Clin Invest*. 104(4):375-381). A primary monoclonal antibody to the C-terminus of the dystrophin protein (F19A12) was used undiluted and rhodamine conjugated anti-mouse IgG was used as the secondary antibody. The F19A12 antibody will detect the full-length protein produced by suppression of the 15 nonsense codon. Staining was viewed using a Leica DMR microscope, digital camera, and associated imaging software at the University of Pennsylvania. As shown in Figure 9, 20 full-length dystrophin protein is produced and localized to the muscle myotubes in cultures treated with 20 μ M Compound A and gentamicin (200 μ M). In addition, cells from untreated cultures exhibited minimal staining. These results indicate that full-length dystrophin protein is produced as a consequence of nonsense suppression.

7.8. Compound A and Compound B Cause Readthrough of Premature Termination Codons in the mdx Mouse

Since the results of the mdx cell culture experiments demonstrated production of full-length dystrophin in cells treated with Compound A, it was asked whether suppression of the nonsense codon in the mdx mouse could be observed. As 30 previously described (Barton-Davis *et al.*, 1999, *J Clin Invest*. 104(4):375-381), compound was delivered by Alzet osmotic pumps implanted under the skin of anesthetized mice. Two doses of Compound A were administered. Gentamicin served as a positive control and pumps filled with solvent only served as the negative control. Pumps were loaded with appropriate compound such that the calculated doses to which tissue was exposed were 10

μM and 20 μM . The gentamicin concentration was calculated to achieve tissue exposure of approximately 200 μM . In the initial experiment, mice were treated for 14 days, after which animals were anesthetized with ketamine and exsanguinated. The tibialis anterior (TA) muscle of the experimental animals was then excised, frozen, and used for immunofluorescence analysis of dystrophin incorporation into striated muscle. The presence of dystrophin in TA muscles was detected by immunostaining, as described previously (Barton-Davis *et al.*, 1999, *J Clin Invest.* 104(4):375-381; see mdx primary cells in Section 7.8 *supra*). As shown in Figure 10, these experiments demonstrated that mice treated with both concentrations of compound elicited production of full-length dystrophin. Importantly, a significant portion of the full-length dystrophin protein was properly localized to the membrane. These important results demonstrate that Compound A can function in an animal model.

8. HUMAN DISEASE GENES SORTED BY CHROMOSOME

Table 3: Genes, Locations and Genetic Disorders on Chromosome 1

| Gene | GDB Accession ID | OMIM Link |
|---------|------------------|--|
| ABCA4 | GDB:370748 | MACULAR DEGENERATION, SENILE STARGARDT DISEASE 1; STGD1 ATP BINDING CASSETTE TRANSPORTER; ABCR RETINITIS PIGMENTOSA-19; RP19 |
| ABCD3 | GDB:131485 | PEROXISOMAL MEMBRANE PROTEIN 1; PXMP1 |
| ACADM | GDB:118958 | ACYL-CoA DEHYDROGENASE, MEDIUM-CHAIN; ACADM |
| AGL | GDB:132644 | GLYCOGEN STORAGE DISEASE III |
| AGT | GDB:118750 | ANGIOTENSIN I; AGT |
| ALDH4A1 | GDB:9958827 | HYPERPROLINEMIA, TYPE II |

| | | |
|---------|-------------|--|
| ALPL | GDB:118730 | PHOSPHATASE, LIVER ALKALINE; ALPL HYPOPHOSPHATASIA, INFANTILE |
| AMPD1 | GDB:119677 | ADENOSINE MONOPHOSPHATE DEAMINASE-1; AMPD1 |
| APOA2 | GDB:119685 | APOLIPOPROTEIN A-II; APOA2 |
| AVSD1 | GDB:265302 | ATRIOVENTRICULAR SEPTAL DEFECT; AVSD |
| BRCD2 | GDB:9955322 | BREAST CANCER, DUCTAL, 2; BRCD2 |
| C1QA | GDB:119042 | COMPLEMENT COMPONENT 1, q SUBCOMPONENT, ALPHA POLYPEPTIDE; C1QA |
| C1QB | GDB:119043 | COMPLEMENT COMPONENT 1, q SUBCOMPONENT, BETA POLYPEPTIDE; C1QB |
| C1QG | GDB:128132 | COMPLEMENT COMPONENT 1, q SUBCOMPONENT, GAMMA POLYPEPTIDE; C1QG |
| C8A | GDB:119735 | COMPLEMENT COMPONENT-8, DEFICIENCY OF |
| C8B | GDB:119736 | COMPLEMENT COMPONENT-8, DEFICIENCY OF, TYPE II |
| CACNA1S | GDB:126431 | CALCIUM CHANNEL, VOLTAGE-DEPENDENT, L TYPE, ALPHA 1S SUBUNIT; CACNA1S PERIODIC PARALYSIS I MALIGNANT HYPERTHERMIA SUSCEPTIBILITY-5; MHSS |

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| CCV | GDB:1336655 | CATARACT, CONGENITAL, VOLKMANN TYPE; CCV |
| CD3Z | GDB:119766 | CD3Z ANTIGEN, ZETA POLYPEPTIDE; CD3Z |
| CDC2L1 | GDB:127827 | PROTEIN KINASE p58; PK58 |
| CHML | GDB:135222 | CHOROIDEREMIA-LIKE; CHML |
| CHS1 | GDB:4568202 | CHEDIAK-HIGASHI SYNDROME; CHS1 |
| CIAS1 | GDB:9957338 | COLD HYPERSENSITIVITY URTICARIA, DEAFNESS, AND AMYLOIDOSIS |
| | | |
| CLCNKB | GDB:698472 | CHLORIDE CHANNEL, KIDNEY, B; CLCNKB |
| CMD1A | GDB:434478 | CARDIOMYOPATHY, DILATED 1A; CMD1A |
| CMH2 | GDB:137324 | CARDIOMYOPATHY, FAMILIAL HYPERTROPHIC, 2; CMH2 |
| CMM | GDB:119059 | MELANOMA, MALIGNANT |
| COL11A1 | GDB:120595 | COLLAGEN, TYPE XI, ALPHA-1; COL11A1 |
| | | |
| COL9A2 | GDB:138310 | COLLAGEN, TYPE IX, ALPHA-2 CHAIN; |

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| | | COL9A2 EPIPHYSEAL DYSPLASIA, MULTIPLE, 2; EDM2 |
| CPT2 | GDB:127272 | MYOPATHY WITH DEFICIENCY OF CARNITINE PALMITOYLTRANSFERASE II HYPOGLYCEMIA, HYPOKETOTIC, WITH DEFICIENCY OF CARNITINE PALMITOYLTRANSFERASE CARNITINE PALMITOYLTRANSFERASE II; CPT2 |
| CRB1 | GDB:333930 | RETINITIS PIGMENTOSA-12; RP12 |
| | | |
| CSE | GDB:596182 | CHOREOATHETOSIS/SPASTICITY, EPISODIC; CSE |
| CSF3R | GDB:126430 | COLONY STIMULATING FACTOR 3 RECEPTOR, GRANULOCYTE; CSF3R |
| CTPA | GDB:9863168 | CATARACT, POSTERIOR POLAR |
| CTSK | GDB:453910 | PYCNODYSOSTOSIS CATHEPSIN K; CTSK |
| DBT | GDB:118784 | MAPLE SYRUP URINE DISEASE, TYPE 2 |
| DIO1 | GDB:136449 | THYROXINE DEIODINASE TYPE I; TDXII |
| DISC1 | GDB:9992707 | DISORDER-2; SCZD2 |
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| DPYD | GDB:364102 | DIHYDROPYRIMIDINE DEHYDROGENASE; DPYD |
| EKV | GDB:119106 | ERYTHROKERATODERMIA VARIABILIS; EKV |
| ENO1 | GDB:119871 | PHOSPHOPYRUVATE HYDRATASE; PPH |
| ENO1P | GDB:135006 | PHOSPHOPYRUVATE HYDRATASE; PPH |
| EPB41 | GDB:119865 | ERYTHROCYTE MEMBRANE PROTEIN BAND 4.1; EPB41 HEREDITARY HEMOLYTIC |
| EPHX1 | GDB:119876 | EPOXIDE HYDROLASE 1, MICROSOMAL; EPHX1 |
| F13B | GDB:119893 | FACTOR XIII, B SUBUNIT; F13B |
| F5 | GDB:119896 | FACTOR V DEFICIENCY |
| FCGR2A | GDB:119903 | Fc FRAGMENT OF IgG, LOW AFFINITY IIa, RECEPTOR FOR; FCGR2A |
| FCGR2B | GDB:128183 | Fc FRAGMENT OF IgG, LOW AFFINITY IIa, RECEPTOR FOR; FCGR2A |
| FCGR3A | GDB:119904 | Fc FRAGMENT OF IgG, LOW AFFINITY IIIa, RECEPTOR FOR; FCGR3A |
| FCHL | GDB:9837503 | HYPERLIPIDEMIA, COMBINED |
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| FH | GDB:119133 | FUMARATE HYDRATASE; FH LEIOMYOMATA, HEREDITARY MULTIPLE, OF SKIN |
| FMO3 | GDB:135136 | FLAVIN-CONTAINING MONOOXYGENASE 3; FMO3 TRIMETHYLAMINURIA |
| FMO4 | GDB:127981 | FLAVIN-CONTAINING MONOOXYGENASE 2; FMO2 |
| FUCA1 | GDB:119237 | FUCOSIDOSIS |
| FY | GDB:119242 | BLOOD GROUP--DUFFY SYSTEM; Fy |
| GALE | GDB:119245 | GALACTOSE EPIMERASE DEFICIENCY |
| GBA | GDB:119262 | GAUCHER DISEASE, TYPE I; GD I |
| | | |
| GFND | GDB:9958222 | GLOMERULAR NEPHRITIS, FAMILIAL, WITH FIBRONECTIN DEPOSITS |
| GJA8 | GDB:696369 | CATARACT, ZONULAR PULVERULENT 1; CZP1 GAP JUNCTION PROTEIN, ALPHA-8, 50-KD; GJA8 |
| GJB3 | GDB:127820 | ERYTHROKERATODERMIA VARIABILIS; EKV DEAFNESS, AUTOSOMAL DOMINANT NONSYNDROMIC SENSORINEURAL, 2; DFNA2 |
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| GLC3B | GDB:3801939 | GLAUCOMA 3, PRIMARY INFANTILE, |

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| | | B; GLC3B |
| HF1 | GDB:120041 | H FACTOR 1; HF1 |
| HMGCL | GDB:138445 | HYDROXYMETHYLGLUTARICACIDURIA; HMGCL |
| HPC1 | GDB:5215209 | PROSTATE CANCER; PRCA1 PROSTATE CANCER, HEREDITARY 1 |
| HRD | GDB:9862254 | HYPOPARATHYROIDISM WITH SHORT STATURE, MENTAL RETARDATION, AND SEIZURES |
| HRPT2 | GDB:125253 | HYPERTHYROIDISM, FAMILIAL PRIMARY, WITH MULTIPLE OSSIFYING JAW |
| HSD3B2 | GDB:134044 | ADRENAL HYPERPLASIA II |
| HSPG2 | GDB:126372 | HEPARAN SULFATE PROTEOGLYCAN OF BASEMENT MEMBRANE; HSPG2 MYOTONIC MYOPATHY, DWARFISM, CHONDRODYSTROPHY, AND OCULAR AND FACIAL |
| KCNQ4 | GDB:439046 | DEAFNESS, AUTOSOMAL DOMINANT NONSYNDROMIC SENSORINEURAL, 2; DFNA2 |
| | | |
| KCS | GDB:9848740 | KENNY-CAFFEY SYNDROME, RECESSIVE FORM |
| KIF1B | GDB:128645 | CHARCOT-MARIE-TOOTH DISEASE, NEURONAL TYPE, A; CMT2A |

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| LAMB3 | GDB:251820 | LAMININ, BETA 3; LAMB3 |
| LAMC2 | GDB:136225 | LAMININ, GAMMA 2; LAMC2 EPIDERMOLYSIS BULLOSA LETALIS |
| LGMD1B | GDB:231606 | MUSCULAR DYSTROPHY, LIMB-GIRDLE, TYPE 1B; LGMD1B |
| LMNA | GDB:132146 | LAMIN A/C; LMNA LIPODYSTROPHY, FAMILIAL PARTIAL, DUNNIGAN TYPE; LDP1 |
| LOR | GDB:132049 | LORICRIN; LOR |
| MCKD1 | GDB:9859381 | POLYCYSTIC KIDNEYS, MEDULLARY TYPE |
| MCL1 | GDB:139137 | MYELOID CELL LEUKEMIA 1; MCL1 |
| MPZ | GDB:125266 | HYPERTROPHIC NEUROPATHY OF DEJERINE-SOTTAS MYELIN PROTEIN ZERO; MPZ |
| MTHFR | GDB:370882 | 5,10-@METHYLENETETRAHYDROFOL ATE REDUCTASE; MTHFR |
| MTR | GDB:119440 | METHYLtetrahydrofolate:L-HO MOCysteine S-METHYLTRANSFERASE; MTR |
| MUTYH | GDB:9315115 | ADENOMATOUS POLYPOSIS OF THE COLON; APC |
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| MYOC | GDB:5584221 | GLAUCOMA 1, OPEN ANGLE; GLC1A MYOCILIN; MYOC |
| NB | GDB:9958705 | NEUROBLASTOMA; NB |
| NCF2 | GDB:120223 | GRANULOMATOUS DISEASE, CHRONIC, AUTOSOMAL CYTOCHROME-b-POSITIVE FORM |
| NEM1 | GDB:127387 | NEMALINE MYOPATHY 1, AUTOSOMAL DOMINANT; NEM1 |
| NPHS2 | GDB:9955617 | ARRHYTHMOGENIC RIGHT VENTRICULAR DYSPLASIA, FAMILIAL, 2; ARVD2 |
| NPPA | GDB:118727 | NATRIURETIC PEPTIDE PRECURSOR A; NPPA |
| NRAS | GDB:119457 | ONCOGENE NRAS; NRAS; NRAS1 |
| NTRK1 | GDB:127897 | ONCOGENE TRK NEUROTROPHIC TYROSINE KINASE, RECEPTOR, TYPE 1; NTRK1 NEUROPATHY, CONGENITAL SENSORY, WITH ANHIDROSIS |
| OPTA2 | GDB:9955577 | OSTEOPETROSIS, AUTOSOMAL DOMINANT, TYPE II; OPA2 |
| PBX1 | GDB:125351 | PRE-B-CELL LEUKEMIA TRANSCRIPTION FACTOR-1; PBX1 |
| PCHC | GDB:9955586 | PHEOCHROMOCYTOMA |
| PGD | GDB:119486 | 6-@PHOSPHOGLUCONATE DEHYDROGENASE, ERYTHROCYTE |

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| PHA2A | GDB:9955628 | PSEUDOHYPOALDOSTERONISM, TYPE II; PHA2 |
| PHGDH | GDB:9958261 | 3-@PHOSPHOGLYCERATE DEHYDROGENASE DEFICIENCY |
| PKLR | GDB:120294 | PYRUVATE KINASE DEFICIENCY OF ERYTHROCYTE |
| PKP1 | GDB:4249598 | PLAKOPHILIN 1; PKP1 |
| PLA2G2A | GDB:120296 | PHOSPHOLIPASE A2, GROUP IIA; PLA2G2A |
| PLOD | GDB:127821 | PROCOLLAGEN-LYSINE, 2-OXOGLUTARATE 5-DIOXYGENASE; PLOD EHRLERS-DANLOS SYNDROME, TYPE VI; E-D VI; EDS VI |
| PPOX | GDB:118852 | PROTOPORPHYRINOGEN OXIDASE; PPOX |
| PPT | GDB:125227 | CEROID-LIPOFUSCINOSIS, NEURONAL 1, INFANTILE; CLN1 PALMITOYL-PROTEIN THIOESTERASE; PPT |
| PRCC | GDB:3888215 | PAPILLARY RENAL CELL CARCINOMA; PRCC |
| PRG4 | GDB:9955719 | ARTHROPATHY-CAMPTODACTYLY SYNDROME |
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| PSEN2 | GDB:633044 | ALZHEIMER DISEASE, FAMILIAL, TYPE 4; AD4 |

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| PTOS1 | GDB:6279920 | PTOSIS, HEREDITARY CONGENITAL 1; PTOS1 |
| REN | GDB:120345 | RENIN; REN |
| RFX5 | GDB:6288464 | REGULATORY FACTOR 5; RFX5 |
| RHD | GDB:119551 | RHESUS BLOOD GROUP, D ANTIGEN; RHD |
| RMD1 | GDB:448902 | RIPPLING MUSCLE DISEASE-1; RMD1 |
| RPE65 | GDB:226519 | RETINAL PIGMENT EPITHELIUM-SPECIFIC PROTEIN, 65-KD; RPE65 AMAUROSIS CONGENITA OF LEBER II |
| SCCD | GDB:9955558 | CORNEAL DYSTROPHY, CRYSTALLINE, OF SCHNYDER |
| SERPINC1 | GDB:119024 | ANTITHROMBIN III DEFICIENCY |
| SJS1 | GDB:1381631 | MYOTONIC MYOPATHY, DWARFISM, CHONDRODYSTROPHY, AND OCULAR AND FACIAL |
| SLC19A2 | GDB:9837779 | THIAMINE-RESPONSIVE MEGALOBLASTIC ANEMIA SYNDROME |
| SLC2A1 | GDB:120627 | SOLUTE CARRIER FAMILY 2, MEMBER 1; SLC2A1 |
| SPTA1 | GDB:119601 | ELLIPTOCYTOSIS, RHESUS-UNLINKED TYPE HEREDITARY HEMOLYTIC SPECTRIN, ALPHA, ERYTHROCYTIC 1; SPTA1 |

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| TAL1 | GDB:120759 | T-CELL ACUTE LYMPHOCYTIC LEUKEMIA 1; TAL1 |
| TNFSF6 | GDB:422178 | APOPTOSIS ANTIGEN LIGAND 1; APT1LG1 |
| TNNT2 | GDB:221879 | TROPONIN-T2, CARDIAC; TNNT2 |
| TPM3 | GDB:127872 | ONCOGENE TRK TROPOMYOSIN 3; TPM3 |
| TSHB | GDB:120467 | THYROID-STIMULATING HORMONE, BETA CHAIN; TSHB |
| UMPK | GDB:120481 | URIDINE MONOPHOSPHATE KINASE; UMPK |
| UOX | GDB:127539 | URATE OXIDASE; UOX |
| UROD | GDB:119628 | PORPHYRIA CUTANEA TARDA; PCT |
| USH2A | GDB:120483 | USHER SYNDROME, TYPE II; USH2 |
| VMGLOM | GDB:9958134 | GLOMUS TUMORS, MULTIPLE |
| VWS | GDB:120532 | CLEFT LIP AND/OR PALATE WITH MUCOUS CYSTS OF LOWER LIP |
| WS2B | GDB:407579 | WAARDENBURG SYNDROME, TYPE 2B; WS2B |

Table 4: Genes, Locations and Genetic Disorders on Chromosome 2

| Gene | GDB Accession ID | Location | OMIM Link |
|------|------------------|----------|-----------|
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|--------|--------------|---|---|
| ABCB11 | GDB:9864786 | 2q24-2q24 2q24.3-2q24.3 | CHOLESTASIS, PROGRESSIVE FAMILIAL INTRAHEPATIC 2; PFIC2 |
| ABCG5 | GDB:10450298 | 2p21-2p21 | PHYTOSTEROLEMIA |
| ABCG8 | GDB:10450300 | 2p21-2p21 | PHYTOSTEROLEMIA |
| ACADL | GDB:118745 | 2q34-2q35 | ACYL-CoA DEHYDROGENASE, LONG-CHAIN, DEFICIENCY OF |
| ACP1 | GDB:118962 | 2p25-2p25 | PHOSPHATASE, ACID, OF ERYTHROCYTE; ACP1 |
| AGXT | GDB:127113 | 2q37.3-2q37.3 | OXALOSIS I |
| AHHR | GDB:118984 | 2pter-2q31 | CYTOCHROME P450, SUBFAMILY I, POLYPEPTIDE 1; CYP1A1 |
| ALMS1 | GDB:9865539 | 2p13-2p12 2p14-2p13 2p13.1-2p13.1 | ALSTROM SYNDROME |
| ALPP | GDB:119672 | 2q37.1-2q37.1 | ALKALINE PHOSPHATASE, PLACENTAL; ALPP |
| ALS2 | GDB:135696 | 2q33-2q35 | AMYOTROPHIC LATERAL SCLEROSIS 2, JUVENILE; ALS2 |
| APOB | GDB:119686 | 2p24-2p23 2p24-2p24 | APOLIPOPROTEIN B; APOB |
| BDE | GDB:9955730 | 2q37-2q37 | BRACHYDACTYLY, TYPE E; BDE |
| BDMR | GDB:533064 | 2q37-2q37 | BRACHYDACTYLY-MENTAL |

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| | | | RETARDATION SYNDROME; BDMR |
| BJS | GDB:9955717 | 2q34-2q36 | TORTI AND NERVE DEAFNESS |
| BMPR2 | GDB:642243 | 2q33-2q33 2q33-2q34 | PULMONARY HYPERTENSION, PRIMARY; PPH1 BONE MORPHOGENETIC RECEPTOR TYPE II; BMPR2 |
| CHRNA1 | GDB:120586 | 2q24-2q32 | CHOLINERGIC RECEPTOR, NICOTINIC, ALPHA POLYPEPTIDE 1; CHRNA1 |
| CMCWTD | GDB:11498919 | 2p22.3-2p21 | FAMILIAL CHRONIC MUCCOCUTANEOUS, DOMINANT TYPE |
| CNGA3 | GDB:434398 | 2q11.2-2q11.2 | COLORBLINDNESS, TOTAL CYCLIC NUCLEOTIDE GATED CHANNEL, OLFACTORY, 3; CNG3 |
| COL3A1 | GDB:118729 | 2q31-2q32.3 2q32.2-2q32.2 | COLLAGEN, TYPE III; COL3A1 EHLERS-DANLOS SYNDROME, TYPE IV, AUTOSOMAL DOMINANT |
| COL4A3 | GDB:128351 | 2q36-2q37 | COLLAGEN, TYPE IV, ALPHA-3 CHAIN; COL4A3 |
| COL4A4 | GDB:132673 | 2q35-2q37 | COLLAGEN, TYPE IV, ALPHA-4 CHAIN; COL4A4 |
| COL6A3 | GDB:119066 | 2q37.3-2q37.3 | COLLAGEN, TYPE VI, ALPHA-3 CHAIN; COL6A3 MYOPATHY, BENIGN CONGENITAL, WITH CONTRACTURES |

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| CPS1 | GDB:119799 | 2q33-2q36 2q34-2q35 2q35-2q35 | HYPERAMMONEMIA DUE TO CARBAMOYLPHOSPHATE SYNTHETASE I DEFICIENCY |
| CRYGA | GDB:119076 | 2q33-2q35 | CRYSTALLIN, GAMMA A; CRYGA |
| CRYGEP1 | GDB:119808 | 2q33-2q35 | CRYSTALLIN, GAMMA A; CRYGA |
| CYP1B1 | GDB:353515 | 2p21-2p21 2p22-2p21 2pter-2qter | GLAUCOMA 3, PRIMARY INFANTILE, A; GLC3A CYTOCHROME P450, SUBFAMILY I (DIOXIN-INDUCIBLE), POLYPEPTIDE 1; CYP1B1 |
| CYP27A1 | GDB:128129 | 2q33-2qter | CEREBROTENDINOUS XANTHOMATOSIS |
| DBI | GDB:119837 | 2q12-2q21 | DIAZEPAM BINDING INHIBITOR; DBI |
| DES | GDB:119841 | 2q35-2q35 | DESMIN; DES |
| DYSF | GDB:340831 | 2p-2p 2p13-2p13 2pter-2p12 | MUSCULAR DYSTROPHY, LIMB-GIRDLE, TYPE 2B; LGMD2B MUSCULAR DYSTROPHY, LATE-ONSET DISTAL |
| EDAR | GDB:9837372 | 2q11-2q13 | DYSPLASIA, HYPOHIDROTIC ECTODERMAL DYSPLASIA, ANHYDROTIC |
| EFEMP1 | GDB:1220111 | 2p16-2p16 | DOYNE HONEYCOMB DEGENERATION OF RETINA FIBRILLIN-LIKE; FBNL |

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|---------|-------------|---------------|--|
| EIF2AK3 | GDB:9956743 | 2p12-2p12 | EPIPHYSEAL DYSPLASIA, MULTIPLE, WITH EARLY-ONSET DIABETES MELLITUS |
| ERCC3 | GDB:119881 | 2q21-2q21 | EXCISION-REPAIR, COMPLEMENTING DEFECTIVE, IN CHINESE HAMSTER, 3; ERCC3 |
| FSHR | GDB:127510 | 2p21-2p16 | FOLLICLE-STIMULATING HORMONE RECEPTOR; FSHR GONADAL DYSGENESIS, XX TYPE |
| GAD1 | GDB:119244 | 2q31-2q31 | PYRIDOXINE DEPENDENCY WITH SEIZURES |
| GINGF | GDB:9848875 | 2p21-2p21 | GINGIVAL SON OF SEVENLESS (DROSOPHILA) HOMOLOG 1; SOS1 |
| GLC1B | GDB:1297553 | 2q1-2q13 | GLAUCOMA 1, OPEN ANGLE, B; GLC1B |
| GPD2 | GDB:354558 | 2q24.1-2q24.1 | GLYCEROL-3-PHOSPHATE DEHYDROGENASE-2; GPD2 |
| GYPC | GDB:120027 | 2q14-2q21 | BLOOD GROUP--GERBICH; Ge |
| HADHA | GDB:434026 | 2p23-2p23 | HYDROXYACYL-CoA DEHYDROGENASE/3-KETOACY L-CoA THIOLASE/ENOYL-CoA HYDRATASE, |
| HADHB | GDB:344953 | 2p23-2p23 | HYDROXYACYL-CoA DEHYDROGENASE/3-KETOACY L-CoA THIOLASE/ENOYL-CoA HYDRATASE, |

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| HOXD13 | GDB:127225 | 2q31-2q31 | HOMEOPBOX-D13; HOXD13 SYNDACTYLY, TYPE II |
| HPE2 | GDB:136066 | 2p21-2p21 | MIDLINE CLEFT SYNDROME |
| IGKC | GDB:120088 | 2p12-2p12 2p11.2-2p11.2 | IMMUNOGLOBULIN KAPPA CONSTANT REGION; IGKC |
| IHH | GDB:511203 | 2q33-2q35 2q35-2q35 2pter-2qter | BRACHYDACTYLY, TYPE A1; BDA1 INDIAN HEDGEHOG, DROSOPHILA, HOMOLOG OF; IHH |
| IRS1 | GDB:133974 | 2q36-2q36 | INSULIN RECEPTOR SUBSTRATE 1; IRS1 |
| ITGA6 | GDB:128027 | 2pter-2qter | INTEGRIN, ALPHA-6; ITGA6 |
| KHK | GDB:391903 | 2p23.3-2p23.2 | FRUCTOSURIA |
| KYNU | GDB:9957925 | 2q22.2-2q23.3 | |
| LCT | GDB:120140 | 2q21-2q21 | DISACCHARIDE INTOLERANCE II |
| LHCGR | GDB:125260 | 2p21-2p21 | LUTEINIZING HORMONE/CHORIOGONADOTROPIN RECEPTOR; LHCGR |
| LSFC | GDB:9956219 | 2p16-2p16 | CYTOCHROME c OXIDASE DEFICIENCY, FRENCH-CANADIAN TYPE |
| MSH2 | GDB:203983 | 2p16-2p16 2p22-2p21 | COLON CANCER, FAMILIAL, NONPOLYPOSIS TYPE 1; FCC1 |
| MSH6 | GDB:632803 | 2p16-2p16 | G/T MISMATCH-BINDING |

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| | | | PROTEIN; GTBP |
| NEB | GDB:120224 | 2q24.1-2q24.2 | NEBULIN; NEB NEMALINE MYOPATHY 2, AUTOSOMAL RECESSIVE; NEM2 |
| NMTC | GDB:11498336 | 2q21-2q21 | THYROID CARCINOMA, PAPILLARY |
| NPHP1 | GDB:128050 | 2q13-2q13 | NEPHRONOPHTHISIS, FAMILIAL JUVENILE 1; NPHP1 |
| PAFAH1P1 | GDB:435099 | 2p11.2-2p11.2 | PLATELET-ACTIVATING FACTOR ACETYLHYDROLASE, GAMMA SUBUNIT |
| PAX3 | GDB:120495 | 2q36-2q36 2q35-2q35 | KLEIN-WAARDENBURG SYNDROME WAARDENBURG SYNDROME; WS1 |
| PAX8 | GDB:136447 | 2q12-2q14 | PAIRED BOX HOMEOTIC GENE 8; PAX8 |
| PMS1 | GDB:386403 | 2q31-2q33 | POSTMEIOTIC SEGREGATION INCREASED (S. CEREVIAE)-1; PMS1 |
| PNKD | GDB:5583973 | 2q33-2q35 | CHOREOATHETOSIS, FAMILIAL PAROXYSMAL; FPDI |
| PPH1 | GDB:1381541 | 2q31-2q32 2q33-2q33 | PULMONARY HYPERTENSION, PRIMARY; PPH1 |
| PROC | GDB:120317 | 2q13-2q21 2q13-2q14 | PROTEIN C DEFICIENCY, CONGENITAL THROMBOTIC DISEASE DUE TO |
| REG1A | GDB:132455 | 2p12-2p12 | REGENERATING |

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| | | | ISLET-DERIVED 1-ALPHA; REGIA |
| SAG | GDB:120365 | 2q37.1-2q37.1 | S-ANTIGEN; SAG |
| SFTPB | GDB:120374 | 2p12-2p11.2 | SURFACTANT-ASSOCIATED PROTEIN, PULMONARY-3; SFTP3 |
| SLC11A1 | GDB:371444 | 2q35-2q35 | CIRRHOSIS, PRIMARY; PBC NATURAL RESISTANCE-ASSOCIATED MACROPHAGE PROTEIN 1; NRAMP1 |
| SLC3A1 | GDB:202968 | 2p16.3-2p16.3 2p21-2p21 | SOLUTE CARRIER FAMILY 3, MEMBER 1; SLC3A1 CYSTINURIA; CSNU |
| SOS1 | GDB:230004 | 2p22-2p21 | GINGIVAL SON OF SEVENLESS (DROSOPHILA) HOMOLOG 1; SOS1 |
| SPG4 | GDB:230127 | 2p24-2p21 | SPASTIC PARAPLEGIA-4, AUTOSOMAL DOMINANT; SPG4 |
| SRD5A2 | GDB:127343 | 2p23-2p23 | PSEUDOVAGINAL PERINEOSCROTAL HYPOSPADIAS; PPSH |
| TCL4 | GDB:136378 | 2q34-2q34 | T-CELL LEUKEMIA/LYMPHOMA-4; TCL4 |
| TGFA | GDB:120435 | 2p13-2p13 | TRANSFORMING GROWTH FACTOR, ALPHA; TGFA |
| TMD | GDB:9837196 | 2q31-2q31 | TIBIAL MUSCULAR DYSTROPHY, TARDIVE |

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| TPO | GDB:120446 | 2p25-2p25 2p25-2p24 | THYROID HORMONOGENESIS, GENETIC DEFECT IN, IIA |
| UGT1 | GDB:120007 | 2q37-2q37 | UDP GLUCURONOSYLTRANSFERAS E 1 FAMILY, A1; UGT1A1 |
| UV24 | GDB:9955737 | 2pter-2qter | UV-DAMAGE, EXCISION REPAIR OF, UV-24 |
| WSS | GDB:9955707 | 2q32-2q32 | WRINKLY SKIN SYNDROME; WSS |
| XDH | GDB:266386 | 2p23-2p22 | XANTHINURIA |
| ZAP70 | GDB:433738 | 2q11-2q13 2q12-2q12 | SYK-RELATED TYROSINE KINASE; SRK |
| ZFHX1B | GDB:9958310 | 2q22-2q22 | DISEASE, MICROCEPHALY, AND IRIS COLOBOMA |

Table 5: Genes, Locations and Genetic Disorders on Chromosome 3

| Gene | GDB Accession ID | Location | OMIM Link |
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| ACAA1 | GDB:119643 | 3p23-3p22 | PEROXISOMAL 3-OXOACYL-COENZYME A THIOLASE DEFICIENCY |
| AGTR1 | GDB:132359 | 3q21-3q25 | ANGIOTENSIN II RECEPTOR, VASCULAR TYPE 1; AT2R1 |
| AHSG | GDB:118985 | 3q27-3q27 | ALPHA-2-HS-GLYCOPROTEIN; AHSG |
| AMT | GDB:132138 | 3p21.3-3p21.2 3p21.2-3p21.1 | HYPERGLYCINEMIA, ISOLATED NONKETOTIC, TYPE II; NKH2 |

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| ARP | GDB:9959049 | 3p21.1-3p21.1 | ARGININE-RICH PROTEIN |
| BBS3 | GDB:376501 | 3p-3p 3p12.3-3q11.1 | BARDET-BIEDL SYNDROME, TYPE 3; BBS3 |
| BCHE | GDB:120558 | 3q26.1-3q26.2 | BUTYRYLCHOLINESTERASE; BCHE |
| BCPM | GDB:433809 | 3q21-3q21 | BENIGN CHRONIC PEMPHIGUS; BCPM |
| BTD | GDB:309078 | 3p25-3p25 | BIOTINIDASE; BTD |
| CASR | GDB:134196 | 3q21-3q24 | HYPOCALCIURIC HYPERCALCEMIA, FAMILIAL; HHC1 |
| CCR2 | GDB:337364 | 3p21-3p21 | CHEMOKINE (C-C) RECEPTOR 2; CMKBR2 |
| CCR5 | GDB:1230510 | 3p21-3p21 | CHEMOKINE (C-C) RECEPTOR 5; CMKBR5 |
| CDL1 | GDB:136344 | 3q26.3-3q26.3 | DE LANGE SYNDROME; CDL |
| CMT2B | GDB:604021 | 3q13-3q22 | CHARCOT-MARIE-TOOTH DISEASE, NEURONAL TYPE, B; CMT2B |
| COL7A1 | GDB:128750 | 3p21-3p21 3p21.3-3p21.3 | COLLAGEN, TYPE VII, ALPHA-1; COL7A1 |
| CP | GDB:119069 | 3q23-3q25 3q21-3q24 | CERULOPLASMIN; CP |
| CRV | GDB:11498333 | 3p21.3-3p21.1 | VASCULOPATHY, RETINAL, WITH CEREBRAL |

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| | | | LEUKODYSTROPHY |
| CTNNB1 | GDB:141922 | 3p22-3p22 3p21.3-3p21.3 | CATENIN, BETA 1; CTNNB1 |
| DEM | GDB:681157 | 3p12-3q11 | DEMENTIA, FAMILIAL NONSPECIFIC; DEM |
| ETM1 | GDB:9732523 | 3q13-3q13 | TREMOR, HEREDITARY ESSENTIAL 1; ETM1 |
| FANCD2 | GDB:698345 | 3p25.3-3p25.3 3pter-3p24.2 | FANCONI PANCYTOPENIA, COMPLEMENTATION GROUP D |
| FIH | GDB:9955790 | 3q13-3q13 | HYPOPARATHYROIDISM, FAMILIAL ISOLATED; FIH |
| FOXL2 | GDB:129025 | 3q23-3q23 3q22-3q23 | BLEPHAROPHIMOSIS, EPICANTHUS INVERSUS, AND PTOSIS; BPES |
| GBE1 | GDB:138442 | 3p12-3p12 | GLYCOGEN STORAGE DISEASE IV |
| GLB1 | GDB:119987 | 3p22-3p21.33 3p21.33-3p21. 33 | GANGLIOSIDOSIS, GENERALIZED GM1, TYPE I |
| GLC1C | GDB:3801941 | 3q21-3q24 | GLAUCOMA 1, OPEN ANGLE, C; GLC1C |
| GNAI2 | GDB:120516 | 3p21.3-3p21.2 | GUANINE NUCLEOTIDE-BINDING PROTEIN, ALPHA-INHIBITING, POLYPEPTIDE-2; |
| GNAT1 | GDB:119277 | 3p21.3-3p21.2 | GUANINE NUCLEOTIDE-BINDING |

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| | | | PROTEIN, ALPHA-TRANSDUCING, POLYPEPTIDE |
| GP9 | GDB:126370 | 3pter-3qter | PLATELET GLYCOPROTEIN IX; GP9 |
| GPX1 | GDB:119282 | 3q11-3q12 3p21.3-3p21.3 | GLUTATHIONE PEROXIDASE; GPX1 |
| HGD | GDB:203935 | 3q21-3q23 | ALKAPTONURIA; AKU |
| HRG | GDB:120055 | 3q27-3q27 | HISTIDINE-RICH GLYCOPROTEIN; HRG; HRGP |
| ITIH1 | GDB:120107 | 3p21.2-3p21.1 | INTER-ALPHA-TRYPSIN INHIBITOR, HEAVY CHAIN-1; ITIH1; IATIH; ITIH |
| KNG | GDB:125256 | 3q27-3q27 | FLAUJEAC FACTOR DEFICIENCY |
| LPP | GDB:1391795 | 3q27-3q28 | LIM DOMAIN-CONTAINING PREFERRED TRANSLOCATION PARTNER IN LIPOMA; LPP |
| LRS1 | GDB:682448 | 3p21.1-3p14.1 | LARSEN SYNDROME, AUTOSOMAL DOMINANT; LRS1 |
| MCCC1 | GDB:135989 | 3q27-3q27 3q25-3q27 | BETA-METHYLCROTONYLGLY CINURIA I |
| MDS1 | GDB:250411 | 3q26-3q26 | MYELODYSPLASIA SYNDROME 1; MDS1 |
| MHS4 | GDB:574245 | 3q13.1-3q13.1 | HYPERTHERMIA SUSCEPTIBILITY-4; MHS4 |

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| MITF | GDB:214776 | 3p14.1-3p12 | MICROPHTHALMIA-ASSOCIATED TRANSCRIPTION FACTOR; MITF WAARDENBURG SYNDROME, TYPE II; WS2 |
| MLH1 | GDB:249617 | 3p23-3p22 3p21.3-3p21.3 | COLON CANCER, FAMILIAL, NONPOLYPOSIS TYPE 2; FCC2 |
| MYL3 | GDB:120218 | 3p21.3-3p21.2 | MYOSIN, LIGHT CHAIN, ALKALI, VENTRICULAR AND SKELETAL SLOW; MYL3 |
| MYMY | GDB:11500610 | 3p26-3p24.2 | DISEASE |
| OPA1 | GDB:118848 | 3q28-3q29 | OPTIC ATROPHY 1; OPA1 |
| PBX1 | GDB:125352 | 3q22-3q23 | PRE-B-CELL LEUKEMIA TRANSCRIPTION FACTOR-1; PBX1 |
| PCCB | GDB:119474 | 3q21-3q22 | GLYCINEMIA, KETOTIC, II |
| POU1F1 | GDB:129070 | 3p11-3p11 | POU DOMAIN, CLASS 1, TRANSCRIPTION FACTOR 1; POU1F1 |
| PPARG | GDB:1223810 | 3p25-3p25 | CANCER OF COLON PEROXISOME PROLIFERATOR ACTIVATED RECEPTOR, GAMMA; PPARG |
| PROS1 | GDB:120721 | 3p11-3q11 3p11.1-3q11.2 | PROTEIN S, ALPHA; PROS1 |
| PTHR1 | GDB:138128 | 3p22-3p21.1 | METAPHYSEAL CHONDRODYSPLASIA, MURK JANSEN TYPE PARATHYROID HORMONE RECEPTOR 1; PTHR1 |

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| RCA1 | GDB:230233 | 3p14.2-3p14.2 | RENAL CARCINOMA, FAMILIAL, ASSOCIATED 1; RCA1 |
| RHO | GDB:120347 | 3q21.3-3q24 | RHODOPSIN; RHO |
| SCA7 | GDB:454471 | 3p21.1-3p12 | SPINOCEBELLAR ATAXIA 7; SCA7 |
| SCLC1 | GDB:9955750 | 3p23-3p21 | SMALL-CELL CANCER OF THE LUNG; SCCL |
| SCN5A | GDB:132152 | 3p21-3p21 | SODIUM CHANNEL, VOLTAGE-GATED, TYPE V, ALPHA POLYPEPTIDE; SCN5A |
| SI | GDB:120377 | 3q25.2-3q26.2 | DISACCHARIDE INTOLERANCE I |
| SLC25A20 | GDB:6503297 | 3p21.31-3p21. 31 | CARNITINE-ACYLCARNITINE TRANSLOCASE; CACT |
| SLC2A2 | GDB:119995 | 3q26.2-3q27 3q26.1-3q26.3 | SOLUTE CARRIER FAMILY 2, MEMBER 2; SLC2A2 FANCONI-BICKEL SYNDROME; FBS |
| TF | GDB:120432 | 3q21-3q21 | TRANSFERRIN; TF |
| TGFBR2 | GDB:224909 | 3p22-3p22 3pter-3p24.2 | TRANSFORMING GROWTH FACTOR-BETA RECEPTOR, TYPE II; TGFBR2 |
| THPO | GDB:374007 | 3q26.3-3q27 | THROMBOPOIETIN; THPO |
| THRΒ | GDB:120731 | 3p24.1-3p22 3p24.3-3p24.3 | THYROID HORMONE RECEPTOR, BETA; THRΒ |

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| TKT | GDB:132402 | 3p14.3-3p14.3 | WERNICKE-KORSAKOFF SYNDROME |
| TM4SF1 | GDB:250815 | 3q21-3q25 | TUMOR-ASSOCIATED ANTIGEN L6; TAAL6 |
| TRH | GDB:128072 | 3pter-3qter | THYROTROPIN-RELEASING HORMONE DEFICIENCY |
| UMPS | GDB:120482 | 3q13-3q13 | OROTICACIDURIA I |
| UQCRC1 | GDB:141850 | 3p21.3-3p21.2 3p21.3-3p21.3 | UBIQUINOL-CYTOCHROME c REDUCTASE CORE PROTEIN I; UQCRC1 |
| USH3A | GDB:392645 | 3q21-3q25 | USHER SYNDROME, TYPE III; USH3 |
| VHL | GDB:120488 | 3p26-3p25 | VON HIPPEL-LINDAU SYNDROME; VHL |
| WS2A | GDB:128053 | 3p14.2-3p13 | MICROPHTHALMIA-ASSOCIATE D TRANSCRIPTION FACTOR; MITF WAARDENBURG SYNDROME, TYPE II; WS2 |
| XPC | GDB:134769 | 3p25.1-3p25.1 | XERODERMA PIGMENTOSUM, COMPLEMENTATION GROUP C; XPC |
| ZNF35 | GDB:120507 | 3p21-3p21 | ZINC FINGER PROTEIN-35; ZNF35 |

Table 6: Genes, Locations and Genetic Disorders on Chromosome 4

| Gene | GDB Accession ID | Location | OMIM Link |
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| ADH1B | GDB:119651 | 4q21-4q23 4q22-4q22 | ALCOHOL DEHYDROGENASE-2; ADH2 |
| ADH1C | GDB:119652 | 4q21-4q23 4q22-4q22 | ALCOHOL DEHYDROGENASE-3; ADH3 |
| AFP | GDB:119660 | 4q11-4q13 | ALPHA-FETOPROTEIN; AFP |
| AGA | GDB:118981 | 4q23-4q35 4q32-4q33 | ASPARTYLGLUCOSAMINURIA ; AGU |
| AIH2 | GDB:118751 | 4q11-4q13 4q13.3-4q21.2 | AMELOGENESIS IMPERFECTA 2, HYPOPLASTIC LOCAL, AUTOSOMAL DOMINANT; |
| ALB | GDB:118990 | 4q11-4q13 | ALBUMIN; ALB |
| ASMD | GDB:119705 | 4q-4q 4q28-4q31 | ANTERIOR SEGMENT OCULAR DYSGENESIS; ASOD |
| BFHD | GDB:11498907 | 4q34.1-4q35 | DYSPLASIA, BEUKES TYPE |
| CNGA1 | GDB:127557 | 4p14-4q13 | CYCLIC NUCLEOTIDE GATED CHANNEL, PHOTORECEPTOR, cGMP GATED, 1; CNCG1 |
| CRBM | GDB:9958132 | 4p16.3-4p16.3 | CHERUBISM |
| DCK | GDB:126810 | 4q13.3-4q21.1 | DEOXYCYTIDINE KINASE; DCK |
| DFNA6 | GDB:636175 | 4p16.3-4p16.3 | DEAFNESS, AUTOSOMAL DOMINANT NONSYNDROMIC SENSORINEURAL, 6; DFNA6 |
| DSPP | GDB:5560457 | 4pter-4qter 4q21.3-4q21.3 | DENTIN PHOSPHOPROTEIN; DPP DENTINOGENESIS |

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| | | | IMPERFECTA; DGI1 |
| DTDP2 | GDB:9955810 | 4q-4q | DENTIN DYSPLASIA, TYPE II |
| ELONG | GDB:11498700 | 4q24-4q24 | |
| ENAM | GDB:9955259 | 4q21-4q21 | AMELOGENESIS IMPERFECTA 2, HYPOPLASTIC LOCAL, AUTOSOMAL DOMINANT; AMELOGENESIS IMPERFECTA, HYPOPLASTIC TYPE |
| ETFDH | GDB:135992 | 4q32-4q35 | GLUTARICACIDURIA IIC; GA IIC |
| EVC | GDB:555573 | 4p16-4p16 | ELLIS-VAN CREVLD SYNDROME; EVC |
| F11 | GDB:119891 | 4q35-4q35 | PTA DEFICIENCY |
| FABP2 | GDB:119127 | 4q28-4q31 | FATTY ACID BINDING PROTEIN 2, INTESTINAL; FABP2 |
| FGA | GDB:119129 | 4q28-4q28 | AMYLOIDOSIS, FAMILIAL VISCERAL FIBRINOGEN, A ALPHA POLYPEPTIDE; FGA |
| FGB | GDB:119130 | 4q28-4q28 | FIBRINOGEN, B BETA POLYPEPTIDE; FGB |
| FGFR3 | GDB:127526 | 4p16.3-4p16.3 | ACHONDROPLASIA; ACH BLADDER CANCER FIBROBLAST GROWTH FACTOR RECEPTOR-3; FGFR3 |
| FGG | GDB:119132 | 4q28-4q28 | FIBRINOGEN, G GAMMA POLYPEPTIDE; FGG |

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| FSHMD1A | GDB:119914 | 4q35-4q35 | FACIOSCAPULOHUMERAL MUSCULAR DYSTROPHY 1A; FSHMD1A |
| GC | GDB:119263 | 4q12-4q13 4q12-4q12 | GROUP-SPECIFIC COMPONENT; GC |
| GNPTA | GDB:119280 | 4q21-4q23 | MUCOLIPIDOSIS II; ML2; ML II |
| GNRHR | GDB:136456 | 4q13-4q13 4q21.2-4q21.2 | GONADOTROPIN-RELEASING HORMONE RECEPTOR; GNRHR |
| GYPA | GDB:118890 | 4q28-4q31 4q28.2-4q31.1 | BLOOD GROUP--MN LOCUS; MN |
| HCA | GDB:9954675 | 4q33-4qter | HYPERCALCIURIA, FAMILIAL IDIOPATHIC |
| HCL2 | GDB:119305 | 4q28-4q31 4q-4q | HAIR COLOR-2; HCL2 |
| HD | GDB:119307 | 4p16.3-4p16.3 | HUNTINGTON DISEASE; HD |
| HTN3 | GDB:125601 | 4q12-4q21 | HISTATIN-3; HTN3 |
| HVBS6 | GDB:120687 | 4q32-4q32 | HEPATOCELLULAR CARCINOMA-2; HCC2 |
| IDUA | GDB:119327 | 4p16.3-4p16.3 | MUCOPOLYSACCHARIDOSIS TYPE I; MPS I |
| IF | GDB:120077 | 4q24-4q25 4q25-4q25 | COMPLEMENT COMPONENT-3 INACTIVATOR, DEFICIENCY OF |
| JPD | GDB:120113 | 4pter-4qter | PERIODONTITIS, JUVENILE; |

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| | | 4q12-4q13 | JPD |
| KIT | GDB:120117 | 4q12-4q12 | V-KIT HARDY-ZUCKERMAN 4 FELINE SARCOMA VIRAL ONCOGENE HOMOLOG; KIT |
| KLKB1 | GDB:127575 | 4q34-4q35 4q35-4q35 | FLETCHER FACTOR DEFICIENCY |
| LQT4 | GDB:682072 | 4q25-4q27 | SYNDROME WITHOUT PSYCHOMOTOR RETARDATION |
| MANBA | GDB:125261 | 4q21-4q25 | MANNOSIDOSIS, BETA; MANB1 |
| MLLT2 | GDB:136792 | 4q21-4q21 | MYELOID/LYMPHOID OR MIXED LINEAGE LEUKEMIA, TRANSLOCATED TO, 2; MLLT2 |
| MSX1 | GDB:120683 | 4p16.3-4p16.1 4p16.1-4p16.1 | MSH, DROSOPHILA, HOMEBOX, HOMOLOG OF, 1; MSX1 |
| MTP | GDB:228961 | 4q24-4q24 | MICROSOMAL TRIGLYCERIDE TRANSFER PROTEIN, 88 KD; MTP |
| NR3C2 | GDB:120188 | 4q31-4q31 4q31.1-4q31.1 | PSEUDOHYPOALDOSTERONISM, TYPE I, AUTOSOMAL RECESSIVE; PHA1 |
| PBT | GDB:120260 | 4q12-4q21 | PIEBALD TRAIT; PBT |
| PDE6B | GDB:125915 | 4p16.3-4p16.3 | NIGHTBLINDNESS, CONGENITAL STATIONARY; CSNB3 PHOSPHODIESTERASE 6B, cGMP-SPECIFIC, ROD, BETA; PDE6B |

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| PEE1 | GDB:7016765 | 4q31-4q34 4q25-4qter | I; PEE1 |
| PITX2 | GDB:134770 | 4q25-4q27 4q25-4q26 4q25-4q25 | IRIDOGONIODYSGENESIS, TYPE 2; IRID2 RIEGER SYNDROME, TYPE 1; RIEG1 RIEG BICOID-RELATED HOMEobox TRANSCRIPTION FACTOR 1; RIEG1 HOMEO BOX 2 |
| PKD2 | GDB:118851 | 4q21-4q23 | POLYCYSTIC KIDNEY DISEASE 2; PKD2 |
| QDPR | GDB:120331 | 4p15.3-4p15.3 4p15.31-4p15. 31 | PHENYLKETONURIA II |
| SGCB | GDB:702072 | 4q12-4q12 | MUSCULAR DYSTROPHY, LIMB-GIRDLE, TYPE 2E; LGMD2E |
| SLC25A4 | GDB:119680 | 4q35-4q35 | ADENINE NUCLEOTIDE TRANSLOCATOR 1; ANT1 PROGRESSIVE EXTERNAL OPHTHALMOPLEGIA; PEO |
| SNCA | GDB:439047 | 4q21.3-4q22 4q21-4q21 | SYNUCLEIN, ALPHA; SNCA PARKINSON DISEASE, FAMILIAL, TYPE 1; PARK1 |
| SOD3 | GDB:125291 | 4p16.3-4q21 | SUPEROXIDE DISMUTASE, EXTRACELLULAR; SOD3 |
| STATH | GDB:120391 | 4q11-4q13 | STATHERIN; STATH; STR |
| TAPVR1 | GDB:392646 | 4p13-4q11 | ANOMALOUS PULMONARY VENOUS RETURN; APVR |

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| TYS | GDB:119624 | 4q-4q | SCLEROTYLOSIS; TYS |
| WBS2 | GDB:132426 | 4q33-4q35.1 | WILLIAMS-BEUREN SYNDROME; WBS |
| WFS1 | GDB:434294 | 4p-4p 4p16-4p16 | DIABETES MELLITUS AND INSIPIDUS WITH OPTIC ATROPHY AND DEAFNESS |
| WHCR | GDB:125355 | 4p16.3-4p16.3 | WOLF-HIRSCHHORN SYNDROME; WHS |

Table 7: Genes, Locations and Genetic Disorders on Chromosome 5

| Gene | GDB Accession ID | OMIM Link |
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| ADAMTS2 | GDB:9957209 | EHLERS-DANLOS SYNDROME, TYPE VII, AUTOSOMAL RECESSIVE |
| ADRB2 | GDB:120541 | BETA-2-ADRENERGIC RECEPTOR; ADRB2 |
| AMCN | GDB:9836823 | ARTHROGRYPOSIS MULTIPLEX CONGENITA, NEUROGENIC TYPE |
| AP3B1 | GDB:9955590 | HERMANSKY-PUDLAK SYNDROME; HPS |
| APC | GDB:119682 | ADENOMATOUS POLYPOSIS OF THE COLON; APC |
| ARSB | GDB:119008 | MUCOPOLYSACCHARIDOSIS TYPE VI; MPS VI |
| B4GALT7 | GDB:9957653 | SYNDROME, PROGEROID FORM |

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| BHR1 | GDB:9956078 | ASTHMA |
| C6 | GDB:119045 | COMPLEMENT COMPONENT-6, DEFICIENCY OF |
| C7 | GDB:119046 | COMPLEMENT COMPONENT-7, DEFICIENCY OF |
| CCAL2 | GDB:5584265 | CHONDROCALCINOSIS, FAMILIAL ARTICULAR |
| CKN1 | GDB:128586 | COCKAYNE SYNDROME, TYPE I; CKN1 |
| CMDJ | GDB:9595425 | CRANIOMETAPHYSEAL DYSPLASIA, JACKSON TYPE; CMDJ |
| CRHBP | GDB:127438 | CORTICOTROPIN RELEASING HORMONE-BINDING PROTEIN; CRHBP |
| CSF1R | GDB:120600 | COLONY-STIMULATING FACTOR-1 RECEPTOR; CSF1R |
| DHFR | GDB:119845 | DIHYDROFOLATE REDUCTASE; DHFR |
| DIAPH1 | GDB:9835482 | DEAFNESS, AUTOSOMAL DOMINANT NONSYNDROMIC SENSORINEURAL, 1; DFNA1 DIAPHANOUS, DROSOPHILA, HOMOLOG OF, 1 |
| DTR | GDB:119853 | DIPHTHERIA TOXIN SENSITIVITY; DTS |
| EOS | GDB:9956083 | EOSINOPHILIA, FAMILIAL |

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| ERVR | GDB:9835857 | HYALOIDEORETINAL DEGENERATION OF WAGNER |
| F12 | GDB:119892 | HAGEMAN FACTOR DEFICIENCY |
| FBN2 | GDB:128122 | CONTRACTURAL ARACHNODACTYLY, CONGENITAL; CCA |
| GDNF | GDB:450609 | GLIAL CELL LINE-DERIVED NEUROTROPHIC FACTOR; GDNF |
| GHR | GDB:119984 | GROWTH HORMONE RECEPTOR; GHR |
| GLRA1 | GDB:118801 | GLYCINE RECEPTOR, ALPHA-1 SUBUNIT; GLRA1 KOK DISEASE |
| GM2A | GDB:120000 | TAY-SACHS DISEASE, AB VARIANT |
| HEXB | GDB:119308 | SANDHOFF DISEASE |
| HSD17B4 | GDB:385059 | 17-@BETA-HYDROXYSTEROID DEHYDROGENASE IV; HSD17B4 |
| ITGA2 | GDB:128031 | INTEGRIN, ALPHA-2; ITGA2 |
| KFS | GDB:9958987 | VERTEBRAL FUSION |
| LGMD1A | GDB:118832 | MUSCULAR DYSTROPHY, LIMB-GIRDLE, TYPE 1A; LGMD1A |
| LOX | GDB:119367 | LYSYL OXIDASE; LOX |
| LTC4S | GDB:384080 | LEUKOTRIENE C4 SYNTHASE; LTC4S |

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| MAN2A1 | GDB:136413 | MANNOSIDASE, ALPHA, II; MANA2 DYSERYTHROPOIETIC ANEMIA, CONGENITAL, TYPE II |
| MCC | GDB:128163 | MUTATED IN COLORECTAL CANCERS; MCC |
| MCCC2 | GDB:135990 | II |
| MSH3 | GDB:641986 | MutS, E. COLI, HOMOLOG OF, 3; MSH3 |
| MSX2 | GDB:138766 | MSH (DROSOPHILA) HOME BOX HOMOLOG 2; MSX2 pariETAL FORAMINA, SYMMETRIC; PFM |
| NR3C1 | GDB:120017 | GLUCOCORTICOID RECEPTOR; GRL |
| PCSK1 | GDB:128033 | PROPROTEIN CONVERTASE SUBTILISIN/KEXIN TYPE 1; PCSK1 |
| PDE6A | GDB:120265 | PHOSPHODIESTERASE 6A, cGMP-SPECIFIC, ROD, ALPHA; PDE6A |
| PFBI | GDB:9956096 | INTENSITY OF INFECTION IN |
| RASA1 | GDB:120339 | RAS p21 PROTEIN ACTIVATOR 1; RASA1 |
| SCZD1 | GDB:120370 | DISORDER-1; SCZD1 |
| SDHA | GDB:378037 | SUCCINATE DEHYDROGENASE COMPLEX, SUBUNIT A, FLAVOPROTEIN; SDHA |
| SGCD | GDB:5886421 | SARCOGLYCAN, DELTA; SGCD |

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| SLC22A5 | GDB:9863277 | CARNITINE DEFICIENCY, SYSTEMIC, DUE TO DEFECT IN RENAL REABSORPTION |
| SLC26A2 | GDB:125421 | DIASTROPHIC DYSPLASIA; DTD EPIPHYSEAL DYSPLASIA, MULTIPLE; MED NEONATAL OSSEOUS DYSPLASIA I ACHONDROGENESIS, TYPE IB; ACG1B |
| SLC6A3 | GDB:132445 | SOLUTE CARRIER FAMILY 6, MEMBER 3; SLC6A3 DEFICIT-HYPERACTIVITY DISORDER; ADHD |
| SM1 | GDB:9834488 | SCHISTOSOMA MANSONI SUSCEPTIBILITY/RESISTANCE |
| SMA@ | GDB:120378 | SPINAL MUSCULAR ATROPHY I; SMA I SURVIVAL OF MOTOR NEURON 1, TELOMERIC; SMN1 |
| SMN1 | GDB:5215173 | SPINAL MUSCULAR ATROPHY I; SMA I SURVIVAL OF MOTOR NEURON 1, TELOMERIC; SMN1 |
| SMN2 | GDB:5215175 | SPINAL MUSCULAR ATROPHY I; SMA I SURVIVAL OF MOTOR NEURON 2, CENTROMERIC; SMN2 |
| SPINK5 | GDB:9956114 | NETHERTON DISEASE |
| TCOF1 | GDB:127390 | TREACHER COLLINS-FRANCESCHETTI SYNDROME 1; TCOF1 |
| TGFBI | GDB:597601 | CORNEAL DYSTROPHY, GRANULAR TYPE CORNEAL DYSTROPHY, LATTICE TYPE I; CDL1 |

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| | | TRANSFORMING GROWTH FACTOR, BETA-INDUCED, 68 KD; TGFBI |
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Table 8: Genes, Locations and Genetic Disorders on Chromosome 6

| Gene | GDB Accession ID | OMIM Link |
|---------|------------------|--|
| ALDH5A1 | GDB:454767 | SUCCINIC SEMIALDEHYDE DEHYDROGENASE, NAD(+)-DEPENDENT; SSADH |
| ARG1 | GDB:119006 | ARGININEMIA |
| AS | GDB:135697 | ANKYLOSING SPONDYLITIS; AS |
| ASSP2 | GDB:119017 | CITRULLINEMIA |
| BCKDHB | GDB:118759 | MAPLE SYRUP URINE DISEASE, TYPE IB |
| BF | GDB:119726 | GLYCINE-RICH BETA-GLYCOPROTEIN; GBG |
| C2 | GDB:119731 | COMPLEMENT COMPONENT-2, DEFICIENCY OF |
| C4A | GDB:119732 | COMPLEMENT COMPONENT 4A; C4A |
| CDKN1A | GDB:266550 | CYCLIN-DEPENDENT KINASE INHIBITOR 1A; CDKN1A |
| COL10A1 | GDB:128635 | COLLAGEN, TYPE X, ALPHA 1; COL10A1 |
| COL11A2 | GDB:119788 | COLLAGEN, TYPE XI, ALPHA-2; COL11A2 STICKLER SYNDROME, TYPE |

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| | | II; STL2 DEAFNESS, AUTOSOMAL DOMINANT NONSYNDROMIC SENSORINEURAL, 13; DFNA13 |
| CYP21A2 | GDB:120605 | ADRENAL HYPERPLASIA, CONGENITAL, DUE TO 21-HYDROXYLASE DEFICIENCY |
| DYX2 | GDB:437584 | DYSLEXIA, SPECIFIC, 2; DYX2 |
| EJM1 | GDB:119864 | MYOCLONIC EPILEPSY, JUVENILE; EJM1 |
| ELOVL4 | GDB:11499609 | STARGARDT DISEASE 3; STGD3 |
| EPM2A | GDB:3763331 | EPILEPSY, PROGRESSIVE MYOCLONIC 2; EPM2 |
| ESR1 | GDB:119120 | ESTROGEN RECEPTOR; ESR |
| EYA4 | GDB:700062 | DEAFNESS, AUTOSOMAL DOMINANT NONSYNDROMIC SENSORINEURAL, 10; DFNA10 |
| F13A1 | GDB:120614 | FACTOR XIII, A1 SUBUNIT; F13A1 |
| FANCE | GDB:1220236 | FANCONI ANEMIA, COMPLEMENTATION GROUP E; FACE |
| GCLC | GDB:132915 | GAMMA-GLUTAMYL CYSTEINE SYNTHETASE DEFICIENCY, HEMOLYTIC ANEMIA DUE |
| GJA1 | GDB:125196 | GAP JUNCTION PROTEIN, ALPHA-1, 43 KD; GJA1 |
| GLYS1 | GDB:136421 | GLYCOSURIA, RENAL |

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| GMPR | GDB:127058 | GUANINE MONOPHOSPHATE REDUCTASE |
| GSE | GDB:9956235 | DISEASE; CD |
| HCR | GDB:9993306 | PSORIASIS, SUSCEPTIBILITY TO HFE GDB:119309 HEMOCHROMATOSIS; HFE |
| HLA-A | GDB:119310 | HLA-A HISTOCOMPATIBILITY TYPE; HLAA HLA-DPB1 GDB:120636 LA-DP HISTOCOMPATIBILITY TYPE, BETA-1 SUBUNIT |
| HLA-DRA | GDB:120641 | HLA-DR HISTOCOMPATIBILITY TYPE; HLA-DRA |
| HPFH | GDB:9849006 | HETEROCELLULAR HEREDITARY PERSISTENCE OF FETAL HEMOGLOBIN |
| ICS1 | GDB:136433 | IMMOTILE CILIA SYNDROME-1; ICS1 |
| IDDM1 | GDB:9953173 | DIABETES MELLITUS, JUVENILE-ONSET INSULIN-DEPENDENT; IDDM |
| IFNGR1 | GDB:120688 | INTERFERON, GAMMA, RECEPTOR-1; IFNGR1 |
| IGAD1 | GDB:6929077 | SELECTIVE DEFICIENCY OF |
| IGF2R | GDB:120083 | INSULIN-LIKE GROWTH FACTOR 2 RECEPTOR; IGF2R |
| ISCW | GDB:9956158 | SUPPRESSION; IS |

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| LAMA2 | GDB:132362 | LAMININ, ALPHA 2; LAMA2 |
| LAP | GDB:9958992 | LARYNGEAL ADDUCTOR PARALYSIS; LAP |
| LCA5 | GDB:11498764 | AMAUROSIS CONGENITA OF LEBER I |
| LPA | GDB:120699 | APOLIPOPROTEIN(a); LPA |
| MCDR1 | GDB:131406 | MACULAR DYSTROPHY, RETINAL, 1, NORTH CAROLINA TYPE; MCDR1 |
| MOCS1 | GDB:9862235 | MOLYBDENUM COFACTOR DEFICIENCY |
| MUT | GDB:120204 | METHYLMALONIC ACIDURIA DUE TO METHYLMALONIC CoA MUTASE DEFICIENCY |
| MYB | GDB:119441 | V-MYB AVIAN MYELOBLASTOSIS VIRAL ONCOGENE HOMOLOG; MYB |
| NEU1 | GDB:120230 | NEURAMINIDASE DEFICIENCY |
| NKS1 | GDB:128100 | SUSCEPTIBILITY TO LYSIS BY ALLOREACTIVE NATURAL KILLER CELLS; EC1 |
| NYS2 | GDB:9848763 | NYSTAGMUS, CONGENITAL |
| OA3 | GDB:136429 | ALBINISM, OCULAR, AUTOSOMAL RECESSIVE; OAR |
| ODDD | GDB:6392584 | OCULODENTODIGITAL DYSPLASIA; ODDD |

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| OFC1 | GDB:120247 | OROFACIAL CLEFT 1; OFC1 |
| PARK2 | GDB:6802742 | PARKINSONISM, JUVENILE |
| PBCA | GDB:9956321 | BETA CELL AGENESIS WITH NEONATAL DIABETES MELLITUS |
| PBCRA1 | GDB:3763333 | CHORIORETINAL ATROPHY, PROGRESSIVE BIFOCAL; CRAPB |
| PDB1 | GDB:136349 | DISEASE OF BONE; PDB |
| PEX3 | GDB:9955507 | ZELLWEGER SYNDROME; ZS |
| PEX6 | GDB:5592414 | ZELLWEGER SYNDROME; ZS PEROXIN-6; PEX6 |
| PEX7 | GDB:6155803 | RHIZOMELIC CHONDRODYSPLASIA PUNCTATA; RCDP PEROXIN-7; PEX7 |
| PKHD1 | GDB:433910 | POLYCYSTIC KIDNEY AND HEPATIC DISEASE-1; PKHD1 |
| PLA2G7 | GDB:9958829 | PLATELET-ACTIVATING FACTOR ACETYLHYDROLASE, SUBUNIT |
| PLG | GDB:119498 | PLASMINOGEN; PLG |
| POLH | GDB:6963323 | PIGMENTOSUM WITH NORMAL DNA REPAIR RATES |
| PPAC | GDB:9956248 | ARTHROPATHY, PROGRESSIVE PSEUDORHEUMATOID, OF CHILDHOOD |

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| PSORS1 | GDB:6381310 | PSORIASIS, SUSCEPTIBILITY TO |
| PUJO | GDB:9956231 | MULTICYSTIC RENAL DYSPLASIA, BILATERAL; MRD |
| RCD1 | GDB:333929 | RETINAL CONE DEGENERATION |
| RDS | GDB:118863 | RETINAL DEGENERATION, SLOW; RDS |
| RHAG | GDB:136011 | RHESUS BLOOD GROUP-ASSOCIATED GLYCOPROTEIN; RHAG RH-NUL, REGULATOR TYPE; RHN |
| RP14 | GDB:433713 | RETINITIS PIGMENTOSA-14; RP14 TUBBY-LIKE PROTEIN 1; TULP1 |
| RUNX2 | GDB:392082 | CLEIDOCRANIAL DYSPLASIA; CCD CORE-BINDING FACTOR, RUNT DOMAIN, ALPHA SUBUNIT 1; CBFA1 |
| RWS | GDB:9956195 | SENSITIVITY |
| SCA1 | GDB:119588 | SPINOCEBELLAR ATAXIA 1; SCA1 |
| SCZD3 | GDB:635974 | DISORDER-3; SCZD3 |
| SIASD | GDB:433552 | SIALIC ACID STORAGE DISEASE; SIASD |
| SOD2 | GDB:119597 | SUPEROXIDE DISMUTASE 2, MITOCHONDRIAL; SOD2 |
| ST8 | GDB:6118456 | OVARIAN TUMOR |
| TAP1 | GDB:132668 | TRANSPORTER 1, ABC; TAP1 |

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| TAP2 | GDB:132669 | TRANSPORTER 2, ABC; TAP2 |
| TFAP2B | GDB:681506 | DUCTUS ARTERIOSUS; PDA TRANSCRIPTION FACTOR AP-2 BETA; TFAP2B |
| TNDM | GDB:9956265 | DIABETES MELLITUS, TRANSIENT NEONATAL |
| TNF | GDB:120441 | TUMOR NECROSIS FACTOR; TNF |
| TPBG | GDB:125568 | TROPHOBlast GLYCOPROTEIN; TPBG; M6P1 |
| TPMT | GDB:209025 | THIOPURINE S-METHYLTRANSFERASE; TPMT |
| TULP1 | GDB:6199353 | TUBBY-LIKE PROTEIN 1; TULP1 |
| WISP3 | GDB:9957361 | ARTHROPATHY, PROGRESSIVE PSEUDORHEUMATOID, OF CHILDHOOD |

Table 9: Genes, Locations and Genetic Disorders on Chromosome 7

| Gene | GDB Accession ID | OMIM Link |
|-------|------------------|--|
| AASS | GDB:11502144 | HYPERLYSINEMIA |
| ABCB1 | GDB:120712 | P-GLYCOPROTEIN-1; PGY1 |
| ABCB4 | GDB:120713 | P-GLYCOPROTEIN-3; PGY3 |
| ACHE | GDB:118746 | ACETYLCHOLINESTERASE BLOOD GROUP--Yt SYSTEM; YT |

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| AQP1 | GDB:129082 | AQUAPORIN-1; AQP1 BLOOD GROUP--COLTON; CO |
| ASL | GDB:119703 | ARGININOSUCCINICACIDURIA |
| ASNS | GDB:119706 | ASPARAGINE SYNTHETASE; ASNS; AS |
| AUTS1 | GDB:9864226 | DISORDER |
| BPGM | GDB:119039 | DIPHOSPHOGLYCERATE MUTASE DEFICIENCY OF ERYTHROCYTE |
| C7orf2 | GDB:10794644 | ACHEIROPODY |
| CACNA2D1 | GDB:132010 | CALCIUM CHANNEL, VOLTAGE-DEPENDENT, L TYPE; ALPHA-2/DELTA SUBUNIT; MALIGNANT HYPERTHERMIA SUSCEPTIBILITY-3 |
| CCM1 | GDB:580824 | CEREBRAL CAVERNOUS MALFORMATIONS 1; CCM1 |
| CD36 | GDB:138800 | CD36 ANTIGEN; CD36 |
| CFTR | GDB:120584 | CYSTIC FIBROSIS; CF DEFERENS, CONGENITAL BILATERAL APLASIA OF; CBAVD; CAVD |
| CHORDOMA | GDB:11498328 | |
| CLCN1 | GDB:134688 | CHLORIDE CHANNEL 1, SKELETAL MUSCLE; CLCN1 |
| CMH6 | GDB:9956392 | CARDIOMYOPATHY, FAMILIAL HYPERTROPHIC, WITH WOLFF-PARKINSON-WHITE |

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| CMT2D | GDB:9953232 | CHARCOT-MARIE-TOOTH DISEASE, NEURONAL TYPE, D |
| COL1A2 | GDB:119062 | COLLAGEN, TYPE I, ALPHA-2 POLYPEPTIDE; COL1A2 OSTEOGENESIS IMPERFECTA TYPE I OSTEOGENESIS IMPERFECTA TYPE IV; OI4 |
| CRS | GDB:119073 | CRANIOSYNOSTOSIS, TYPE 1; CRS1 |
| CYMD | GDB:366594 | MACULAR EDEMA, CYSTOID |
| DFNA5 | GDB:636174 | DEAFNESS, AUTOSOMAL DOMINANT NONSYNDROMIC SENSORINEURAL, 5; DFNA5 |
| DLD | GDB:120608 | LIPOAMIDE DEHYDROGENASE DEFICIENCY, LACTIC ACIDOSIS DUE TO |
| DYT11 | GDB:10013754 | MYOCLONUS, HEREDITARY ESSENTIAL |
| EEC1 | GDB:136338 | ECTRODACTYLY, ECTODERMAL DYSPLASIA, AND CLEFT LIP/PALATE; EEC |
| ELN | GDB:119107 | ELASTIN; ELN WILLIAMS-BEUREN SYNDROME; WBS |
| ETV1 | GDB:335229 | ETS VARIANT GENE 1; ETV1 |
| FKBP6 | GDB:9955215 | WILLIAMS-BEUREN SYNDROME; WBS |
| GCK | GDB:127550 | DIABETES MELLITUS, AUTOSOMAL DOMINANT, TYPE II GLUCOKINASE; GCK |

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| GHRHR | GDB:138465 | GROWTH HORMONE-RELEASING HORMONE RECEPTOR; GHRHR |
| GHS | GDB:9956363 | MICROSOMIA WITH RADIAL DEFECTS |
| GLI3 | GDB:119990 | PALLISTER-HALL SYNDROME; PHS GLI-KRUPPEL FAMILY MEMBER 3; GLI3 POSTAXIAL POLYDACTYLY, TYPE A1 GREIG CEPHALOPOLYSYNDACTYLY SYNDROME; GCPS |
| GPDS1 | GDB:9956410 | GLAUCOMA, PIGMENT-DISPERSION TYPE |
| GUSB | GDB:120025 | MUCOPOLYSACCHARIDOSIS TYPE VII |
| HADH | GDB:120033 | HYDROXYACYL-CoA DEHYDROGENASE/3-KETOACYL-CoA THIOLASE/ENOYL-CoA HYDRATASE, |
| HLXB9 | GDB:136411 | HOME BOX GENE HB9; HLXB9 SACRAL AGENESIS, HEREDITARY, WITH PRESACRAL MASS, ANTERIOR MENINGOCELE, |
| HOXA13 | GDB:120656 | HOME BOX A13; HOXA13 |
| HPFH2 | GDB:128071 | HEREDITARY PERSISTENCE OF FETAL HEMOGLOBIN, HETEROCELLULAR, INDIAN |
| HRX | GDB:9958999 | HRX |
| IAB | GDB:11498909 | ANEURYSM, INTRACRANIAL BERRY |
| IMMP2L | GDB:11499195 | GILLES DE LA TOURETTE SYNDROME; GTS |

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| KCNH2 | GDB:138126 | LONG QT SYNDROME, TYPE 2; LQT2 |
| LAMB1 | GDB:119357 | LAMININ BETA 1; LAMB1 |
| LEP | GDB:136420 | LEPTIN; LEP |
| MET | GDB:120178 | MET PROTO-ONCOGENE; MET |
| NCF1 | GDB:120222 | GRANULOMATOUS DISEASE, CHRONIC, AUTOSOMAL CYTOCHROME-b-POSITIVE FORM |
| NM | GDB:119454 | NEUTROPHIL CHEMOTACTIC RESPONSE; NCR |
| OGDH | GDB:118847 | ALPHA-KETOGLUTARATE DEHYDROGENASE DEFICIENCY |
| OPN1SW | GDB:119032 | TRITANOPIA |
| PEX1 | GDB:9787110 | ZELLWEGER SYNDROME; ZS PEROXIN-1; PEX1 |
| PGAM2 | GDB:120280 | PHOSPHOGLYCERATE MUTASE, DEFICIENCY OF M SUBUNIT OF |
| PMS2 | GDB:386406 | POSTMEIOTIC SEGREGATION INCREASED (S. CEREVISIAE)-2; PMS2 |
| PON1 | GDB:120308 | PARAOXONASE 1; PON1 |
| PPP1R3A | GDB:136797 | PROTEIN PHOSPHATASE 1, REGULATORY (INHIBITOR) SUBUNIT 3; PPP1R3 |

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| PRSS1 | GDB:119620 | PANCREATITIS, HEREDITARY; PCTT PROTEASE, SERINE, 1; PRSS1 |
| PTC | GDB:118744 | PHENYLTHIOCARBAMIDE TASTING |
| PTPN12 | GDB:136846 | PROTEIN-TYROSINE PHOSPHATASE, NONRECEPTOR TYPE, 12; PTPN12 |
| RP10 | GDB:138786 | RETINITIS PIGMENTOSA-10; RP10 |
| RP9 | GDB:333931 | RETINITIS PIGMENTOSA-9; RP9 |
| SERPINE1 | GDB:120297 | PLASMINOGEN ACTIVATOR INHIBITOR, TYPE I; PAI1 |
| SGCE | GDB:9958714 | MYOCLONUS, HEREDITARY ESSENTIAL |
| SHFM1 | GDB:128195 | SPLIT-HAND/FOOT DEFORMITY, TYPE I; SHFD1 |
| SHH | GDB:456309 | HOLOPROSENCEPHALY, TYPE 3; HPE3 SONIC HEDGEHOG, DROSOPHILA, HOMOLOG OF; SHH |
| SLC26A3 | GDB:138165 | DOWN-REGULATED IN ADENOMA; DRA CHLORIDE DIARRHEA, FAMILIAL; CLD |
| SLC26A4 | GDB:5584511 | PENDRED SYNDROME; PDS DEAFNESS, NEUROSENSORY, AUTOSOMAL RECESSIVE, 4; DFNB4 |
| SLOS | GDB:385950 | SMITH-LEMLI-OPITZ SYNDROME |
| SMAD1 | GDB:3763345 | SPINAL MUSCULAR ATROPHY, DISTAL, WITH UPPER LIMB |

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| | | PREDOMINANCE; SMAD1 |
| TBXAS1 | GDB:128744 | THROMBOXANE A SYNTHASE 1; TBXAS1 |
| TWIST | GDB:135694 | ACROCEPHALOSYNDACTYLY TYPE III TWIST, DROSOPHILA, HOMOLOG OF; TWIST |
| ZWS1 | GDB:120511 | ZELLWEGER SYNDROME; ZS |

Table 10: Genes, Locations and Genetic Disorders on Chromosome 8

| Gene | GDB AccessionID | OMIM Link |
|-------|-----------------|--|
| ACHM3 | GDB:9120558 | PINGELAPESE BLINDNESS |
| ADRB3 | GDB:203869 | BETA-3-ADRENERGIC RECEPTOR; ADRB3 |
| ANK1 | GDB:118737 | SPHEROCYTOSIS, HEREDITARY; HS |
| CA1 | GDB:119047 | CARBONIC ANHYDRASE I, ERYTHROCYTE, ELECTROPHORETIC VARIANTS OF; CA1 |
| CA2 | GDB:119739 | OSTEOPETROSIS WITH RENAL TUBULAR ACIDOSIS |
| CCAL1 | GDB:512892 | CHONDROCALCINOSIS WITH EARLY-ONSET OSTEOARTHRITIS; CCAL2 |
| CLN8 | GDB:252118 | EPILEPSY, PROGRESSIVE, WITH MENTAL RETARDATION; EPMR |
| CMT4A | GDB:138755 | CHARCOT-MARIE-TOOTH NEUROPATHY 4A; CMT4A |

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| CNGB3 | GDB:9993286 | PINGELAPESE BLINDNESS |
| COH1 | GDB:252122 | COHEN SYNDROME; COH1 |
| CPP | GDB:119798 | CERULOPLASMIN; CP |
| CRH | GDB:119804 | CORTICOTROPIN-RELEASING HORMONE; CRH |
| CYP11B1 | GDB:120603 | ADRENAL HYPERPLASIA, CONGENITAL, DUE TO 11-@BETA-HYDROXYLASE DEFICIENCY |
| CYP11B2 | GDB:120514 | CYTOCHROME P450, SUBFAMILY XIB, POLYPEPTIDE 2; CYP11B2 |
| DECR1 | GDB:453934 | 2,4-@DIENOYL-CoA REDUCTASE; DECR |
| DPYS | GDB:5885803 | DIHYDROPYRIMIDINASE; DPYS |
| DURS1 | GDB:9958126 | DUANE SYNDROME |
| EBS1 | GDB:119856 | EPIDERMOLYSIS BULLOSA SIMPLEX, OGNA TYPE |
| ECA1 | GDB:10796318 | JUVENILE ABSENCE |
| EGI | GDB:128830 | EPILEPSY, GENERALIZED, IDIOPATHIC; EGI |
| EXT1 | GDB:135994 | EXOSTOSES, MULTIPLE, TYPE I; EXT1 CHONDROSARCOMA |
| EYA1 | GDB:5215167 | BRANCHIOOTORENAL DYSPLASIA EYES ABSENT 1; EYA1 |

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| FGFR1 | GDB:119913 | ACROCEPHALOSYNDACTYLY TYPE V FIBROBLAST GROWTH FACTOR RECEPTOR-1; FGFR1 |
| GNRH1 | GDB:133746 | GONADOTROPIN-RELEASING HORMONE 1; GNRH1 FAMILIAL HYPOGONADOTROPHIC |
| GSR | GDB:119288 | GLUTATHIONE REDUCTASE; GSR |
| GULOP | GDB:128078 | SCURVY |
| HR | GDB:595499 | ALOPECIA UNIVERSALIS ATRICHLA WITH PAPULAR LESIONS HAIRLESS, MOUSE, HOMOLOG OF |
| KCNQ3 | GDB:9787230 | CONVULSIONS, BENIGN FAMILIAL NEONATAL, TYPE 2; BFNC2 POTASSIUM CHANNEL, VOLTAGE-GATED, SUBFAMILY Q, MEMBER 3 |
| KFM | GDB:265291 | KLIPPEL-FEIL SYNDROME; KFS; KFM |
| KWE | GDB:9315120 | KERATOLYTIC WINTER ERYTHEMA |
| LGCR | GDB:120698 | LANGER-GIEDION SYNDROME; LGS |
| LPL | GDB:120700 | HYPERLIPOPROTEINEMIA, TYPE I |
| MCPH1 | GDB:9834525 | MICROCEPHALY; MCT |
| MOS | GDB:119396 | TRANSFORMATION GENE: ONCOGENE MOS |
| MYC | GDB:120208 | TRANSFORMATION GENE: ONCOGENE MYC; MYC |

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| NAT1 | GDB:125364 | ARYLAMIDE ACETYLASE 1; AAC1 |
| NAT2 | GDB:125365 | ISONIAZID INACTIVATION |
| NBS1 | GDB:9598211 | NIJMEGEN BREAKAGE SYNDROME |
| PLAT | GDB:119496 | PLASMINOGEN ACTIVATOR, TISSUE; PLAT |
| PLEC1 | GDB:4119073 | EPIDERMOLYSIS BULLOSA SIMPLEX AND LIMB-GIRDLE MUSCULAR DYSTROPHY PLECTIN 1; PLEC1 |
| PRKDC | GDB:234702 | SEVERE COMBINED IMMUNODEFICIENCY DISEASE-1; SCID1 PROTEIN KINASE, DNA-ACTIVATED, CATALYTIC SUBUNIT; PRKDC |
| PXMP3 | GDB:131487 | PEROXIN-2; PEX2 ZELLWEGER SYNDROME; ZS |
| RP1 | GDB:120352 | RETINITIS PIGMENTOSA-1; RP1 |
| SCZD6 | GDB:9864736 | DISORDER-2; SCZD2 |
| SFTPC | GDB:120373 | PULMONARY SURFACTANT APOPROTEIN PSP-C |
| SGM1 | GDB:135350 | KLIPPEL-FEIL SYNDROME; KFS; KFM |
| SPG5A | GDB:250332 | SPASTIC PARAPLEGIA-5A, AUTOSOMAL RECESSIVE; SPG5A |
| STAR | GDB:635457 | STEROIDOGENIC ACUTE REGULATORY PROTEIN; STAR |

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| TG | GDB:120434 | THYROGLOBULIN; TG |
| TRPS1 | GDB:594960 | TRICORRHINOPHALANGEAL SYNDROME, TYPE I; TRPS1 |
| TTPA | GDB:512364 | VITAMIN E, FAMILIAL ISOLATED DEFICIENCY OF; VED TOCOPHEROL (ALPHA) TRANSFER PROTEIN; TTPA |
| VMD1 | GDB:119631 | MACULAR DYSTROPHY, ATYPICAL VITELLIFORM; VMD1 |
| WRN | GDB:128446 | WERNER SYNDROME; WRN |

Table 11: Genes, Locations and Genetic Disorders on Chromosome 9

| Gene | GDB AccessionID | OMIM Link |
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| ABCA1 | GDB:305294 | ANALPHALIPOPROTEINEMIA ATP-BINDING CASSETTE 1; ABC1 |
| ABL1 | GDB:119640 | ABELSON MURINE LEUKEMIA VIRAL ONCOGENE HOMOLOG 1; ABL1 |
| ABO | GDB:118956 | ABO BLOOD GROUP; ABO |
| ADAMTS13 | GDB:9956467 | THROMBOCYTOPENIC PURPURA |
| AK1 | GDB:119664 | ADENYLATE KINASE-1; AK1 |
| ALAD | GDB:119665 | DELTA-AMINOLEVULINATE DEHYDRATASE; ALAD |
| ALDH1A1 | GDB:119667 | ALDEHYDE DEHYDROGENASE-1; ALDH1 |

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| ALDOB | GDB:119669 | FRUCTOSE INTOLERANCE, HEREDITARY |
| AMBP | GDB:120696 | PROTEIN HC; HCP |
| AMCD1 | GDB:437519 | ARTHROGRYPOSIS MULTIPLEX CONGENITA, DISTAL, TYPE 1; AMCD1 |
| ASS | GDB:119010 | CITRULLINEMIA |
| BDMF | GDB:9954424 | BONE DYSPLASIA WITH MEDULLARY FIBROSARCOMA |
| BSCL | GDB:9957720 | SEIP SYNDROME |
| C5 | GDB:119734 | COMPLEMENT COMPONENT-5, DEFICIENCY OF |
| CDKN2A | GDB:335362 | MELANOMA, CUTANEOUS MALIGNANT, 2; CMM2 CYCLIN-DEPENDENT KINASE INHIBITOR 2A; CDKN2A |
| CHAC | GDB:6268491 | CHOREOACANTHOCYTOSIS; CHAC |
| CHH | GDB:138268 | CARTILAGE-HAIR HYPOPLASIA; CHH |
| CMD1B | GDB:677147 | CARDIOMYOPATHY, DILATED 1B; CMD1B |
| COL5A1 | GDB:131457 | COLLAGEN, TYPE V, ALPHA-1 POLYPEPTIDE; COL5A1 |
| CRAT | GDB:359759 | CARNITINE ACETYLTRANSFERASE; CRAT |
| DBH | GDB:119836 | DOPAMINE BETA-HYDROXYLASE, PLASMA; DBH |

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| DFNB11 | GDB:1220180 | DEAFNESS, NEUROSENSORY, AUTOSOMAL RECESSIVE, 7; DFNB7 |
| DFNB7 | GDB:636178 | DEAFNESS, NEUROSENSORY, AUTOSOMAL RECESSIVE, 7; DFNB7 |
| DNAI1 | GDB:11500297 | IMMOTILE CILIA SYNDROME-1; ICS1 |
| DYS | GDB:137085 | DYSAUTONOMIA, FAMILIAL; DYS |
| DYT1 | GDB:119854 | DYSTONIA 1, TORSION; DYT1 |
| ENG | GDB:137193 | ENDOGLIN; ENG |
| EPB72 | GDB:128993 | ERYTHROCYTE SURFACE PROTEIN BAND 7.2; EPB72 STOMATOCYTOSIS I |
| FANCC | GDB:132672 | FANCONI ANEMIA, COMPLEMENTATION GROUP C; FACC |
| FBP1 | GDB:141539 | FRUCTOSE-1,6-BISPHOPHATASE 1; FBP1 |
| FCMD | GDB:250412 | FUKUYAMA-TYPE CONGENITAL MUSCULAR DYSTROPHY; FCMD |
| FRDA | GDB:119951 | FRIEDREICH ATAXIA 1; FRDA1 |
| GALT | GDB:119971 | GALACTOSEMIA |
| GLDC | GDB:128611 | HYPERGLYCINEMIA, ISOLATED NONKETOTIC, TYPE I; NKH1 |
| GNE | GDB:9954891 | INCLUSION BODY MYOPATHY; IBM2 |

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| GSM1 | GDB:9784210 | GENIOSPASM 1; GSM1 |
| GSN | GDB:120019 | AMYLOIDOSIS V GELSOLIN; GSN |
| HSD17B3 | GDB:347487 | PSEUDOHERMAPHRODITISM, MALE, WITH GYNECOMASTIA |
| HSN1 | GDB:3853677 | NEUROPATHY, HEREDITARY SENSORY, TYPE 1 |
| IBM2 | GDB:3801447 | INCLUSION BODY MYOPATHY; IBM2 |
| LALL | GDB:9954426 | LEUKEMIA, ACUTE, WITH LYMPHOMATOUS FEATURES; LALL |
| LCCS | GDB:386141 | LETHAL CONGENITAL CONTRACTURE SYNDROME; LCCS |
| LGMD2H | GDB:9862233 | DYSTROPHY, HUTTERITE TYPE |
| LMX1B | GDB:9834526 | NAIL-PATELLA SYNDROME; NPS1 |
| MLLT3 | GDB:138172 | MYELOID/LYMPHOID OR MIXED LINEAGE LEUKEMIA, TRANSLOCATED TO, 3; MLLT3 |
| MROS | GDB:9954430 | MELKERSSON SYNDROME |
| MSSE | GDB:128019 | EPITHELIOMA, SELF-HEALING SQUAMOUS |
| NOTCH1 | GDB:131400 | NOTCH, DROSOPHILA, HOMOLOG OF, 1; NOTCH1 |
| ORM1 | GDB:120250 | OROSOMUCOID 1; ORM1 |

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| PAPPA | GDB:134729 | PREGNANCY-ASSOCIATED PLASMA PROTEIN A; PAPPA |
| PIP5K1B | GDB:686238 | FRIEDREICH ATAXIA 1; FRDA1 |
| PTCH | GDB:119447 | BASAL CELL NEVUS SYNDROME; BCNS PATCHED, DROSOPHILA, HOMOLOG OF; PTCH |
| PTGS1 | GDB:128070 | PROSTAGLANDIN-ENDOPEROXIDASE SYNTHASE 1; PTGS1 |
| RLN1 | GDB:119552 | RELAXIN; RLN1 |
| RLN2 | GDB:119553 | RELAXIN, OVARIAN, OF PREGNANCY |
| RMRP | GDB:120348 | MITOCHONDRIAL RNA-PROCESSING ENDORIBONUCLEASE, RNA COMPONENT OF; RMRP; CARTILAGE-HAIR HYPOPLASIA; CHH |
| ROR2 | GDB:136454 | BRACHYDACTYLY, TYPE B; BDB ROBINOW SYNDROME, RECESSIVE FORM NEUROTROPHIC TYROSINE KINASE, RECEPTOR-RELATED 2; NTRKR2 |
| RPD1 | GDB:9954440 | RETINITIS PIGMENTOSA-DEAFNESS SYNDROME 1, AUTOSOMAL DOMINANT |
| SARDH | GDB:9835149 | SARCOSINEMIA |
| TDFA | GDB:9954420 | FACTOR, AUTOSOMAL |
| TEK | GDB:344185 | VENOUS MALFORMATIONS, MULTIPLE CUTANEOUS AND MUCOSAL; VMCM TEK TYROSINE KINASE, ENDOTHELIAL; TEK |

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| TSC1 | GDB:120735 | TUBEROUS SCLEROSIS-1; TSC1 |
| TYRP1 | GDB:126337 | TYROSINASE-RELATED PROTEIN 1; TYRP1 ALBINISM III XANTHISM |
| XPA | GDB:125363 | XERODERMA PIGMENTOSUM I |

Table 12: Genes, Locations and Genetic Disorders on Chromosomes 10

| Gene | GDB Accession ID | OMIM Link |
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| CACNB2 | GDB:132014 | CALCIUM CHANNEL, VOLTAGE-DEPENDENT, BETA-2 SUBUNIT; CACNB2 |
| COL17A1 | GDB:131396 | COLLAGEN, TYPE XVII, ALPHA-1 POLYPEPTIDE; COL17A1 |
| CUBN | GDB:636049 | MEGALOBLASTIC ANEMIA 1; MGA1 |
| CYP17 | GDB:119829 | ADRENAL HYPERPLASIA, CONGENITAL, DUE TO 17-ALPHA-HYDROXYLASE DEFICIENCY |
| CYP2C19 | GDB:119831 | CYTOCHROME P450, SUBFAMILY IIC, POLYPEPTIDE 19; CYP2C19 |
| CYP2C9 | GDB:131455 | CYTOCHROME P450, SUBFAMILY IIC, POLYPEPTIDE 9; CYP2C9 |
| EGR2 | GDB:120611 | EARLY GROWTH RESPONSE-2; EGR2 |
| EMX2 | GDB:277886 | EMPTY SPIRACLES, DROSOPHILA, 2, HOMOLOG OF; EMX2 |

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| EPT | GDB:9786112 | EPILEPSY, PARTIAL; EPT |
| ERCC6 | GDB:119882 | EXCISION-REPAIR CROSS-COMPLEMENTING RODENT REPAIR DEFICIENCY, COMPLEMENTATION |
| FGFR2 | GDB:127273 | ACROCEPHALOSYNDACTYLY TYPE V FIBROBLAST GROWTH FACTOR RECEPTOR-2; FGFR2 |
| HK1 | GDB:120044 | HEXOKINASE-1; HK1 |
| HOX11 | GDB:119607 | HOME BOX-11; HOX11 |
| HPS | GDB:127359 | HERMANSKY-PUDLAK SYNDROME; HPS |
| IL2RA | GDB:119345 | INTERLEUKIN-2 RECEPTOR, ALPHA; IL2RA |
| LGI1 | GDB:9864936 | EPILEPSY, PARTIAL; EPT |
| LIPA | GDB:120153 | WOLMAN DISEASE |
| MAT1A | GDB:129077 | METHIONINE ADENOSYLTRANSFERASE DEFICIENCY |
| MBL2 | GDB:120167 | MANNOSE-BINDING PROTEIN, SERUM; MBP1 |
| MKI67 | GDB:120185 | PROLIFERATION-RELATED Ki-67 ANTIGEN; MKI67 |
| MXI1 | GDB:137182 | MAX INTERACTING PROTEIN 1; MXII |

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| OAT | GDB:120246 | ORNITHINE AMINOTRANSFERASE DEFICIENCY |
| OATL3 | GDB:215803 | ORNITHINE AMINOTRANSFERASE DEFICIENCY |
| PAX2 | GDB:138771 | PAIRED BOX HOMEOTIC GENE 2; PAX2 |
| PCBD | GDB:138478 | PTERIN-4-ALPHA-CARBINOLAMINE DEHYDRATASE; PCBD PRIMAPTERINURIA |
| PEO1 | GDB:632784 | PROGRESSIVE EXTERNAL OPHTHALMOPLEGIA; PEO |
| PHYH | GDB:9263423 | REFSUM DISEASE PHYTANOYL-CoA HYDROXYLASE; PHYH |
| PNLIP | GDB:127916 | LIPASE, CONGENITAL ABSENCE OF PANCREATIC |
| PSAP | GDB:120366 | PROSAPOSIN; PSAP |
| PTEN | GDB:6022948 | MACROCEPHALY, MULTIPLE LIPOSAS AND HEMANGIOMATA MULTIPLE HAMARTOMA SYNDROME; MHAM POLYPOSIS, JUVENILE INTESTINAL PHOSPHATASE AND TENSIN HOMOLOG; PTEN |
| RBP4 | GDB:120342 | RETINOL-BINDING PROTEIN, PLASMA; RBP4 |
| RDPA | GDB:9954445 | REFSUM DISEASE WITH INCREASED PIPECOLICACIDEMIA; RDPA |

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| RET | GDB:120346 | RET PROTO-ONCOGENE; RET |
| SDF1 | GDB:433267 | STROMAL CELL-DERIVED FACTOR 1; SDF1 |
| SFTPA1 | GDB:119593 | PULMONARY SURFACTANT APOPROTEIN PSP-A; PSAP |
| SFTPД | GDB:132674 | PULMONARY SURFACTANT APOPROTEIN PSP-D; PSP-D |
| SHFM3 | GDB:386030 | SPLIT-HAND/FOOT MALFORMATION, TYPE 3; SHFM3 |
| SIAL | GDB:6549924 | NEURAMINIDASE DEFICIENCY |
| THC2 | GDB:10794765 | THROMBOCYTOPENIA |
| TNFRSF6 | GDB:132671 | APOPTOSIS ANTIGEN 1; APT1 |
| UFS | GDB:6380714 | UROFACIAL SYNDROME; UFS |
| UROS | GDB:128112 | PORPHYRIA, CONGENITAL ERYTHROPOIETIC; CEP |

Table 13: Genes, Locations and Genetic Disorders on Chromosome 11

| Gene | GDB Accession ID | OMIM Link |
|-------|------------------|--|
| AA | GDB:568984 | ATROPHIA AREATA; AA |
| ABCC8 | GDB:591370 | SULFONYLUREA RECEPTOR; SUR PERSISTENT HYPERINSULINEMIC HYPOGLYCEMIA OF INFANCY |

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| ACAT1 | GDB:126861 | ALPHA-METHYLACETOACETICACIDURIA |
| ALX4 | GDB:10450304 | PARIETAL FORAMINA, SYMMETRIC; PFM |
| AMPD3 | GDB:136013 | ADENOSINE MONOPHOSPHATE DEAMINASE-3; AMPD3 |
| ANC | GDB:9954484 | CANAL CARCINOMA |
| APOA1 | GDB:119684 | AMYLOIDOSIS, FAMILIAL VISCERAL APOLIPOPROTEIN A-I OF HIGH DENSITY LIPOPROTEIN; APOA1 |
| APOA4 | GDB:119000 | APOLIPOPROTEIN A-IV; APOA4 |
| APOC3 | GDB:119001 | APOLIPOPROTEIN C-III; APOC3 |
| ATM | GDB:593364 | ATAxia-TELANGIECTASIA; AT |
| BSCL2 | GDB:9963996 | SEIP SYNDROME |
| BWS | GDB:120567 | BECKWITH-WIEDEMANN SYNDROME; BWS |
| CALCA | GDB:120571 | CALCITONIN/CALCITONIN-RELATED POLYPEPTIDE, ALPHA; CALCA |
| CAT | GDB:119049 | CATALASE; CAT |
| CCND1 | GDB:128222 | LEUKEMIA, CHRONIC LYMPHATIC; CLL CYCLIN D1; CCND1 |
| CD3E | GDB:119764 | CD3E ANTIGEN, EPSILON POLYPEPTIDE; CD3E |

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| CD3G | GDB:119765 | T3 T-CELL ANTIGEN, GAMMA CHAIN; T3G; CD3G |
| CD59 | GDB:119769 | CD59 ANTIGEN P18-20; CD59 HUMAN LEUKOCYTE ANTIGEN MIC11; MIC11 |
| CDKN1C | GDB:593296 | CYCLIN-DEPENDENT KINASE INHIBITOR 1C; CDKN1C |
| CLN2 | GDB:125228 | CEROID-LIPOFUSCINOSIS, NEURONAL 2, LATE INFANTILE TYPE; CLN2 |
| CNTF | GDB:125919 | CILIARY NEUROTROPHIC FACTOR; CNTF |
| CPT1A | GDB:597642 | HYPOGLYCEMIA, HYPOKETOTIC, WITH DEFICIENCY OF CARNITINE PALMITOYLTRANSFERASE CARNITINE PALMITOYLTRANSFERASE I, LIVER; CPT1A |
| CTSC | GDB:642234 | KERATOSIS PALMOPLANTARIS WITH PERIODONTOPATHIA KERATOSIS PALMOPLANTARIS WITH PERIODONTOPATHIA ANDONYCHOGRYPOSIS CATHEPSIN C; CTSC |
| DDB1 | GDB:595014 | DNA DAMAGE-BINDING PROTEIN; DDB1 |
| DDB2 | GDB:595015 | DNA DAMAGE-BINDING PROTEIN-2; DDB2 |
| DHCR7 | GDB:9835302 | SMITH-LEMLI-OPITZ SYNDROME |
| DLAT | GDB:118785 | CIRRHOSIS, PRIMARY; PBC |
| DRD4 | GDB:127782 | DOPAMINE RECEPTOR D4; DRD4 |
| ECB2 | GDB:9958955 | POLYCYTHEMIA, BENIGN FAMILIAL |

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| ED4 | GDB:9837373 | DYSPLASIA, MARGARITA TYPE |
| EVR1 | GDB:134029 | EXUDATIVE VITREORETINOPATHY, FAMILIAL; EVR EXT2GDB:344921EXOSTOSES, MULTIPLE, TYPE II; EXT2 CHONDROSARCOMA |
| F2 | GDB:119894 | COAGULATION FACTOR II; F2 |
| FSHB | GDB:119955 | FOLLICLE-STIMULATING HORMONE, BETA POLYPEPTIDE; FSHB |
| FTH1 | GDB:120617 | FERRITIN HEAVY CHAIN 1; FTH1 |
| GIF | GDB:118800 | PERNICIOUS ANEMIA, CONGENITAL, DUE TO DEFECT OF INTRINSIC FACTOR |
| GSD1B | GDB:9837619 | GLYCOGEN STORAGE DISEASE Ib |
| GSD1C | GDB:9837637 | STORAGE DISEASE Ic |
| HBB | GDB:119297 | HEMOGLOBIN--BETA LOCUS; HBB |
| HBBP1 | GDB:120035 | HEMOGLOBIN--BETA LOCUS; HBB |
| HBD | GDB:119298 | HEMOGLOBIN--DELTA LOCUS; HBD |
| HBE1 | GDB:119299 | HEMOGLOBIN--EPSILON LOCUS; HBE1 |
| HBG1 | GDB:119300 | HEMOGLOBIN, GAMMA A; HBG1 |
| HBG2 | GDB:119301 | HEMOGLOBIN, GAMMA G; HBG2 |
| HMBS | GDB:120528 | PORPHYRIA, ACUTE INTERMITTENT; AIP |

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| HND | GDB:9954478 | HARTNUP DISORDER |
| HOMG2 | GDB:9956484 | MAGNESIUM WASTING, RENAL |
| HRAS | GDB:120684 | BLADDER CANCER V-HA-RAS HARVEY RAT SARCOMA VIRAL ONCOGENE HOMOLOG; HRAS |
| HVBS1 | GDB:120069 | CANCER, HEPATOCELLULAR |
| IDDM2 | GDB:128530 | DIABETES MELLITUS, INSULIN-DEPENDENT, 2 DIABETES MELLITUS, JUVENILE-ONSET INSULIN-DEPENDENT; IDDM |
| IGER | GDB:119696 | IgE RESPONSIVENESS, ATOPIC; IGER |
| INS | GDB:119349 | INSULIN; INS |
| JBS | GDB:120111 | JACOBSEN SYNDROME; JBS |
| KCNJ11 | GDB:7009893 | POTASSIUM CHANNEL, INWARDLY-RECTIFYING, SUBFAMILY J, MEMBER 11; KCNJ11 PERSISTENT HYPERINSULINEMIC HYPOGLYCEMIA OF INFANCY |
| KCNJ1 | GDB:204206 | POTASSIUM CHANNEL, INWARDLY-RECTIFYING, SUBFAMILY J, MEMBER 1; KCNJ1 |
| KCNQ1 | GDB:741244 | LONG QT SYNDROME, TYPE 1; LQT1 |
| LDHA | GDB:120141 | LACTATE DEHYDROGENASE-A; LDHA |
| LRP5 | GDB:9836818 | OSTEOPOROSIS-PSEUDOGLIOMA SYNDROME; OPPG HIGH BONE MASS |

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| MEN1 | GDB:120173 | MULTIPLE ENDOCRINE NEOPLASIA, TYPE 1; MEN1 |
| MLL | GDB:128819 | MYELOID/LYMPHOID OR MIXED-LINEAGE LEUKEMIA; MLL |
| MTACR1 | GDB:125743 | MULTIPLE TUMOR ASSOCIATED CHROMOSOME REGION 1; MTACR1 |
| MYBPC3 | GDB:579615 | CARDIOMYOPATHY, FAMILIAL HYPERTROPHIC, 4; CMH4 MYOSIN-BINDING PROTEIN C, CARDIAC; MYBPC3 |
| MYO7A | GDB:132543 | MYOSIN VIIA; MYO7A DEAFNESS, NEUROSENSORY, AUTOSOMAL RECESSIVE, 2; DFNB2 DEAFNESS, AUTOSOMAL DOMINANT NONSYNDROMIC SENSORINEURAL, 11; DFNA11 |
| NNO1 | GDB:10450513 | SIMPLE, AUTOSOMAL DOMINANT |
| OPPG | GDB:3789438 | OSTEOPOROSIS-PSEUDOGLIOMA SYNDROME; OPPG |
| OPTB1 | GDB:9954474 | OSTEOPETROSIS, AUTOSOMAL RECESSIVE |
| PAX6 | GDB:118997 | PAIRED BOX HOMEOTIC GENE 6; PAX6 |
| PC | GDB:119472 | PYRUVATE CARBOXYLASE DEFICIENCY |
| PDX1 | GDB:9836634 | PYRUVATE DEHYDROGENASE COMPLEX, COMPONENT X |
| PGL2 | GDB:511177 | PARAGANGLIOMAS, FAMILIAL NONCHROMAFFIN, 2; PGL2 |

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| PGR | GDB:119493 | PROGESTERONE RESISTANCE |
| PORC | GDB:128610 | PORPHYRIA, CHESTER TYPE; PORC |
| PTH | GDB:119522 | PARATHYROID HORMONE; PTH |
| PTS | GDB:118856 | 6-@PYRUVOYLtetrahydropterin SYNTHASE; PTS |
| PVRL1 | GDB:583951 | ECTODERMAL DYSPLASIA, CLEFT LIP AND PALATE, HAND AND FOOT DEFORMITY, DYSPLASIA, MARGARITA TYPE POLIOVIRUS RECEPTOR RELATED; PVRR |
| PYGM | GDB:120329 | GLYCOGEN STORAGE DISEASE V |
| RAG1 | GDB:120334 | RECOMBINATION ACTIVATING GENE-1; RAG1 |
| RAG2 | GDB:125186 | RECOMBINATION ACTIVATING GENE-2; RAG2 |
| ROM1 | GDB:120350 | ROD OUTER SEGMENT PROTEIN-1; ROM1 |
| SAA1 | GDB:120364 | SERUM AMYLOID A1; SAA1 |
| SCA5 | GDB:378219 | SPINOCEREBELLAR ATAXIA 5; SCA5 |
| SCZD2 | GDB:118874 | DISORDER-2; SCZD2 |
| SDHD | GDB:132456 | PARAGANGLIOMAS, FAMILIAL NONCHROMAFFIN, 1; PGL1 |
| SERPING1 | GDB:119041 | ANGIONEUROTIC EDEMA, HEREDITARY; HANE |

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| SMPD1 | GDB:128144 | NIEMANN-PICK DISEASE |
| TCIRG1 | GDB:9956269 | OSTEOPETROSIS, AUTOSOMAL RECESSIVE |
| TCL2 | GDB:9954468 | LEUKEMIA, ACUTE T-CELL; ATL |
| TECTA | GDB:6837718 | DEAFNESS, AUTOSOMAL DOMINANT NONSYNDROMIC SENSORINEURAL, 8; DFNA8 DEAFNESS, AUTOSOMAL DOMINANT NONSYNDROMIC SENSORINEURAL, 12; DFNA12 |
| TH | GDB:119612 | TYROSINE HYDROXYLASE; TH |
| TREH | GDB:9958953 | TREHALASE |
| TSG101 | GDB:1313414 | TUMOR SUSCEPTIBILITY GENE 101; TSG101 |
| TYR | GDB:120476 | ALBINISM I |
| USH1C | GDB:132544 | USHER SYNDROME, TYPE IC; USH1C |
| VMD2 | GDB:133795 | VITELLIFORM MACULAR DYSTROPHY; VMD2 |
| VRNI | GDB:135662 | VITREORETINOPATHY, NEOVASCULAR INFLAMMATORY; VRNI |
| WT1 | GDB:120496 | FRASIER SYNDROME WILMS TUMOR; WT1 |
| WT2 | GDB:118886 | MULTIPLE TUMOR ASSOCIATED CHROMOSOME REGION 1; MTACR1 |
| ZNF145 | GDB:230064 | PROMYELOCYTIC LEUKEMIA ZINC FINGER; PLZF |

Table 14: Genes, Locations and Genetic Disorders on Chromosome 12

| Gene | GDB Accession ID | OMIM Link |
|--------|------------------|--|
| A2M | GDB:119639 | ALPHA-2-MACROGLOBULIN; A2M |
| AAAS | GDB:9954498 | GLUCOCORTICOID DEFICIENCY AND ACHALASIA |
| ACADS | GDB:118959 | ACYL-CoA DEHYDROGENASE, SHORT-CHAIN; ACADS |
| ACLS | GDB:136346 | ACROCALLOSAL SYNDROME; ACLS |
| ACVRL1 | GDB:230240 | OSLER-RENDU-WEBER SYNDROME 2; ORW2 ACTIVIN A RECEPTOR, TYPE II-LIKE KINASE 1; ACVRL1 |
| ADHR | GDB:9954488 | VITAMIN D-RESISTANT RICKETS, AUTOSOMAL DOMINANT |
| ALDH2 | GDB:119668 | ALDEHYDE DEHYDROGENASE-2; ALDH2 |
| AMHR2 | GDB:696210 | ANTI-MULLERIAN HORMONE TYPE II RECEPTOR; AMHR2 |
| AOM | GDB:118998 | STICKLER SYNDROME, TYPE I; STL1 |
| AQP2 | GDB:141853 | AQUAPORIN-2; AQP2 DIABETES INSIPIDUS, RENAL TYPE DIABETES INSIPIDUS, RENAL TYPE, AUTOSOMAL RECESSIVE |
| ATD | GDB:696353 | ASPHYXIATING THORACIC DYSTROPHY; ATD |

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| ATP2A2 | GDB:119717 | ATPase, Ca(2+)-TRANSPORTING, SLOW-TWITCH; ATP2A2 DARIER-WHITE DISEASE; DAR |
| BDC | GDB:5584359 | BRACHYDACTYLY, TYPE C; BDC |
| C1R | GDB:119729 | COMPLEMENT COMPONENT-C1r, DEFICIENCY OF |
| CD4 | GDB:119767 | T-CELL ANTIGEN T4/LEU3; CD4 |
| CDK4 | GDB:204022 | CYCLIN-DEPENDENT KINASE 4; CDK4 |
| CNA1 | GDB:252119 | CORNEA PLANA 1; CNA1 |
| COL2A1 | GDB:119063 | STICKLER SYNDROME, TYPE I; STL1 COLLAGEN, TYPE II, ALPHA-1 CHAIN; COL2A1 ACHONDROGENESIS, TYPE II; ACG2 |
| CYP27B1 | GDB:9835730 | PSEUDOVITAMIN D DEFICIENCY RICKETS; PDDR |
| DRPLA | GDB:270336 | DENTATORUBRAL-PALLIDOLUYSIAN ATROPHY; DRPLA |
| ENUR2 | GDB:666422 | ENURESIS, NOCTURNAL, 2; ENUR2 |
| FEOM1 | GDB:345037 | FIBROSIS OF EXTRAOCULAR MUSCLES, CONGENITAL; FEOM |
| FPF | GDB:9848880 | PERIODIC FEVER, AUTOSOMAL DOMINANT |
| GNB3 | GDB:120005 | GUANINE NUCLEOTIDE-BINDING PROTEIN, BETA POLYPEPTIDE-3; GNB3 |

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| GNS | GDB:120006 | MUCOPOLYSACCHARIDOSIS TYPE III |
| HAL | GDB:120746 | HISTIDINEMIA |
| HBP1 | GDB:701889 | BRACHYDACTYLY WITH HYPERTENSION |
| HMGIC | GDB:362658 | HIGH MOBILITY GROUP PROTEIN ISOFORM I-C; HMGIC |
| HMN2 | GDB:9954508 | MUSCULAR ATROPHY, ADULT SPINAL |
| HPD | GDB:135978 | TYROSINEMIA, TYPE III |
| IGF1 | GDB:120081 | INSULINLIKE GROWTH FACTOR 1; IGF1 |
| KCNA1 | GDB:127903 | POTASSIUM VOLTAGE-GATED CHANNEL, SHAKER-RELATED SUBFAMILY, MEMBER |
| KERA | GDB:252121 | CORNEA PLANA 2; CNA2 |
| KRAS2 | GDB:120120 | V-KI-RAS2 KIRSTEN RAT SARCOMA 2 VIRAL ONCOGENE HOMOLOG; KRAS2 |
| KRT1 | GDB:128198 | KERATIN 1; KRT1 |
| KRT2A | GDB:407640 | ICHTHYOSIS, BULLOUS TYPE KERATIN 2A; KRT2A |
| KRT3 | GDB:136276 | KERATIN 3; KRT3 |

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| KRT4 | GDB:120697 | KERATIN 4; KRT4 |
| KRT5 | GDB:128110 | EPIDERMOLYSIS BULLOSA HERPETIFORMIS, DOWLING-MEARA TYPE KERATIN 5; KRT5 |
| KRT6A | GDB:128111 | KERATIN 6A; KRT6A |
| KRT6B | GDB:128113 | KERATIN 6B; KRT6B PACHYONYCHIA CONGENITA, JACKSON-LAWLER TYPE |
| KRTHB6 | GDB:702078 | MONILETHRICK KERATIN, HAIR BASIC (TYPE II) 6 |
| LDHB | GDB:120147 | LACTATE DEHYDROGENASE-B; LDHB |
| LYZ | GDB:120160 | AMYLOIDOSIS, FAMILIAL VISCERAL LYSOZYME; LYZ |
| MGCT | GDB:9954504 | TESTICULAR TUMORS |
| MPE | GDB:120191 | MALIGNANT PROLIFERATION OF |
| MVK | GDB:134189 | MEVALONICACIDURIA |
| MYL2 | GDB:128829 | MYOSIN, LIGHT CHAIN, REGULATORY VENTRICULAR; MYL2 |
| NS1 | GDB:439388 | NOONAN SYNDROME 1; NS1 |
| OAP | GDB:120245 | OSTEOARTHROSIS, PRECOCIOUS; OAP |

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| PAH | GDB:119470 | PHENYLKETONURIA; PKU1 |
| PPKB | GDB:696352 | PALMOPLANTAR KERATODERMA, BOTHNIAN TYPE; PPKB |
| PRB3 | GDB:119513 | PAROTID SALIVARY GLYCOPROTEIN; G1 |
| PXR1 | GDB:433739 | ZELLWEGER SYNDROME; ZS PEROXISOME RECEPTOR 1; PXR1 |
| RLS | GDB:11501392 | ACROMELALGIA, HEREDITARY |
| RSN | GDB:139158 | RESTIN; RSN |
| SAS | GDB:128054 | SARCOMA AMPLIFIED SEQUENCE; SAS |
| SCA2 | GDB:128034 | SPINOCEREBELLAR ATAXIA 2; SCA2 ATAxin-2; ATX2 |
| SCNN1A | GDB:366596 | SODIUM CHANNEL, NONVOLTAGE-GATED, 1; SCNN1A |
| SMAL | GDB:9954506 | SPINAL MUSCULAR ATROPHY, CONGENITAL NONPROGRESSIVE, OF LOWER LIMBS |
| SPPM | GDB:9954502 | SCAPULOPERONEAL MYOPATHY; SPM |
| SPSMA | GDB:9954510 | SCAPULOPERONEAL AMYOTROPHY, NEUROGENIC, NEW ENGLAND TYPE |
| TBX3 | GDB:681969 | ULNAR-MAMMARY SYNDROME; UMS T-BOX 3; TBX3 |

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| TBX5 | GDB:6175917 | HOLT-ORAM SYNDROME; HOS T-BOX 5; TBX5 |
| TCF1 | GDB:125297 | TRANSCRIPTION FACTOR 1, HEPATIC; TCF1 MATURITY-ONSET DIABETES OF THE YOUNG, TYPE III; MODY3 |
| TPI1 | GDB:119617 | TRIOSEPHOSPHATE ISOMERASE 1; TPI1 |
| TSC3 | GDB:127930 | SCLEROSIS-3; TSC3 |
| ULR | GDB:594089 | UTERINE |
| VDR | GDB:120487 | VITAMIN D-RESISTANT RICKETS WITH END-ORGAN UNRESPONSIVENESS TO 1,25-DIHYDROXYCHOLECALCIFEROL VITAMIN D RECEPTOR; VDR |
| VWF | GDB:119125 | VON WILLEBRAND DISEASE; VWD |

Table 15: Genes, Locations and Genetic Disorders on Chromosome 13

| Gene | GDB Accession ID | OMIM Link |
|-------|------------------|---|
| ATP7B | GDB:120494 | WILSON DISEASE; WND |
| BRCA2 | GDB:387848 | BREAST CANCER 2, EARLY-ONSET; BRCA2 |
| BRCD1 | GDB:9954522 | BREAST CANCER, DUCTAL, 1; BRCD1 |
| CLN5 | GDB:230991 | CEROID-LIPOFUSCINOSIS, NEURONAL 5; CLN5 |

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|-------|-------------|---|
| CPB2 | GDB:129546 | CARBOXYPEPTIDASE B2, PLASMA; CPB2 |
| ED2 | GDB:9834522 | ECTODERMAL DYSPLASIA, HIDROTIC; HED |
| EDNRB | GDB:129075 | ENDOTHELIN-B RECEPTOR; EDNRB HIRSCHSPRUNG DISEASE-2; HSCR2 |
| ENUR1 | GDB:594516 | ENURESIS, NOCTURNAL, 1; ENUR1 |
| ERCC5 | GDB:120515 | EXCISION-REPAIR, COMPLEMENTING DEFECTIVE, IN CHINESE HAMSTER, 5; ERCC5 |
| F10 | GDB:119890 | X, QUANTITATIVE VARIATION IN FACTOR X DEFICIENCY; F10 |
| F7 | GDB:119897 | FACTOR VII DEFICIENCY |
| GJB2 | GDB:125247 | GAP JUNCTION PROTEIN, BETA-2, 26 KD; GJB2 DEAFNESS, NEUROSENSORY, AUTOSOMAL RECESSIVE, 1; DFNB1 DEAFNESS, AUTOSOMAL DOMINANT NONSYNDROMIC SENSORINEURAL, 3; DFNA3 |
| GJB6 | GDB:9958357 | ECTODERMAL DYSPLASIA, HIDROTIC; HED DEAFNESS, AUTOSOMAL DOMINANT NONSYNDROMIC SENSORINEURAL, 3; DFNA3 |
| IPF1 | GDB:448899 | INSULIN PROMOTER FACTOR 1; IPF1 |
| MBS1 | GDB:128365 | MOEBIUS SYNDROME; MBS |
| MCOR | GDB:9954520 | CONGENITAL |

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| PCCA | GDB:119473 | GLYCINEMIA, KETOTIC, I |
| RB1 | GDB:118734 | BLADDER CANCER RETINOBLASTOMA; RB1 |
| RHOK | GDB:371598 | RHODOPSIN KINASE; RHOK |
| SCZD7 | GDB:9864734 | DISORDER-2; SCZD2 |
| SGCG | GDB:3763329 | MUSCULAR DYSTROPHY, LIMB GIRDLE, TYPE 2C; LGMD2C |
| SLC10A2 | GDB:677534 | SOLUTE CARRIER FAMILY 10, MEMBER 2; SLC10A2 |
| SLC25A15 | GDB:120042 | HYPERNORNITHINEMIA-HYPERAMMONEMIA-HOMOCITRULLINURIA SYNDROME |
| STARP1 | GDB:635459 | STEROIDOGENIC ACUTE REGULATORY PROTEIN; STAR |
| ZNF198 | GDB:6382650 | ZINC FINGER PROTEIN-198; ZNF198 |

Table 16: Genes, Locations and Genetic Disorders on Chromosome 14

| Gene | GDB Accession ID | OMIM Link |
|-------|------------------|--|
| ACHM1 | GDB:132458 | COLORBLINDNESS, TOTAL |
| ARVD1 | GDB:371339 | ARRHYTHMOGENIC RIGHT VENTRICULAR DYSPLASIA, FAMILIAL, 1; ARVD1 |
| CTAA1 | GDB:265299 | CATARACT, ANTERIOR POLAR 1; CTAA1 |
| DAD1 | GDB:407505 | DEFENDER AGAINST CELL DEATH; DAD1 |

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| DFNB5 | GDB:636176 | DEAFNESS, NEUROSENSORY, AUTOSOMAL RECESSIVE, 5; DFNB5 |
| EML1 | GDB:6328385 | USHER SYNDROME, TYPE IA; USH1A |
| GALC | GDB:119970 | KRABBE DISEASE |
| GCH1 | GDB:118798 | DYSTONIA, PROGRESSIVE, WITH DIURNAL VARIATION GTP CYCLOHYDROLASE I DEFICIENCY GTP CYCLOHYDROLASE I; GCH1 |
| HE1 | GDB:9957680 | MALFORMATIONS, MULTIPLE, WITH LIMB ABNORMALITIES AND HYPOPITUITARISM |
| IBGC1 | GDB:10450404 | CEREBRAL CALCIFICATION, NONARTERIOSCLEROTIC |
| IGH@ | GDB:118731 | IgA CONSTANT HEAVY CHAIN 1; IGHA1 IMMUNOGLOBULIN: D (DIVERSITY) REGION OF HEAVY CHAIN IgA CONSTANT HEAVY CHAIN 2; IGHA2 IMMUNOGLOBULIN: J (JOINING) LOCI OF HEAVY CHAIN; IGHJ IMMUNOGLOBULIN: HEAVY Mu CHAIN; Mu1; IGHM1 IMMUNOGLOBULIN: VARIABLE REGION OF HEAVY CHAINS--Hv1; IGHV IgG HEAVY CHAIN LOCUS; IGHG1 IMMUNOGLOBULIN Gm-2; IGHG2 IMMUNOGLOBULIN Gm-3; IGHG3 IMMUNOGLOBULIN Gm-4; IGHG4 IMMUNOGLOBULIN: HEAVY DELTA CHAIN; IGHD IMMUNOGLOBULIN: HEAVY EPSILON CHAIN; IGHE |
| IGHC group | GDB:9992632 | IgA CONSTANT HEAVY CHAIN 1; IGHA1 IgA CONSTANT HEAVY CHAIN 2; IGHA2 IMMUNOGLOBULIN: HEAVY Mu CHAIN; Mu1; IGHM1 IgG HEAVY CHAIN LOCUS; IGHG1 IMMUNOGLOBULIN Gm-2; IGHG2 IMMUNOGLOBULIN Gm-3; IGHG3 IMMUNOGLOBULIN Gm-4; IGHG4 IMMUNOGLOBULIN: HEAVY DELTA CHAIN; |

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| | | IGHD IMMUNOGLOBULIN: HEAVY EPSILON CHAIN; IGHE |
| IGHG1 | GDB:120085 | IgG HEAVY CHAIN LOCUS; IGHG1 |
| IGHM | GDB:120086 | IMMUNOGLOBULIN: HEAVY Mu CHAIN; Mu1; IGHM1 |
| IGHR | GDB:9954529 | G1(A1) SYNDROME |
| IV | GDB:139274 | INVERSUS VISCRUM |
| LTBP2 | GDB:453890 | LATENT TRANSFORMING GROWTH FACTOR-BETA BINDING PROTEIN 2; LTBP2 |
| MCOP | GDB:9954527 | MICROPHTHALMOS |
| MJD | GDB:118840 | MACHADO-JOSEPH DISEASE; MJD |
| MNG1 | GDB:6540062 | GOITER, MULTINODULAR 1; MNG1 |
| MPD1 | GDB:230271 | MYOPATHY, LATE DISTAL HEREDITARY |
| MPS3C | GDB:9954532 | MUCOPOLYSACCHARIDOSIS TYPE IIIC |
| MYH6 | GDB:120214 | MYOSIN, HEAVY POLYPEPTIDE 6; MYH6 |
| MYH7 | GDB:120215 | MYOSIN, CARDIAC, HEAVY CHAIN, BETA; MYH7 |
| NP | GDB:120239 | NUCLEOSIDE PHOSPHORYLASE; NP |
| PABPN1 | GDB:567135 | OCULOPHARYNGEAL MUSCULAR DYSTROPHY; OPMD OCULOPHARYNGEAL |

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| | | MUSCULAR DYSTROPHY, AUTOSOMAL RECESSIVE POLYADENYLATE-BINDING PROTEIN-2; PABP2 |
| PSEN1 | GDB:135682 | ALZHEIMER DISEASE, FAMILIAL, TYPE 3; AD3 |
| PYGL | GDB:120328 | GLYCOGEN STORAGE DISEASE VI |
| RPGRIP1 | GDB:11498766 | AMAUROSIS CONGENITA OF LEBER I |
| SERPINA1 | GDB:120289 | PROTEASE INHIBITOR 1; PI |
| SERPINA3 | GDB:118955 | ALPHA-1-ANTICHYMOTRYPSIN; AACT |
| SERPINA6 | GDB:127865 | CORTICOSTEROID-BINDING GLOBULIN; CBG |
| SLC7A7 | GDB:9863033 | DIBASICAMINOACIDURIA II |
| SPG3A | GDB:230126 | SPASTIC PARAPLEGIA-3, AUTOSOMAL DOMINANT; SPG3A |
| SPTB | GDB:119602 | ELLIPTOCYTOSIS, RHESUS-UNLINKED TYPE HEREDITARY HEMOLYTIC SPECTRIN, BETA, ERYTHROCYTIC; SPTB |
| TCL1A | GDB:250785 | T-CELL LYMPHOMA OR LEUKEMIA |
| TCRAV17S1 | GDB:642130 | T-CELL ANTIGEN RECEPTOR, ALPHA SUBUNIT; TCRA |
| TCRAV5S1 | GDB:451966 | T-CELL ANTIGEN RECEPTOR, ALPHA SUBUNIT; TCRA |

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| TGM1 | GDB:125299 | TRANSGLUTAMINASE 1; TGM1 ICHTHYOSIS CONGENITA |
| TITF1 | GDB:132588 | THYROID TRANSCRIPTION FACTOR 1; TITF1 |
| TMIP | GDB:9954523 | AND ULNA, DUPLICATION OF, WITH ABSENCE OF TIBIA AND RADIUS |
| TRA@ | GDB:120404 | T-CELL ANTIGEN RECEPTOR, ALPHA SUBUNIT; TCRA |
| TSHR | GDB:125313 | THYROTROPIN, UNRESPONSIVENESS TO |
| USH1A | GDB:118885 | USHER SYNDROME, TYPE IA; USH1A |
| VP | GDB:120492 | PORPHYRIA VARIEGATA |

Table 17: Genes, Locations and Genetic Disorders on Chromosome 15

| Gene | GDB Accession ID | OMIM Link |
|-------|------------------|---|
| ACCPN | GDB:5457725 | CORPUS CALLOSUM, AGENESIS OF, WITH NEURONOPATHY |
| AHO2 | GDB:9954535 | HEREDITARY OSTEODYSTROPHY-2; AHO2 |
| ANCR | GDB:119678 | ANGELMAN SYNDROME |
| B2M | GDB:119028 | BETA-2-MICROGLOBULIN; B2M |
| BBS4 | GDB:511199 | BARDET-BIEDL SYNDROME, TYPE 4; BBS4 |
| BLM | GDB:135698 | BLOOM SYNDROME; BLM |

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| CAPN3 | GDB:119751 | CALPAIN, LARGE POLYPEPTIDE L3; CAPN3 MUSCULAR DYSTROPHY, LIMB-GIRDLE, TYPE 2; LGMD2 |
| CDAN1 | GDB:9823267 | DYSERYTHROPOIETIC ANEMIA, CONGENITAL, TYPE I |
| CDAN3 | GDB:386192 | DYSERYTHROPOIETIC ANEMIA, CONGENITAL, TYPE III; CDAN3 |
| CLN6 | GDB:4073043 | CEROID-LIPOFUSCINOSIS, NEURONAL 6, LATE INFANTILE, VARIANT; CLN6 |
| CMH3 | GDB:138299 | CARDIOMYOPATHY, FAMILIAL HYPERTROPHIC, 3; CMH3 |
| CYP19 | GDB:119830 | CYTOCHROME P450, SUBFAMILY XIX; CYP19 |
| CYP1A1 | GDB:120604 | CYTOCHROME P450, SUBFAMILY I, POLYPEPTIDE 1; CYP1A1 |
| CYP1A2 | GDB:118780 | CYTOCHROME P450, SUBFAMILY I, POLYPEPTIDE 2; CYP1A2 |
| DYX1 | GDB:1391796 | DYSLEXIA, SPECIFIC, 1; DYX1 |
| EPB42 | GDB:127385 | HEREDITARY HEMOLYTIC PROTEIN 4.2, ERYTHROCYTIC; EPB42 |
| ETFA | GDB:119121 | GLUTARICACIDURIA IIA; GA IIA |
| EYCL3 | GDB:4590306 | EYE COLOR-3; EYCL3 |
| FAH | GDB:119901 | TYROSINEMIA, TYPE I |

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| FBN1 | GDB:127115 | FIBRILLIN-1; FBN1 MARFAN SYNDROME; MFS |
| FES | GDB:119906 | V-FES FELINE SARCOMA VIRAL/V-FPS FUJINAMI AVIAN SARCOMA VIRAL ONCOGENE |
| HCVS | GDB:119306 | CORONAVIRUS 229E SUSCEPTIBILITY; CVS |
| HEXA | GDB:120040 | TAY-SACHS DISEASE; TSD |
| IVD | GDB:119354 | ISOVALERICACIDEMIA; IVA |
| LCS1 | GDB:11500552 | CHOLESTASIS-LYMPHEDEMA SYNDROME |
| LIPC | GDB:119366 | LIPASE, HEPATIC; LIPC |
| MYO5A | GDB:218824 | MYOSIN VA; MYO5A |
| OCA2 | GDB:136820 | ALBINISM II |
| OTSC1 | GDB:9860473 | OTOSCLEROSIS |
| PWCR | GDB:120325 | PRADER-WILLI SYNDROME |
| RLBP1 | GDB:127341 | RETINALDEHYDE-BINDING PROTEIN 1;; RLBP1 |
| SLC12A1 | GDB:386121 | SOLUTE CARRIER FAMILY 12, MEMBER 1; SLC12A1 |
| SPG6 | GDB:511201 | SPASTIC PARAPLEGIA 6, AUTOSOMAL DOMINANT; SPG6 |

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| TPM1 | GDB:127875 | TROPOMYOSIN 1; TPM1 |
| UBE3A | GDB:228487 | ANGELMAN SYNDROME UBIQUITIN-PROTEIN LIGASE E3A; UBE3A |
| WMS | GDB:5583902 | WEILL-MARCHESANI SYNDROME |

Table 18: Genes, Locations and Genetic Disorders on Chromosome 16

| Gene | GDB Accession ID | OMIM Link |
|--------|------------------|---|
| ABCC6 | GDB:9315106 | PSEUDOXANTHOMA ELASTICUM, AUTOSOMAL DOMINANT; PXE PSEUDOXANTHOMA ELASTICUM, AUTOSOMAL RECESSIVE; PXE |
| ALDOA | GDB:118993 | ALDOLASE A, FRUCTOSE-BISPHOSPHATE; ALDOA |
| APRT | GDB:119003 | ADENINE PHOSPHORIBOSYLTRANSFERASE; APRT |
| ATP2A1 | GDB:119716 | ATPase, Ca(2+)-TRANSPORTING, FAST-TWITCH 1; ATP2A1 BRODY MYOPATHY |
| BBS2 | GDB:229992 | BARDET-BIEDL SYNDROME, TYPE 2; BBS2 |
| CARD15 | GDB:11026232 | SYNOVITIS, GRANULOMATOUS, WITH UVEITIS AND CRANIAL NEUROPATHIES REGIONAL ENTERITIS |
| CATM | GDB:701219 | MICROPHTHALMIA-CATARACT |
| CDH1 | GDB:120484 | CADHERIN 1; CDH1 |

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| CETP | GDB:119773 | CHOLESTERYL ESTER TRANSFER PROTEIN, PLASMA; CETP |
| CHST6 | GDB:131407 | CORNEAL DYSTROPHY, MACULAR TYPE |
| CLN3 | GDB:120593 | CEROID-LIPOFUSCINOSIS, NEURONAL 3, JUVENILE; CLN3 |
| CREBBP | GDB:437159 | RUBINSTEIN SYNDROME CREB-BINDING PROTEIN; CREBBP |
| CTH | GDB:119086 | CYSTATHIONINURIA |
| CTM | GDB:119819 | CATARACT, ZONULAR |
| CYBA | GDB:125238 | GRANULOMATOUS DISEASE, CHRONIC, AUTOSOMAL CYTOCHROME-b-NEGATIVE FORM |
| CYLD | GDB:701216 | EPITHELIOMA, HEREDITARY MULTIPLE BENIGN CYSTIC |
| DHS | GDB:9958268 | XEROCYTOSIS, HEREDITARY |
| DNASE1 | GDB:132846 | DEOXYRIBONUCLEASE I; DNASE1 |
| DPEP1 | GDB:128059 | RENAL DIPEPTIDASE |
| ERCC4 | GDB:119113 | EXCISION-REPAIR, COMPLEMENTING DEFECTIVE, IN CHINESE HAMSTER, 4; ERCC4 XERODERMA PIGMENTOSUM, COMPLEMENTATION GROUP F; XPF |
| FANCA | GDB:701221 | FANCONI ANEMIA, COMPLEMENTATION GROUP A; FACA |

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| GALNS | GDB:129085 | MUCOPOLYSACCHARIDOSIS TYPE IVA |
| GAN | GDB:9864885 | NEUROPATHY, GIANT AXONAL; GAN |
| HAGH | GDB:119292 | HYDROXYACYL GLUTATHIONE HYDROLASE; HAGH |
| HBA1 | GDB:119293 | HEMOGLOBIN--ALPHA LOCUS-1; HBA1 |
| HBA2 | GDB:119294 | HEMOGLOBIN--ALPHA LOCUS-2; HBA2 |
| HBHR | GDB:9954541 | HEMOGLOBIN H-RELATED MENTAL RETARDATION |
| HBQ1 | GDB:120036 | HEMOGLOBIN--THETA-1 LOCUS; HBQ1 |
| HBZ | GDB:119302 | HEMOGLOBIN--ZETA LOCUS; HBZ |
| HBZP | GDB:120037 | HEMOGLOBIN--ZETA LOCUS; HBZ |
| HP | GDB:119314 | HAPTOGLOBIN; HP |
| HSD11B2 | GDB:409951 | CORTISOL 11-BETA-KETOREDUCTASE DEFICIENCY |
| IL4R | GDB:118823 | INTERLEUKIN-4 RECEPTOR; IL4R |
| LIPB | GDB:119365 | LIPASE B, LYSOSOMAL ACID; LIPB |
| MC1R | GDB:135162 | MELANOCORTIN-1 RECEPTOR; MC1R |
| MEFV | GDB:125263 | MEDITERRANEAN FEVER, FAMILIAL; MEFV |

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| MHC2TA | GDB:6268475 | MHC CLASS II TRANSACTIVATOR; MHC2TA |
| MLYCD | GDB:11500940 | MALONYL CoA DECARBOXYLASE DEFICIENCY |
| PHKB | GDB:120286 | PHOSPHORYLASE KINASE, BETA SUBUNIT; PHKB |
| PHKG2 | GDB:140316 | PHOSPHORYLASE KINASE, TESTIS/LIVER, GAMMA 2; PHKG2 |
| PKD1 | GDB:120293 | POLYCYSTIC KIDNEYS POLYCYSTIC KIDNEY DISEASE 1; PKD1 |
| PKDTS | GDB:9954545 | POLYCYSTIC KIDNEY DISEASE, INFANTILE SEVERE, WITH TUBEROUS SCLEROSIS; |
| PMM2 | GDB:438697 | CARBOHYDRATE-DEFICIENT GLYCOPROTEIN SYNDROME, TYPE I; CDG1 PHOSPHOMANNOMUTASE 2; PMM2 |
| PXE | GDB:6053895 | PSEUDOXANTHOMA ELASTICUM, AUTOSOMAL DOMINANT; PXE PSEUDOXANTHOMA ELASTICUM, AUTOSOMAL RECESSIVE; PXE |
| SALL1 | GDB:4216161 | TOWNES-BROCKS SYNDROME; TBS SAL-LIKE 1; SALL1 |
| SCA4 | GDB:250364 | SPINOCEBELLAR ATAXIA 4; SCA4 |
| SCNN1B | GDB:434471 | SODIUM CHANNEL, NONVOLTAGE-GATED 1 BETA; SCNN1B |
| SCNN1G | GDB:568759 | SODIUM CHANNEL, NONVOLTAGE-GATED 1 GAMMA; SCNN1G |

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| TAT | GDB:120398 | TYROSINE TRANSAMINASE DEFICIENCY |
| TSC2 | GDB:120466 | TUBEROUS SCLEROSIS-2; TSC2 |
| VDI | GDB:119629 | DEFECTIVE INTERFERING PARTICLE INDUCTION, CONTROL OF |
| WT3 | GDB:9958957 | WILMS TUMOR, TYPE III; WT3 |

Table 19: Genes, Locations and Genetic Disorders on Chromosome 17

| Gene | GDB Accession ID | OMIM Link |
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| ABR | GDB:119642 | ACTIVE BCR-RELATED GENE; ABR |
| ACACA | GDB:120534 | ACETYL-CoA CARBOXYLASE DEFICIENCY |
| ACADVL | GDB:1248185 | ACYL-CoA DEHYDROGENASE, VERY-LONG-CHAIN, DEFICIENCY OF |
| ACE | GDB:119840 | Dipeptidyl Carboxypeptidase-1; DCP1 |
| ALDH3A2 | GDB:1316855 | SJOGREN-LARSSON SYNDROME; SLS |
| APOH | GDB:118887 | APOLIPOPROTEIN H; APOH |
| ASPA | GDB:231014 | SPONGY DEGENERATION OF CENTRAL NERVOUS SYSTEM |
| AXIN2 | GDB:9864782 | CANCER OF COLON |
| BCL5 | GDB:125178 | LEUKEMIA/LYMPHOMA, CHRONIC B-CELL, 5; BCLS |

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| BHD | GDB:11498904 | WITH TRICHODISCOMAS AND ACROCHORDONS |
| BLMH | GDB:3801467 | BLEOMYCIN HYDROLASE |
| BRCA1 | GDB:126611 | BREAST CANCER, TYPE 1; BRCA1 |
| CACD | GDB:5885801 | CHOROIDAL DYSTROPHY, CENTRAL AREOLAR; CACD |
| CCA1 | GDB:118763 | CATARACT, CONGENITAL, CERULEAN TYPE 1; CCA1 |
| CCZS | GDB:681973 | CATARACT, CONGENITAL ZONULAR, WITH SUTURAL OPACITIES; CCZS |
| CHRNBI | GDB:120587 | CHOLINERGIC RECEPTOR, NICOTINIC, BETA POLYPEPTIDE 1; CHRNBI |
| CHRNE | GDB:132246 | CHOLINERGIC RECEPTOR, NICOTINIC, EPSILON POLYPEPTIDE; CHRNE |
| CMT1A | GDB:119785 | CHARCOT-MARIE-TOOTH DISEASE, TYPE 1A; CMT1A NEUROPATHY, HEREDITARY, WITH LIABILITY TO PRESSURE PALSIES; HNPP |
| COL1A1 | GDB:119061 | COLLAGEN, TYPE I, ALPHA-1 CHAIN; COL1A1 OSTEOGENESIS IMPERFECTA TYPE I OSTEOGENESIS IMPERFECTA TYPE IV; OI4 |
| CORDS | GDB:568473 | CONE-ROD DYSTROPHY-5; CORDS |
| CTNS | GDB:700761 | CYSTINOSIS, EARLY-ONSET OR INFANTILE NEPHROPATHIC TYPE |

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| EPX | GDB:377700 | EOSINOPHIL PEROXIDASE; EPX |
| ERBB2 | GDB:120613 | V-ERB-B2 AVIAN ERYTHROBLASTIC LEUKEMIA VIRAL ONCOGENE HOMOLOG 2; ERBB2 |
| G6PC | GDB:231927 | GLYCOGEN STORAGE DISEASE I; GSD-I |
| GAA | GDB:119965 | GLYCOGEN STORAGE DISEASE II |
| GALK1 | GDB:119246 | GALACTOKINASE DEFICIENCY |
| GCGR | GDB:304516 | GLUCAGON RECEPTOR; GCGR |
| GFAP | GDB:118799 | GLIAL FIBRILLARY ACIDIC PROTEIN; GFAP ALEXANDER DISEASE |
| GH1 | GDB:119982 | GROWTH HORMONE 1; GH1 |
| GH2 | GDB:119983 | GROWTH HORMONE 2; GH2 |
| GP1BA | GDB:118806 | GIANT PLATELET SYNDROME |
| GPSC | GDB:9954564 | FAMILIAL PROGRESSIVE SUBCORTICAL |
| GUCY2D | GDB:136012 | AMAUROSIS CONGENITA OF LEBER I GUANYLATE CYCLASE 2D, MEMBRANE; GUC2D CONE-ROD DYSTROPHY-6; CORD6 |
| ITGA2B | GDB:120012 | THROMBASTHENIA OF GLANZMANN AND NAEGELI |
| ITGB3 | GDB:120013 | INTEGRIN, BETA-3; ITGB3 |
| ITGB4 | GDB:128028 | INTEGRIN, BETA-4; ITGB4 |

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| KRT10 | GDB:118828 | KERATIN 10; KRT10 |
| KRT12 | GDB:5583953 | CORNEAL DYSTROPHY, JUVENILE EPITHELIAL, OF MEESMANN KERATIN 12; KRT12 |
| KRT13 | GDB:120740 | KERATIN 13; KRT13 |
| KRT14 | GDB:132145 | KERATIN 14; KRT14 GLUTATHIONE SYNTHETASE; GSS |
| KRT14L1 | GDB:120121 | KERATIN 14; KRT14 |
| KRT14L2 | GDB:120122 | KERATIN 14; KRT14 |
| KRT14L3 | GDB:120123 | KERATIN 14; KRT14 |
| KRT16 | GDB:136207 | KERATIN 16; KRT16 |
| KRT16L1 | GDB:120125 | KERATIN 16; KRT16 |
| KRT16L2 | GDB:120126 | KERATIN 16; KRT16 |
| KRT17 | GDB:136211 | KERATIN 17; KRT17 PACHYONYCHIA CONGENITA, JACKSON-LAWLER TYPE |
| KRT9 | GDB:303970 | HYPERKERATOSIS, LOCALIZED EPIDERMOLYTIC |
| MAPT | GDB:119434 | MICROTUBULE-ASSOCIATED PROTEIN TAU; MAPT PALLIDOPONTONIGRAL DEGENERATION; PPND DISINHIBITION-DEMENTIA-PARKINSONIS M-AMYOTROPHY COMPLEX; DDPAC |
| MDB | GDB:9958959 | MEDULLOBLASTOMA; MDB |

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| MDCR | GDB:120525 | MILLER-DIEKER LISSENCEPHALY SYNDROME; MDLS PLATELET-ACTIVATING FACTOR ACETYLHYDROLASE, GAMMA SUBUNIT |
| MGI | GDB:9954550 | MYASTHENIA GRAVIS, FAMILIAL INFANTILE; FIMG |
| MHS2 | GDB:132580 | MALIGNANT HYPERTERMIA SUSCEPTIBILITY-2; MHS2 |
| MKS1 | GDB:681967 | MECKEL SYNDROME; MKS |
| MPO | GDB:120192 | MYELOPEROXIDASE DEFICIENCY |
| MUL | GDB:636050 | MULIBREY NANISM; MUL |
| MYO15A | GDB:9838006 | DEAFNESS, NEUROSENSORY, AUTOSOMAL RECESSIVE, 3; DFNB3 |
| NAGLU | GDB:636533 | MUCOPOLYSACCHARIDOSIS TYPE IIIB |
| NAPB | GDB:9954572 | NEURITIS WITH BRACHIAL PREDILECTION; NAPB |
| NF1 | GDB:120231 | NEUROFIBROMATOSIS, TYPE I; NF1 |
| NME1 | GDB:127965 | NON-METASTATIC CELLS 1, PROTEIN EXPRESSED IN; NME1 |
| P4HB | GDB:120708 | PROLYL-4-HYDROXYLASE, BETA POLYPEPTIDE; PHDB; PROHB |
| PAFAH1B1 | GDB:677430 | MILLER-DIEKER LISSENCEPHALY SYNDROME; MDLS PLATELET-ACTIVATING FACTOR ACETYLHYDROLASE, GAMMA SUBUNIT |

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| PECAM1 | GDB:696372 | PLATELET-ENDOTHELIAL CELL ADHESION MOLECULE; PECAM1 |
| PEX12 | GDB:6155804 | ZELLWEGER SYNDROME; ZS PEROXIN-12; PEX12 |
| PHB | GDB:126600 | PROHIBITIN; PHB |
| PMP22 | GDB:134190 | CHARCOT-MARIE-TOOTH DISEASE, TYPE 1A; CMT1A HYPERTROPHIC NEUROPATHY OF DEJERINE-SOTTAS PERIPHERAL MYELIN PROTEIN 22; PMP22 |
| PRKAR1A | GDB:120313 | MYXOMA, SPOTTY PIGMENTATION, AND ENDOCRINE OVERACTIVITY PROTEIN KINASE, cAMP-DEPENDENT, REGULATORY, TYPE I, ALPHA; PRKAR1A |
| PRKCA | GDB:128015 | PROTEIN KINASE C, ALPHA; PRKCA |
| PRKWNK4 | GDB:9954566 | PSEUDOHYPOALDOSTERONISM TYPE II, LOCUS B; PHA2B |
| PRP8 | GDB:9957697 | RETINITIS PIGMENTOSA-13; RP13 |
| PRPF8 | GDB:392647 | RETINITIS PIGMENTOSA-13; RP13 |
| PTLAH | GDB:9957342 | APLASIA OR HYPOPLASIA |
| RARA | GDB:120337 | RETINOIC ACID RECEPTOR, ALPHA; RARA |
| RCV1 | GDB:135477 | RECOVERIN; RCV1 |
| RMSA1 | GDB:304519 | REGULATOR OF MITOTIC SPINDLE ASSEMBLY 1; RMSA1 |

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| RP17 | GDB:683199 | RETINITIS PIGMENTOSA-17; RP17 |
| RSS | GDB:439249 | RUSSELL-SILVER SYNDROME; RSS |
| SCN4A | GDB:125181 | PERIODIC PARALYSIS II |
| SERPINF2 | GDB:120301 | PLASMIN INHIBITOR DEFICIENCY |
| SGCA | GDB:384077 | ADHALIN; ADL |
| SGSH | GDB:1319101 | MUCOPOLYSACCHARIDOSIS TYPE IIIA |
| SHBG | GDB:125280 | SEX HORMONE BINDING GLOBULIN; SHBG |
| SLC2A4 | GDB:119997 | SOLUTE CARRIER FAMILY 2, MEMBER 4; SLC2A4 |
| SLC4A1 | GDB:119874 | SOLUTE CARRIER FAMILY 4, ANION EXCHANGER, MEMBER 1; SLC4A1 BLOOD GROUP--DIEGO SYSTEM; DI BLOOD GROUP--WRIGHT ANTIGEN; Wr ELLIPTOCYTOSIS, RHESUS-UNLINKED TYPE HEREDITARY HEMOLYTIC |
| SLC6A4 | GDB:134713 | SOLUTE CARRIER FAMILY 6, MEMBER 4; SLC6A4 |
| SMCR | GDB:120379 | SMITH-MAGENIS SYNDROME; SMS |
| SOST | GDB:10450629 | SCLEROSTEOSIS |
| SOX9 | GDB:134730 | DYSPLASIA |
| SSTR2 | GDB:134186 | SOMATOSTATIN RECEPTOR-2; SSTR2 |

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| SYM1 | GDB:512174 | SYMPHALANGISM, PROXIMAL; SYM1 |
| SYNS1 | GDB:9862343 | SYNOSTOSES, MULTIPLE, WITH BRACHYDACTYLY |
| TCF2 | GDB:125298 | TRANSCRIPTION FACTOR-2, HEPATIC; TCF2 |
| THRA | GDB:120730 | THYROID HORMONE RECEPTOR, ALPHA 1; THRA |
| TIMP2 | GDB:132612 | TISSUE INHIBITOR OF METALLOPROTEINASE-2; TIMP2 |
| TOC | GDB:451978 | TYLOSIS WITH ESOPHAGEAL CANCER; TOC |
| TOP2A | GDB:118884 | TOPOISOMERASE (DNA) II, ALPHA; TOP2A |
| TP53 | GDB:120445 | CANCER, HEPATOCELLULAR LI-FRAUMENI SYNDROME; LFS TUMOR PROTEIN p53; TP53 CARCINOMA |
| VBCH | GDB:9954554 | HYPERTOSTOSIS CORTICALIS GENERALISATA |

Table 20: Genes, Locations and Genetic Disorders on Chromosome 18

| Gene | GDB Accession ID | OMIM Link |
|--------|------------------|---|
| ATP8B1 | GDB:453352 | CHOLESTASIS, PROGRESSIVE FAMILIAL INTRAHEPATIC 1; PFIC1 INTRAHEPATIC CHOLESTASIS FAMILIAL INTRAHEPATIC CHOLESTASIS-1; FIC1 |
| BCL2 | GDB:119031 | B-CELL CLL/LYMPHOMA 2; BCL2 |

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| CNSN | GDB:9954580 | CARNOSINEMIA |
| CORD1 | GDB:118773 | CONE-ROD DYSTROPHY-1; CORD1 |
| CYB5 | GDB:125236 | METHEMOGLOBINEMIA DUE TO DEFICIENCY OF CYTOCHROME b5 |
| DCC | GDB:119838 | DELETED IN COLORECTAL CARCINOMA; DCC |
| F5F8D | GDB:6919858 | FACTOR V AND FACTOR VIII, COMBINED DEFICIENCY OF; F5F8D |
| FECH | GDB:127282 | PROTOPORPHYRIA, ERYTHROPOIETIC |
| FEO | GDB:4378120 | POLYOSTOTIC OSTEOLYTIC DYSPLASIA, HEREDITARY EXPANSILE; HEPOD |
| LAMA3 | GDB:251818 | LAMININ, ALPHA 3; LAMA3 |
| LCFS2 | GDB:9954578 | CANCER |
| MADH4 | GDB:4642788 | POLYPOSIS, JUVENILE INTESTINAL MOTHERS AGAINST DECAPENTAPLEGIC, DROSOPHILA, HOMOLOG OF, 4; MADH4 |
| MAFD1 | GDB:120163 | MANIC-DEPRESSIVE PSYCHOSIS, AUTOSOMAL |
| MC2R | GDB:135163 | ADRENAL UNRESPONSIVENESS TO ACTH |
| MCL | GDB:9954574 | LEIOMYOMATA, HEREDITARY MULTIPLE, OF SKIN |
| MYP2 | GDB:9862232 | MYOPIA |

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| NPC1 | GDB:138178 | NIEMANN-PICK DISEASE, TYPE C1; NPC1 |
| SPPK | GDB:606444 | PALMOPLANTARIS STRIATA |
| TGFBRE | GDB:250852 | TRANSFORMING GROWTH FACTOR, BETA 1 RESPONSE ELEMENT |
| TGIF | GDB:9787150 | HOLOPROSENCEPHALY, TYPE 4; HPE4 |
| TTR | GDB:119471 | TRANSTHYRETIN; TTR |

Table 21: Genes, Locations and Genetic Disorders on Chromosome 19

| Gene | GDB Accession ID | OMIM Link |
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| AD2 | GDB:118748 | ALZHEIMER DISEASE-2; AD2 |
| AMH | GDB:118996 | PERSISTENT MULLERIAN DUCT SYNDROME, TYPES I AND II; PMDS ANTI-MULLERIAN HORMONE; AMH |
| APOC2 | GDB:119689 | APOLIPOPROTEIN C-II DEFICIENCY, TYPE I HYPERLIPOPROTEINEMIA DUE TO |
| APOE | GDB:119691 | APOLIPOPROTEIN E; APOE |
| ATHS | GDB:128803 | LIPOPROTEIN PHENOTYPE; ALP |
| BAX | GDB:228082 | BCL2-ASSOCIATED X PROTEIN; BAX |
| BCKDHA | GDB:119723 | MAPLE SYRUP URINE DISEASE |
| BCL3 | GDB:120561 | B-CELL LEUKEMIA/LYMPHOMA-3; BCL3 |

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| BFIC | GDB:9954584 | BENIGN FAMILIAL INFANTILE CONVULSIONS |
| C3 | GDB:119044 | COMPLEMENT COMPONENT-3; C3 |
| CACNA1A | GDB:126432 | ATAXIA, PERIODIC VESTIBULOCEREBELLAR HEMIPLEGIC MIGRAINE, FAMILIAL; MHP SPINOCEREBELLAR ATAXIA 6; SCA6 CALCIUM CHANNEL, VOLTAGE-DEPENDENT, P/Q TYPE, ALPHA 1A SUBUNIT; CACNA1A |
| CCO | GDB:119755 | CENTRAL CORE DISEASE OF MUSCLE |
| CEACAM5 | GDB:119054 | CARCINOEMBRYONIC ANTIGEN; CEA |
| COMP | GDB:344263 | EPIPHYSEAL DYSPLASIA, MULTIPLE; MED PSEUDOACHONDROPLASTIC DYSPLASIA CARTILAGE OLIGOMERIC MATRIX PROTEIN; COMP |
| CRX | GDB:333932 | CONE-ROD DYSTROPHY-2; CORD2 AMAUROSIS CONGENITA OF LEBER I CONE-ROD HOME BOX-CONTAINING GENE |
| DBA | GDB:9600353 | ANEMIA, CONGENITAL HYPOPLASTIC, OF BLACKFAN AND DIAMOND |
| DDU | GDB:10796026 | URTICARIA; DDU |
| DFNA4 | GDB:606540 | DEAFNESS, AUTOSOMAL DOMINANT NONSYNDROMIC SENSORINEURAL, 4; DFNA4 |
| DLL3 | GDB:9959026 | VERTEBRAL ANOMALIES |

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| DMPK | GDB:119097 | DYSTROPHIA MYOTONICA; DM |
| DMWD | GDB:7178354 | DYSTROPHIA MYOTONICA; DM |
| DPD1 | GDB:10796170 | ENGELMANN DISEASE |
| E11S | GDB:119101 | ECHO 11 SENSITIVITY; E11S |
| ELA2 | GDB:118792 | ELASTASE-2; ELA2 NEUTROPENIA, CYCLIC |
| EPOR | GDB:125242 | ERYTHROPOIETIN RECEPTOR; EPOR |
| ERCC2 | GDB:119112 | EXCISION-REPAIR, COMPLEMENTING DEFECTIVE, IN CHINESE HAMSTER, 2; ERCC2 XERODERMA PIGMENTOSUM IV; XP4 |
| ETFB | GDB:119887 | ELECTRON TRANSFER FLAVOPROTEIN, BETA POLYPEPTIDE; ETFB |
| EXT3 | GDB:383780 | EXOSTOSES, MULTIPLE, TYPE III; EXT3 |
| EYCL1 | GDB:119269 | EYE COLOR-1; EYCL1 |
| FTL | GDB:119234 | FERRITIN LIGHT CHAIN; FTL |
| FUT1 | GDB:120618 | FUCOSYLTRANSFERASE-1; FUT1 |
| FUT2 | GDB:120619 | FUCOSYLTRANSFERASE-2; FUT2 |
| FUT6 | GDB:135180 | FUCOSYLTRANSFERASE-6; FUT6 |
| GAMT | GDB:1313736 | GUANIDINOACETATE |

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| | | METHYLTRANSFERASE; GAMT |
| GCDH | GDB:136004 | GLUTARICACIDEMIA I |
| GPI | GDB:120015 | GLUCOSEPHOSPHATE ISOMERASE; GPI |
| GUSM | GDB:119291 | GLUCURONIDASE, MOUSE, MODIFIER OF; GUSM |
| HB1 | GDB:9954586 | BUNDLE BRANCH BLOCK |
| HCL1 | GDB:119304 | HAIR COLOR-1; HCL1 |
| HHC2 | GDB:249836 | HYPOCALCIURIC HYPERCALCEMIA, FAMILIAL, TYPE II; HHC2 |
| HHC3 | GDB:9955121 | HYPOCALCIURIC HYPERCALCEMIA, FAMILIAL, TYPE III; HHC3 |
| ICAM3 | GDB:136236 | INTERCELLULAR ADHESION MOLECULE-3; ICAM3 |
| INSR | GDB:119352 | INSULIN RECEPTOR; INSR |
| JAK3 | GDB:376460 | JANUS KINASE 3 JAK3 |
| KLK3 | GDB:119695 | ANTIGEN, PROSTATE-SPECIFIC; APS |
| LDLR | GDB:119362 | HYPERCHOLESTEROLEMIA, FAMILIAL; FHC |
| LHB | GDB:119364 | LUTEINIZING HORMONE, BETA POLYPEPTIDE; LHB |

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| LIG1 | GDB:127274 | LIGASE I, DNA, ATP-DEPENDENT; LIG1 |
| LOH19CR1 | GDB:9837482 | ANEMIA, CONGENITAL HYPOPLASTIC, OF BLACKFAN AND DIAMOND |
| LYL1 | GDB:120158 | LEUKEMIA, LYMPHOID, 1; LYL1 |
| MAN2B1 | GDB:119376 | MANNOSIDOSIS, ALPHA B, LYSOSOMAL |
| MCOLN1 | GDB:10013974 | MUCOLIPIDOSIS IV |
| MDRV | GDB:6306714 | MUSCULAR DYSTROPHY, AUTOSOMAL DOMINANT, WITH RIMMED VACUOLES; MDRV |
| MLLT1 | GDB:136791 | MYELOID/LYMPHOID OR MIXED LINEAGE LEUKEMIA, TRANSLOCATED TO, 1; MLLT1 |
| NOTCH3 | GDB:361163 | DEMENTIA, HEREDITARY MULTI-INFARCT TYPE NOTCH, DROSOPHILA, HOMOLOG OF, 3; NOTCH3 |
| NPHS1 | GDB:342105 | NEPHROSIS 1, CONGENITAL, FINNISH TYPE; NPHS1 |
| OFC3 | GDB:128060 | OROFACIAL CLEFT-3; OFC3 |
| OPA3 | GDB:9954590 | OPTIC ATROPHY, INFANTILE, WITH CHOREA AND SPASTIC PARAPLEGIA |
| PEPD | GDB:120273 | PEPTIDASE D; PEPD |
| PRPF31 | GDB:333911 | RETINITIS PIGMENTOSA 11; RP11 |

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| PRTN3 | GDB:126876 | PROTEINAŞE 3; PRTN3; PR3 |
| PRX | GDB:11501256 | HYPERTROPHIC NEUROPATHY OF DEJERINE-SOTTAS |
| PSG1 | GDB:120321 | PREGNANCY-SPECIFIC BETA-1-GLYCOPROTEIN 1; PSG1 |
| PVR | GDB:120324 | POLIOVIRUS SUSCEPTIBILITY, OR SENSITIVITY; PVS |
| RYR1 | GDB:120359 | CENTRAL CORE DISEASE OF MUSCLE HYPERHERMIA OF ANESTHESIA RYANODINE RECEPTOR-1; RYR1 |
| SLC5A5 | GDB:5892184 | SOLUTE CARRIER FAMILY 5, MEMBER 5; SLC5A5 |
| SLC7A9 | GDB:9958852 | CYSTINURIA, TYPE III; CSNU3 |
| STK11 | GDB:9732383 | PEUTZ-JEGHERS SYNDROME SERINE/THREONINE PROTEIN KINASE 11; STK11 |
| TBXA2R | GDB:127517 | THROMBOXANE A2 RECEPTOR, PLATELET; TBXA2R |
| TGFB1 | GDB:120729 | ENGELMANN DISEASE TRANSFORMING GROWTH FACTOR, BETA-1; TGFB1 |
| TNNI3 | GDB:125309 | TROPONIN I, CARDIAC; TNNI3 |
| TYROBP | GDB:9954457 | POLYCYSTIC LIPOMEMBRANOUS OSTEODYSPLOASIA WITH SCLEROSING LEUKOENCEPHALOPATHY |

Table 22: Genes, Locations and Genetic Disorders on Chromosome 20

| Gene | GDB Accession ID | OMIM Link |
|--------|------------------|---|
| ADA | GDB:119649 | ADENOSINE DEAMINASE; ADA |
| AHCY | GDB:118983 | S-ADENOSYLHOMOCYSTEINE HYDROLASE; AHCY |
| AVP | GDB:119009 | DIABETES INSIPIDUS, NEUROHYPOPHYSAL TYPE ARGinine VASOPRESSIN; AVP |
| CDAN2 | GDB:9823270 | DYSERYTHROPOIETIC ANEMIA, CONGENITAL, TYPE II |
| CDMP1 | GDB:438940 | CHONDRODYSPLASIA, GREBE TYPE CARTILAGE-DERIVED MORPHOGENETIC PROTEIN 1 |
| CHED1 | GDB:3837719 | CORNEAL DYSTROPHY, CONGENITAL ENDOTHELIAL; CHED |
| CHRNA4 | GDB:128169 | CHOLINERGIC RECEPTOR, NEURONAL NICOTINIC, ALPHA POLYPEPTIDE 4; CHRNA4 EPILEPSY, BENIGN NEONATAL; EBN1 |
| CST3 | GDB:119817 | AMYLOIDOSIS VI |
| EDN3 | GDB:119862 | ENDOTHELIN-3; EDN3 WAARDENBURG-SHAH SYNDROME |
| EEGV1 | GDB:127525 | ELECTROENCEPHALogram, LOW-VOLTAGE |
| FTLL1 | GDB:119235 | FERRITIN LIGHT CHAIN; FTL |
| GNAS | GDB:120628 | GUANINE NUCLEOTIDE-BINDING |

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| | | PROTEIN, ALPHA-STIMULATING POLYPEPTIDE; |
| GSS | GDB:637022 | GLUTATHIONE SYNTHETASE DEFICIENCY OF ERYTHROCYTES, HEMOLYTIC ANEMIA PYROGLUTAMICACIDURIA HNF4AGDB:393281DIABETES MELLITUS, AUTOSOMAL DOMINANT TRANSCRIPTION FACTOR 14, HEPATIC NUCLEAR FACTOR; TCF14 |
| JAG1 | GDB:6175920 | CHOLESTASIS WITH PERIPHERAL PULMONARY STENOSIS JAGGED 1; JAG1 |
| KCNQ2 | GDB:9787229 | EPILEPSY, BENIGN NEONATAL; EBN1 POTASSIUM CHANNEL, VOLTAGE-GATED, SUBFAMILY Q, MEMBER 2 |
| MKKS | GDB:9860197 | HYDROMETROCOLPOS SYNDROME |
| NBIA1 | GDB:4252819 | HALLERVORDEN-SPATZ DISEASE |
| PCK1 | GDB:125349 | PHOSPHOENOLPYRUVATE CARBOXYKINASE 1, SOLUBLE; PCK1 |
| PI3 | GDB:203940 | PROTEINASE INHIBITOR 3; PI3 |
| PPGB | GDB:119507 | NEURAMINIDASE DEFICIENCY WITH BETA-GALACTOSIDASE DEFICIENCY |
| PPMD | GDB:702144 | CORNEAL DYSTROPHY, HEREDITARY POLYMORPHOUS POSTERIOR; PPCD |
| PRNP | GDB:120720 | GERSTMANN-STRAUSSLER DISEASE; GSD PRION PROTEIN; PRNP |
| THBD | GDB:119613 | THROMBOMODULIN; THBD |

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| TOP1 | GDB:120444 | TOPOISOMERASE (DNA) I; TOP1 |
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Table 23: Genes, Locations and Genetic Disorders on Chromosome 21

| Gene | GDB Accession ID | OMIM Link |
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| AIRE | GDB:567198 | AUTOIMMUNE POLYENDOCRINOPATHY-CANDIDIASIS-ECTODERMAL DYSTROPHY; APECED |
| APP | GDB:119692 | ALZHEIMER DISEASE; AD AMYLOID BETA A4 PRECURSOR PROTEIN; APP |
| CBS | GDB:119754 | HOMOCYSTINURIA |
| COL6A1 | GDB:119065 | COLLAGEN, TYPE VI, ALPHA-1 CHAIN; COL6A1 MYOPATHY, BENIGN CONGENITAL, WITH CONTRACTURES |
| COL6A2 | GDB:119793 | COLLAGEN, TYPE VI, ALPHA-2 CHAIN; COL6A2 MYOPATHY, BENIGN CONGENITAL, WITH CONTRACTURES |
| CSTB | GDB:5215249 | MYOCLONUS EPILEPSY OF UNVERRICHT AND LUNDBORG CYSTATIN B; CSTB |
| DCR | GDB:125354 | TRISOMY 21 |
| DSCR1 | GDB:731000 | TRISOMY 21 |
| FPDMM | GDB:9954610 | CORE-BINDING FACTOR, RUNT DOMAIN, ALPHA SUBUNIT 2; CBFA2 PLATELET DISORDER, FAMILIAL, WITH ASSOCIATED MYELOID MALIGNANCY |

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| HLCS | GDB:392648 | MULTIPLE CARBOXYLASE DEFICIENCY, BIOTIN-RESPONSIVE; MCD |
| HPE1 | GDB:136065 | HOLOPROSENCEPHALY, FAMILIAL ALOBAR |
| ITGB2 | GDB:120574 | INTEGRIN BETA-2; ITGB2 |
| KCNE1 | GDB:127909 | POTASSIUM VOLTAGE-GATED CHANNEL, ISK-RELATED SUBFAMILY, MEMBER 1; |
| KNO | GDB:4073044 | KNOBLOCH SYNDROME; KNO |
| PRSS7 | GDB:384083 | ENTEROKINASE DEFICIENCY |
| RUNX1 | GDB:128313 | CORE-BINDING FACTOR, RUNT DOMAIN, ALPHA SUBUNIT 2; CBFA2 PLATELET DISORDER, FAMILIAL, WITH ASSOCIATED MYELOID MALIGNANCY |
| SOD1 | GDB:119596 | AMYOTROPHIC LATERAL SCLEROSIS SUPEROXIDE DISMUTASE-1; SOD1 MUSCULAR ATROPHY, PROGRESSIVE, WITH AMYOTROPHIC LATERAL SCLEROSIS |
| TAM | GDB:9958709 | MYELOPROLIFERATIVE SYNDROME, TRANSIENT |

Table 24: Genes, Locations and Genetic Disorders on Chromosome 22

| Gene | GDB Accession ID | OMIM Link |
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| ADSL | GDB:119655 | ADENYLOSUCCINATE LYASE; ADSL |

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| ARSA | GDB:119007 | METACHROMATIC LEUKODYSTROPHY, LATE-INFANTILE |
| BCR | GDB:120562 | BREAKPOINT CLUSTER REGION; BCR |
| CECR | GDB:119772 | CAT EYE SYNDROME; CES |
| CHEK2 | GDB:9958730 | LI-FRAUMENI SYNDROME; LFS OSTEOGENIC SARCOMA |
| COMT | GDB:119795 | CATECHOL-O-METHYLTRANSFERASE; COMT |
| CRYBB2 | GDB:119075 | CRYSTALLIN, BETA B2; CRYBB2 CATARACT, CONGENITAL, CERULEAN TYPE, 2; CCA2 |
| CSF2RB | GDB:126838 | GRANULOCYTE-MACROPHAGE COLONY-STIMULATING FACTOR RECEPTOR, BETA SUBUNIT; |
| CTHM | GDB:439247 | HEART MALFORMATIONS; CTHM |
| CYP2D6 | GDB:132127 | CYTOCHROME P450, SUBFAMILY IID; CYP2D |
| CYP2D@ | GDB:119832 | CYTOCHROME P450, SUBFAMILY IID; CYP2D |
| DGCR | GDB:119843 | DIGEORGE SYNDROME; DGS |
| DIA1 | GDB:119848 | METHEMOGLOBINEMIA DUE TO DEFICIENCY OF METHEMOGLOBIN REDUCTASE |
| EWSR1 | GDB:135984 | EWING SARCOMA; EWS |

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| GGT1 | GDB:120623 | GLUTATHIONURIA |
| MGCR | GDB:120180 | MENINGIOMA; MGM |
| MN1 | GDB:580528 | MENINGIOMA; MGM |
| NAGA | GDB:119445 | ALPHA-GALACTOSIDASE B; GALB |
| NF2 | GDB:120232 | NEUROFIBROMATOSIS, TYPE II; NF2 |
| OGS2 | GDB:9954619 | HYPERTELORISM WITH ESOPHAGEAL ABNORMALITY AND HYPOSPADIAS |
| PDGFB | GDB:120709 | V-SIS PLATELET-DERIVED GROWTH FACTOR BETA POLYPEPTIDE; PDGFB |
| PPARA | GDB:202877 | PEROXISOME PROLIFERATOR ACTIVATED RECEPTOR, ALPHA; PPARA |
| PRODH | GDB:5215168 | HYPERPROLINEMIA, TYPE I |
| SCO2 | GDB:9958568 | CYTOCHROME c OXIDASE DEFICIENCY |
| SCZD4 | GDB:1387047 | SCHIZOPHRENIA DISORDER-4; SCZD4 |
| SERPIND1 | GDB:120038 | HEPARIN COFACTOR II; HCF2 |
| SLC5A1 | GDB:120375 | SOLUTE CARRIER FAMILY 5, MEMBER 1; SLC5A1 |
| SOX10 | GDB:9834028 | SRY-BOX 10; SOX10 |
| TCN2 | GDB:119608 | TRANSCOBALAMIN II DEFICIENCY |

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| TIMP3 | GDB:138175 | TISSUE INHIBITOR OF METALLOPROTEINASE-3; TIMP3 |
| VCF | GDB:136422 | VELOCARDIOFACIAL SYNDROME |

Table 25: Genes, Locations and Genetic Disorders on Chromosome X

| Gene | GDB Accession ID | OMIM Link |
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| ABCD1 | GDB:118991 | ADRENOLEUKODYSTROPHY; ALD |
| ACTL1 | GDB:119648 | ACTIN-LIKE SEQUENCE-1; ACTL1 |
| ADFN | GDB:118977 | ALBINISM-DEAFNESS SYNDROME; ADFN; ALDS |
| AGMX2 | GDB:119661 | AGAMMAGLOBULINEMIA, X-LINKED, TYPE 2; AGMX2; XLA2 |
| AHDS | GDB:125899 | MENTAL RETARDATION, X-LINKED, WITH HYPOTONIA |
| AIC | GDB:118986 | CORPUS CALLOSUM, AGENESIS OF, WITH CHORIORETINAL ABNORMALITY |
| AIED | GDB:119663 | ALBINISM, OCULAR, TYPE 2; OA2 |
| AIH3 | GDB:131443 | AMELOGENESIS IMPERFECTA-3, HYPOPLASTIC TYPE; AIH3 |
| ALAS2 | GDB:119666 | ANEMIA, HYPOCHROMIC |
| AMCD | GDB:5584286 | ARTHROGRYPOSIS MULTIPLEX CONGENITA, DISTAL |

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| AMELX | GDB:119675 | AMELOGENESIS IMPERFECTA-1, HYPOPLASTIC TYPE; AIH1 |
| ANOP1 | GDB:128454 | CLINICAL; ANOP1 |
| AR | GDB:120556 | ANDROGEN INSENSITIVITY SYNDROME; AIS ANDROGEN RECEPTOR; AR |
| ARAF1 | GDB:119004 | V-RAF MURINE SARCOMA 3611 VIRAL ONCOGENE HOMOLOG 1; ARAF1 |
| ARSC2 | GDB:119702 | ARYLSULFATASE C, f FORM; ARSC2 |
| ARSE | GDB:555743 | CHONDRODYSPLASIA PUNCTATA 1, X-LINKED RECESSIVE; CDPX1 |
| ARTS | GDB:9954651 | FATAL X-LINKED, WITH DEAFNESS AND LOSS OF VISION |
| ASAT | GDB:9954649 | SIDEROBLASTIC, AND SPINOCEREBELLAR ATAxia; ASAT |
| ASSP5 | GDB:119019 | CITRULLINEMIA |
| ATP7A | GDB:119395 | ATPase, Cu(2+)-TRANSPORTING, ALPHA POLYPEPTIDE; ATP7A MENKES SYNDROME |
| ATRX | GDB:136052 | ALPHA-THALASSEMIA/MENTAL RETARDATION SYNDROME, X-LINKED; ATRX ALPHA-THALASSEMIA/MENTAL RETARDATION SYNDROME, NONDELETION TYPE |
| AVPR2 | GDB:131475 | DIABETES INSIPIDUS, NEPHROGENIC |
| BFLS | GDB:120566 | BORJESON SYNDROME; BORJ |

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| BGN | GDB:119727 | BIGLYCAN; BGN |
| BTK | GDB:120542 | BRUTON AGAMMAGLOBULINEMIA TYROSINE KINASE; BTK |
| BZX | GDB:5205912 | BAZEX SYNDROME; BZX |
| C1HR | GDB:119040 | TATA BOX BINDING PROTEIN (TBP)-ASSOCIATED FACTOR 2A; TAF2A |
| CACNA1F | GDB:6053864 | NIGHTBLINDNESS, CONGENITAL STATIONARY, X-LINKED, TYPE 2; CSNB2 CALCIUM CHANNEL, VOLTAGE-DEPENDENT, ALPHA 1F SUBUNIT; CACNA1F |
| CALB3 | GDB:133780 | CALBINDIN 3; CALB3 |
| CBBM | GDB:9958963 | COLORBLINDNESS, BLUE-MONO-CONE-MONOCHROMATIC TYPE; CBBM |
| CCT | GDB:119756 | CATARACT, CONGENITAL TOTAL, WITH POSTERIOR SUTURAL OPACITIES IN HETEROZYGOTES; |
| CDR1 | GDB:119053 | CEREBELLAR DEGENERATION-RELATED AUTOANTIGEN-1; CDR1; CDR34 |
| CFNS | GDB:9579470 | CRANIOFRONTONASAL SYNDROME; CFNS |
| CGF1 | GDB:6275867 | COGNITION |
| CHM | GDB:120400 | CHOROIDEREMIA; CHM |
| CHR39C | GDB:119779 | CHOLESTEROL REPRESSIBLE PROTEIN 39C; CHR39C |

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| CIDX | GDB:127736 | SEVERE COMBINED IMMUNODEFICIENCY DISEASE, X-LINKED, 2; SCIDX2 |
| CLA2 | GDB:119782 | CEREBELLAR ATAXIA, X-LINKED; CLA2 |
| CLCN5 | GDB:270667 | CHLORIDE CHANNEL 5; CLCN5 FANCONI SYNDROME, RENAL, WITH NEPHROCALCINOSIS AND RENAL STONES NEPHROLITHIASIS, X-LINKED RECESSIVE, WITH RENAL FAILURE; XRN |
| CLS | GDB:119784 | RIBOSOMAL PROTEIN S6 KINASE, 90 KD, POLYPEPTIDE 3; RPS6KA3 COFFIN-LOWRY SYNDROME; CLS |
| CMTX2 | GDB:128311 | CHARCOT-MARIE-TOOTH NEUROPATHY, X-LINKED RECESSIVE, 2; CMTX2 |
| CMTX3 | GDB:128151 | CHARCOT-MARIE-TOOTH NEUROPATHY, X-LINKED RECESSIVE, 3; CMTX3 |
| CND | GDB:9954627 | DERMOIDS OF CORNEA; CND |
| COD1 | GDB:119787 | CONE DYSTROPHY, X-LINKED, 1; COD1 |
| COD2 | GDB:6520166 | CONE DYSTROPHY, X-LINKED, 2; COD2 |
| COL4A5 | GDB:120596 | COLLAGEN, TYPE IV, ALPHA-5 CHAIN; COL4A5 LEIOMYOMATOSIS, ESOPHAGEAL AND VULVAL, WITH NEPHROPATHY |
| COL4A6 | GDB:222775 | COLLAGEN, TYPE IV, ALPHA-6 CHAIN; COL4A6 LEIOMYOMATOSIS, ESOPHAGEAL AND VULVAL, WITH NEPHROPATHY |
| CPX | GDB:120598 | CLEFT PALATE, X-LINKED; CPX |

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| CVD1 | GDB:9954659 | CARDIAC VALVULAR DYSPLASIA, X-LINKED |
| CYBB | GDB:120513 | GRANULOMATOUS DISEASE, CHRONIC; CGD |
| DCX | GDB:9823272 | LISSENCEPHALY, X-LINKED |
| DFN2 | GDB:119091 | DEAFNESS, X-LINKED 2, PERCEPTIVE CONGENITAL; DFN2 |
| DFN4 | GDB:433255 | DEAFNESS, X-LINKED 4, CONGENITAL SENSORINEURAL; DFN4 |
| DFN6 | GDB:1320698 | DEAFNESS, X-LINKED, 6, PROGRESSIVE; DFN6 |
| DHOF | GDB:119847 | FOCAL DERMAL HYPOPLASIA; DHOF |
| DIAPH2 | GDB:9835484 | DIAPHANOUS, DROSOPHILA, HOMOLOG OF, 2 DKC1GDB:119096 DYSKERATOSIS CONGENITA; DKC |
| DMD | GDB:119850 | MUSCULAR DYSTROPHY, PSEUDOhypertrophic PROGRESSIVE, DUCHENNE AND BECKER |
| DSS | GDB:433750 | DOSAGE-SENSITIVE SEX REVERSAL; DSS |
| DYT3 | GDB:118789 | TORSION DYSTONIA-3, X-LINKED TYPE; DYT3 |
| EBM | GDB:119102 | BULLOUS DYSTROPHY, HEREDITARY MACULAR TYPE |
| EBP | GDB:125212 | CHONDRODYSPLASIA PUNCTATA, X-LINKED DOMINANT; CDPX2; CDPXD; |

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| | | CPXD |
| ED1 | GDB:119859 | ECTODERMAL DYSPLASIA, ANHIDROTIC; EDA |
| ELK1 | GDB:119867 | ELK1, MEMBER OF ETS ONCOGENE FAMILY; ELK1 |
| EMD | GDB:119108 | MUSCULAR DYSTROPHY, TARDIVE, DREIFUSS-EMERY TYPE, WITH CONTRACTURES |
| EVR2 | GDB:136068 | EXUDATIVE VITREORETINOPATHY, FAMILIAL, X-LINKED RECESSIVE; EVR2 |
| F8C | GDB:119124 | HEMOPHILIA A |
| F9 | GDB:119900 | HEMOPHILIA B; HEMB |
| FCP1 | GDB:347490 | F-CELL PRODUCTION, X-LINKED; FCPX |
| FDPSL5 | GDB:119922 | SYNTHETASE-5; FPSL5 |
| FGD1 | GDB:119131 | SYNDROME FACIOGENITAL DYSPLASIA; FGDY |
| FGS1 | GDB:9836950 | FG SYNDROME |
| FMR1 | GDB:129038 | FRAGILE SITE MENTAL RETARDATION-1; FMR1 |
| FMR2 | GDB:141566 | FRAGILE SITE, FOLIC ACID TYPE, RARE, FRA(X)(q28); FRAXE |
| G6PD | GDB:120621 | GLUCOSE-6-PHOSPHATE |

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| | | DEHYDROGENASE; G6PD |
| GABRA3 | GDB:119968 | GAMMA-AMINOBUTYRIC ACID RECEPTOR, ALPHA-3; GABRA3 |
| GATA1 | GDB:125373 | GATA-BINDING PROTEIN 1; GATA1 |
| GDI1 | GDB:1347097 | GDP DISSOCIATION INHIBITOR 1; GDI1 MENTAL RETARDATION, X-LINKED NONSPECIFIC, TYPE 3; MRX3 |
| GDXY | GDB:9954629 | DYSGENESIS, XY FEMALE TYPE; GDXY |
| GJB1 | GDB:125246 | CHARCOT-MARIE-TOOTH PERONEAL MUSCULAR ATROPHY, X-LINKED; CMTX1 GAP JUNCTION PROTEIN, BETA-1, 32 KD; GJB1 |
| GK | GDB:119271 | HYPERGLYCEROLEMIA |
| GLA | GDB:119272 | ANGIOKERATOMA, DIFFUSE |
| GPC3 | GDB:3770726 | GLYPICAN-3; GPC3 SIMPSON DYSMORPHIA SYNDROME; SDYS |
| GRPR | GDB:128035 | GASTRIN-RELEASING PEPTIDE RECEPTOR; GRPR |
| GTD | GDB:9954635 | GONADOTROPIN DEFICIENCY; GTD |
| GUST | GDB:9954655 | MENTAL RETARDATION WITH OPTIC ATROPHY, DEAFNESS, AND SEIZURES |
| HMS1 | GDB:251827 | I; HMS1 |

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| HPRT1 | GDB:119317 | HYPOXANTHINE GUANINE PHOSPHORIBOSYLTRANSFERASE 1; HPRT1 |
| HPT | GDB:119322 | HYPOPARATHYROIDISM, X-LINKED; HYPX |
| HTC2 | GDB:700980 | HYPERTRICHOSIS, CONGENITAL GENERALIZED; CGH; HCG |
| HTR2C | GDB:378202 | 5-@HYDROXYTRYPTAMINE RECEPTOR 2C; HTR2C |
| HYR | GDB:9954625 | REGULATOR; HYR |
| IDS | GDB:120521 | MUCOPOLYSACCHARIDOSIS TYPE II |
| IHG1 | GDB:119343 | HYPOPLASIA OF, WITH GLAUCOMA; IHG |
| IL2RG | GDB:134807 | INTERLEUKIN-2 RECEPTOR, GAMMA; IL2RG SEVERE COMBINED IMMUNODEFICIENCY DISEASE, X-LINKED, 2; SCIDX2 |
| INDX | GDB:9954657 | IMMUNONEUROLOGIC DISORDER, X-LINKED |
| IP1 | GDB:120105 | INCONTINENTIA PIGMENTI, TYPE I; IP1 |
| IP2 | GDB:120106 | INCONTINENTIA PIGMENTI, TYPE II; IP2 |
| JMS | GDB:204055 | MENTAL RETARDATION, X-LINKED, WITH GROWTH RETARDATION, DEAFNESS, AND |
| KAL1 | GDB:120116 | KALLMANN SYNDROME 1; KAL1 |
| KFSD | GDB:128174 | KERATOSIS FOLLICULARIS SPINULOSA |

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| | | DECALVANS CUM OPHIASI; KFSD |
| L1CAM | GDB:120133 | CLASPED THUMB AND MENTAL RETARDATION L1 CELL ADHESION MOLECULE; L1CAM |
| LAMP2 | GDB:125376 | LYSOSOME-ASSOCIATED MEMBRANE PROTEIN B; LAMP2; LAMPB |
| MAA | GDB:119372 | MICROPHTHALMIA OR ANOPHTHALMOS, WITH ASSOCIATED ANOMALIES; MAA |
| MAFD2 | GDB:119373 | PSYCHOSIS, X-LINKED |
| MAOA | GDB:120164 | MONOAMINE OXIDASE A; MAOA |
| MAOB | GDB:119377 | MONOAMINE OXIDASE B; MAOB |
| MCF2 | GDB:120168 | MCF.2 CELL LINE DERIVED TRANSFORMING SEQUENCE; MCF2 |
| MCS | GDB:128370 | MENTAL RETARDATION, X-LINKED, SYNDROMIC-4, WITH CONGENITAL CONTRACTURES |
| MEAX | GDB:119383 | X-LINKED, WITH EXCESSIVE AUTOPHAGY; XMEA; MEAX |
| MECP2 | GDB:3851454 | SYNDROME; RTT |
| MF4 | GDB:119386 | METACARPAL 4-5 FUSION; MF4 |
| MGC1 | GDB:120179 | MEGALOCORNEA; MGC1; MGCN |
| MIC5 | GDB:120526 | SURFACE ANTIGEN, X-LINKED; SAX |

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| MID1 | GDB:9772232 | OPITZ SYNDROME |
| MLLT7 | GDB:392309 | MYELOID/LYMPHOID OR MIXED-LINEAGE LEUKEMIA, TRANSLOCATED TO, 7; MLLT7 |
| MLS | GDB:262123 | MICROPHTHALMIA WITH LINEAR SKIN DEFECTS; MLS |
| MRSD | GDB:119398 | MENTAL RETARDATION, SKELETAL DYSPLASIA, AND ABDUCENS PALSY; MRSD |
| MRX14 | GDB:138453 | RETARDATION, X-LINKED 14; MRX14 |
| MRX1 | GDB:120193 | MENTAL RETARDATION, X-LINKED NONSPECIFIC, TYPE 1; MRX1 |
| MRX20 | GDB:217050 | MENTAL RETARDATION, X-LINKED 20; MRX20 |
| MRX2 | GDB:120194 | RETARDATION, X-LINKED NONSPECIFIC, TYPE 2; MRX2 |
| MRX3 | GDB:128105 | GDP DISSOCIATION INHIBITOR 1; GDI1 MENTAL RETARDATION, X-LINKED NONSPECIFIC, TYPE 3; MRX3 |
| MRX40 | GDB:700754 | MENTAL RETARDATION, X-LINKED, WITH HYPOTONIA |
| MRXA | GDB:9954641 | MENTAL RETARDATION, X-LINKED NONSPECIFIC, WITH APHASIA; MRXA |
| MSD | GDB:119399 | SYNDROME |
| MTM1 | GDB:119439 | MYOTUBULAR MYOPATHY 1; MTM1 |

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| MYCL2 | GDB:120209 | MYCL-RELATED PROCESSED GENE; MYCL2 |
| MYP1 | GDB:127783 | MYOPIA, X-LINKED; MYP1 |
| NDP | GDB:119449 | NORRIE DISEASE; NDP |
| NHS | GDB:120235 | CATARACT-DENTAL SYNDROME |
| NPHL1 | GDB:433705 | NEPHROLITHIASIS, X-LINKED RECESSIVE, WITH RENAL FAILURE; XRN |
| NR0B1 | GDB:118982 | ADRENAL HYPOPLASIA, CONGENITAL; AHC |
| NSX | GDB:125596 | SYNDROME; NSX |
| NYS1 | GDB:119458 | NYSTAGMUS, X-LINKED; NYS |
| NYX | GDB:119814 | NIGHTBLINDNESS, CONGENITAL STATIONARY, WITH MYOPIA; CSNB1 |
| OA1 | GDB:119459 | ALBINISM, OCULAR, TYPE 1; OA1 |
| OASD | GDB:138457 | OCULAR, WITH LATE-ONSET SENSORINEURAL DEAFNESS; OASD |
| OCRL | GDB:119461 | LOWE OCULOCEREBRORENAL SYNDROME; OCRL |
| ODT1 | GDB:125360 | TEETH, ABSENCE OF |
| OFD1 | GDB:120248 | OROFACIODIGITAL SYNDROME 1; OFD1 |

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| OPA2 | GDB:125358 | OPTIC ATROPHY 2; OPA2 |
| OPD1 | GDB:120249 | OTOPALATODIGITAL SYNDROME |
| OPEM | GDB:119467 | OPHTHALMOPLEGIA, EXTERNAL, AND MYOPIA; OPEM |
| OPN1LW | GDB:120724 | COLORBLINDNESS, PARTIAL, PROTAN SERIES; CBP |
| OPN1MW | GDB:120622 | COLORBLINDNESS, PARTIAL, DEUTAN SERIES; CBD; DCB |
| OTC | GDB:119468 | ORNITHINE TRANSCARBAMYLASE DEFICIENCY, HYPERAMMONEMIA DUE TO; OTC |
| P3 | GDB:9954667 | PROTEIN P3 |
| PDHA1 | GDB:118895 | PYRUVATE DEHYDROGENASE COMPLEX, E1-ALPHA POLYPEPTIDE-1; PDHA1 |
| PDR | GDB:203409 | AMYLOIDOSIS, FAMILIAL CUTANEOUS |
| PFC | GDB:120275 | PROPERDIN DEFICIENCY, X-LINKED |
| PFKFB1 | GDB:125375 | 6-@PHOSPHOFRUCTO-2-KINASE; PFKFB1 |
| PGK1 | GDB:120282 | PHOSPHOGLYCERATE KINASE 1; PGK1 |
| PGK1P1 | GDB:120283 | PHOSPHOGLYCERATE KINASE 1; PGK1 |
| PGS | GDB:128372 | DANDY-WALKER MALFORMATION WITH MENTAL RETARDATION, BASAL GANGLIA DISEASE, |

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| PHEX | GDB:120520 | HYPOPHOSPHATEMIA, VITAMIN D-RESISTANT RICKETS; HYP |
| PHKA1 | GDB:120285 | PHOSPHORYLASE KINASE, ALPHA I SUBUNIT (MUSCLE); PHKA1 |
| PHKA2 | GDB:127279 | GLYCOGEN STORAGE DISEASE VIII |
| PHP | GDB:119494 | PANHYPOPITUITARISM; PHP |
| PIGA | GDB:138138 | PHOSPHATIDYLINOSITOL GLYCAN, CLASS A; PIGA |
| PLP1 | GDB:120302 | PROTEOLIPID PROTEIN, MYELIN; PLP |
| POF1 | GDB:120716 | PREMATURE OVARIAN FAILURE 1; POF1 |
| POLA | GDB:120304 | POLYMERASE, DNA, ALPHA; POLA |
| POU3F4 | GDB:351386 | DEAFNESS, CONDUCTIVE, WITH STAPES FIXATION |
| PPMX | GDB:9954669 | RETARDATION WITH PSYCHOSIS, PYRAMIDAL SIGNS, AND MACROORCHIDISM |
| PRD | GDB:371323 | DYSPLASIA, PRIMARY |
| PRPS1 | GDB:120318 | PHOSPHORIBOSYL PYROPHOSPHATE SYNTHETASE-I; PRPS1 |
| PRPS2 | GDB:120320 | PHOSPHORIBOSYL PYROPHOSPHATE SYNTHETASE-II; PRPS2 |
| PRS | GDB:128368 | MENTAL RETARDATION, X-LINKED, |

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| | | SYNDROMIC-2, WITH DYSMORPHISM AND CEREBRAL |
| PRTS | GDB:128367 | PARTINGTON X-LINKED MENTAL RETARDATION SYNDROME; PRTS |
| PSF2 | GDB:119519 | TRANSPORTER 2, ABC; TAP2 |
| RENBP | GDB:133792 | RENIN-BINDING PROTEIN; RENBP |
| RENS1 | GDB:9806348 | MENTAL RETARDATION, X-LINKED, RENPENNING TYPE |
| RP2 | GDB:120353 | RETINITIS PIGMENTOSA-2; RP2 |
| RP6 | GDB:125381 | PIGMENTOSA-6; RP6 |
| RPGR | GDB:118736 | RETINITIS PIGMENTOSA-3; RP3 |
| RPS4X | GDB:128115 | RIBOSOMAL PROTEIN S4, X-LINKED; RPS4X |
| RPS6KA3 | GDB:365648 | RIBOSOMAL PROTEIN S6 KINASE, 90 KD, POLYPEPTIDE 3; RPS6KA3 |
| RS1 | GDB:119581 | RETINOSCHISIS; RS |
| S11 | GDB:120361 | ANTIGEN, X-LINKED, SECOND; SAX2 |
| SDYS | GDB:119590 | GLYPLICAN-3; GPC3 SIMPSON DYSMORPHIA SYNDROME; SDYS |
| SEDL | GDB:120372 | SPONDYLOEPIPHYSEAL DYSPLASIA, LATE; SEDL |

| | | |
|----------|--------------|--|
| SERPINA7 | GDB:120399 | THYROXINE-BINDING GLOBULIN OF SERUM; TBG |
| SH2D1A | GDB:120701 | IMMUNODEFICIENCY, X-LINKED PROGRESSIVE COMBINED VARIABLE |
| SHFM2 | GDB:226635 | SPLIT-HAND/SPLIT-FOOT ANOMALY, X-LINKED |
| SHOX | GDB:6118451 | SHORT STATURE; SS |
| SLC25A5 | GDB:125190 | ADENINE NUCLEOTIDE TRANSLOCATOR 2; ANT2 |
| SMAX2 | GDB:9954643 | SPINAL MUSCULAR ATROPHY, X-LINKED LETHAL INFANTILE |
| SRPX | GDB:3811398 | RETINITIS PIGMENTOSA-3; RP3 |
| SRS | GDB:136337 | MENTAL RETARDATION, X-LINKED, SNYDER-ROBINSON TYPE |
| STS | GDB:120393 | ICHTHYOSIS, X-LINKED |
| SYN1 | GDB:119606 | SYNAPSIN I; SYN1 |
| SYP | GDB:125295 | SYNAPTOPHYSIN; SYP |
| TAF1 | GDB:120573 | TATA BOX BINDING PROTEIN (TBP)-ASSOCIATED FACTOR 2A; TAF2A |
| TAZ | GDB:120609 | CARDIOMYOPATHY, DILATED 3A; CMD3A ENDOCARDIAL FIBROELASTOSIS-2; EFE2 |
| TBX22 | GDB:10796448 | CLEFT PALATE, X-LINKED; CPX |

| | | |
|--------|------------|---|
| TDD | GDB:119610 | MALE PSEUDOHERMAPHRODITISM: DEFICIENCY OF TESTICULAR 17,20-DESMOLASE; |
| TFE3 | GDB:125870 | TRANSCRIPTION FACTOR FOR IMMUNOGLOBULIN HEAVY-CHAIN ENHANCER-3; TFE3 |
| THAS | GDB:128158 | THORACOABDOMINAL SYNDROME; TAS |
| THC | GDB:125361 | THROMBOCYTOPENIA, X-LINKED; THC; XLT |
| TIMM8A | GDB:119090 | DEAFNESS 1, PROGRESSIVE; DFN1 |
| TIMP1 | GDB:119615 | TISSUE INHIBITOR OF METALLOPROTEINASE-1; TIMP1 |
| TKCR | GDB:119616 | TORTICOLLIS, KELOIDS, CRYPTORCHIDISM, AND RENAL DYSPLASIA; TKC |
| TNFSF5 | GDB:120632 | IMMUNODEFICIENCY WITH INCREASED IgM |
| UBE1 | GDB:118954 | UBIQUITIN-ACTIVATING ENZYME 1; UBE1 |
| UBE2A | GDB:131647 | UBIQUITIN-CONJUGATING ENZYME E2A; UBE2A |
| WAS | GDB:120736 | WISKOTT-ALDRICH SYNDROME; WAS |
| WSN | GDB:125864 | PARKINSONISM, EARLY-ONSET, WITH MENTAL RETARDATION |
| WTS | GDB:128373 | MENTAL RETARDATION, X-LINKED, SYNDROMIC-6, WITH GYNECOMASTIA |

| | | |
|--------|-------------|--|
| | | AND OBESITY; |
| WWS | GDB:120497 | WIEACKER SYNDROME |
| XIC | GDB:120498 | X-INACTIVATION-SPECIFIC TRANSCRIPT; XIST |
| XIST | GDB:126428 | X-INACTIVATION-SPECIFIC TRANSCRIPT; XIST |
| XK | GDB:120499 | Xk LOCUS |
| XM | GDB:119634 | XM SYSTEM |
| XS | GDB:119636 | LUTHERAN SUPPRESSOR, X-LINKED; XS; LUXS |
| ZFX | GDB:120502 | ZINC FINGER PROTEIN, X-LINKED; ZFX |
| ZIC3 | GDB:249141 | HETEROTAXY, X-LINKED VISCERAL; HTX1 |
| ZNF261 | GDB:9785766 | MENTAL RETARDATION, X-LINKED; DXS6673E |
| ZNF41 | GDB:125865 | ZINC FINGER PROTEIN-41; ZNF41 |
| ZNF6 | GDB:120508 | ZINC FINGER PROTEIN-6; ZNF6 |

Table 26: Genes, Locations and Genetic Disorders on Chromosome Y

| Gene | GDB Accession ID | OMIM Link |
|-------|------------------|----------------------------------|
| AMELY | GDB:119676 | AMELOGENIN, Y-CHROMOSOMAL; AMELY |

| | | |
|-------|-------------|---------------------------------------|
| ASSP6 | GDB:119020 | CITRULLINEMIA |
| AZF1 | GDB:119027 | AZOOSPERMIA FACTOR 1; AZF1 |
| AZF2 | GDB:456131 | AZOOSPERMIA FACTOR 2; AZF2 |
| DAZ | GDB:635890 | DELETED IN AZOOSPERMIA; DAZ |
| GCY | GDB:119267 | CONTROL, Y-CHROMOSOME INFLUENCED; GCY |
| RPS4Y | GDB:128052 | RIBOSOMAL PROTEIN S4, Y-LINKED; RPS4Y |
| SMCY | GDB:5875390 | HISTOCOMPATIBILITY Y ANTIGEN; HY; HYA |
| SRY | GDB:125556 | SEX-DETERMINING REGION Y; SRY |
| ZFY | GDB:120503 | ZINC FINGER PROTEIN, Y-LINKED; ZFY |

Table 27: Genes, Locations and Genetic Disorders in Unknown or Multiple Locations

| Gene | GDB Accession ID | OMIM Link |
|------|------------------|---|
| ABAT | GDB:581658 | GAMMA-AMINOBUTYRATE TRANSAMINASE |
| AEZ | GDB:128360 | ACRODERMATITIS ENTEROPATHICA, ZINC-DEFICIENCY TYPE; AEZ |
| AFA | GDB:265277 | FILIFORME ADNATUM AND CLEFT PALATE |
| AFD1 | GDB:265292 | DYSOSTOSIS, TREACHER COLLINS TYPE, WITH LIMB ANOMALIES |

| | | |
|--------|--------------|--|
| AGS1 | GDB:10795417 | ENCEPHALOPATHY, FAMILIAL INFANTILE, WITH CALCIFICATION OF BASAL GANGLIA |
| ASA1 | GDB:6837715 | FARBER LIPOGRANULOMATOSIS |
| ASD1 | GDB:6276019 | atrial septal defect; ASD |
| ASMT | GDB:136259 | CETYL SEROTONIN METHYLTRANSFERASE; ASMT ACETYL SEROTONIN METHYLTRANSFERASE, Y-CHROMOSOMAL; ASMTY; HIOMTY |
| BCH | GDB:118758 | CHOREA, HEREDITARY BENIGN; BCH |
| CCAT | GDB:118738 | CATARACT, CONGENITAL OR JUVENILE |
| CECR9 | GDB:10796163 | CAT EYE SYNDROME; CES |
| CEPA | GDB:581848 | CONTROL, CONGENITAL FAILURE OF |
| CHED2 | GDB:9957389 | CORNEAL DYSTROPHY, CONGENITAL HEREDITARY |
| CLA1 | GDB:119781 | CEREBELLOPARENCHYMAL DISORDER III |
| CLA3 | GDB:128453 | CEREBELLOPARENCHYMAL DISORDER I; CPD I |
| CLN4 | GDB:125229 | CEROID-LIPOFUSCINOSIS, NEURONAL 4; CLN4 |
| CPO | GDB:119070 | COPROPORPHYRIA |
| CSF2RA | GDB:118777 | COLONY STIMULATING FACTOR 2 RECEPTOR, ALPHA; CSF2RA |

| | | |
|---------------|-------------|---|
| | | GRANULOCYTE-MACROPHAGE COLONY-STIMULATING FACTOR RECEPTOR, ALPHA SUBUNIT, |
| CTS1 | GDB:118779 | CARPAL TUNNEL SYNDROME; CTS; CTS1 |
| DF | GDB:132645 | FACTOR D |
| DIH1 | GDB:439243 | DIAPHRAGMATIC |
| DWS | GDB:128371 | SYNDROME; DWS |
| DYT2 | GDB:118788 | DYSTONIA MUSCULORUM DEFORMANS 2; DYT2 |
| DYT4 | GDB:433751 | DYSTONIA MUSCULORUM DEFORMANS 4; DYT4 |
| EBR3 | GDB:118739 | EPIDERMOLYSIS BULLOSA DYSTROPHICA NEUROTROPHICA |
| ECT | GDB:128640 | CENTRALOPATHIC EPILEPSY |
| EEF1A1L1 4 | GDB:1327185 | PROSTATIC CARCINOMA ONCOGENE PTI-1 |
| EYCL2 | GDB:4642815 | EYE COLOR-3; EYCL3 |
| FA1 | GDB:118795 | FANCONI ANEMIA, COMPLEMENTATION GROUP A; FACA |
| FANCB | GDB:9864269 | FANCONI PANCYTOPENIA, TYPE 2 |
| GCSH | GDB:126842 | HYPERGLYCINEMIA, ISOLATED NONKETOTIC, TYPE III; NKH3 |

| | | |
|--------|-------------|---|
| GCSL | GDB:132139 | ISOLATED NONKETOTIC, TYPE IV; NKH4 |
| GDF5 | GDB:433948 | CARTILAGE-DERIVED MORPHOGENETIC PROTEIN 1 |
| GIP | GDB:119985 | GASTRIC INHIBITORY POLYPEPTIDE; GIP |
| GTS | GDB:118807 | GILLES DE LA TOURETTE SYNDROME; GTS |
| HHG | GDB:118740 | HYPERGONADOTROPIC HYPOGONADISM; HHG |
| HMI | GDB:265275 | OF ITO; HMI |
| HOAC | GDB:118812 | DEAFNESS, CONGENITAL, AUTOSOMAL RECESSIVE |
| HOKPP2 | GDB:595535 | HYPOKALEMIC PERIODIC PARALYSIS, TYPE II; HOKPP2 |
| HRPT1 | GDB:125252 | HYPERPARATHYROIDISM, FAMILIAL PRIMARY |
| HSD3B3 | GDB:676973 | GIANT CELL HEPATITIS, NEONATAL |
| HTC1 | GDB:265286 | HYPERTRICHOSIS UNIVERSALIS CONGENITA, AMBRAS TYPE; HTC1 |
| HV1S | GDB:9955009 | HERPES VIRUS SENSITIVITY; HV1S |
| ICR1 | GDB:127785 | LAMELLAR, AUTOSOMAL DOMINANT FORM |
| ICR5 | GDB:127789 | ICHTHYOSIS CONGENITA, HARLEQUIN FETUS TYPE |

| | | |
|-------|-------------|---|
| IL3RA | GDB:128985 | INTERLEUKIN-3 RECEPTOR, ALPHA; IL3RA INTERLEUKIN-3 RECEPTOR, Y-CHROMOSOMAL; IL3RA |
| KAL2 | GDB:265288 | KALLMANN SYNDROME 2; KAL2 |
| KMS | GDB:118827 | SYNDROME; KMS |
| KRT18 | GDB:120127 | KERATIN 18; KRT18 |
| KSS | GDB:9957718 | KEARNS-SAYRE SYNDROME; KSS |
| LCAT | GDB:119359 | FISH-EYE DISEASE; FED LECITHIN:CHOLESTEROL ACYLTRANSFERASE DEFICIENCY |
| LIMM | GDB:9958161 | MYOPATHY, MITOCHONDRIAL, LETHAL INFANTILE; LIMM |
| MANBB | GDB:125262 | MANNOSIDOSIS, BETA; MANB1 |
| MCPH2 | GDB:9863035 | MICROCEPHALY; MCT |
| MEB | GDB:599557 | DISEASE |
| MELAS | GDB:9955855 | MELAS SYNDROME |
| MIC2 | GDB:120184 | SURFACE ANTIGEN MIC2; MIC2; CD99 MIC2 SURFACE ANTIGEN, Y-CHROMOSOMAL; MIC2Y |
| MPFD | GDB:439372 | CONGENITAL, WITH FIBER-TYPE DISPROPORTION |
| MS | GDB:229116 | SCLEROSIS; MS |

| | | |
|--------|------------|--|
| MSS | GDB:118743 | MARINESCO-SJOGREN SYNDROME; MSS |
| MTATP6 | GDB:118897 | ATP SYNTHASE 6; MTATP6 |
| MTCO1 | GDB:118900 | COMPLEX IV, CYTOCHROME c OXIDASE SUBUNIT I; MTCO1; COI |
| MTCO3 | GDB:118902 | CYTOCHROME c OXIDASE III; MTCO3 |
| MTCYB | GDB:118906 | COMPLEX III, CYTOCHROME b SUBUNIT |
| MTND1 | GDB:118911 | COMPLEX I, SUBUNIT ND1; MTND1 |
| MTND2 | GDB:118912 | COMPLEX I, SUBUNIT ND2; MTND2 |
| MTND4 | GDB:118914 | COMPLEX I, SUBUNIT ND4; MTND4 |
| MTND5 | GDB:118916 | COMPLEX I, SUBUNIT ND5; MTND5 |
| MTND6 | GDB:118917 | COMPLEX I, SUBUNIT ND6; MTND6 |
| MTRNR1 | GDB:118920 | RIBOSOMAL RNA, MITOCHONDRIAL, 12S; MTRNR1 |
| MTRNR2 | GDB:118921 | RIBOSOMAL RNA, MITOCHONDRIAL, 16S; MTRNR2 |
| MTTE | GDB:118926 | TRANSFER RNA, MITOCHONDRIAL, GLUTAMIC ACID; MTTE |
| MTTG | GDB:118933 | TRANSFER RNA, MITOCHONDRIAL, GLYCINE; MTTG |
| MTTI | GDB:118935 | TRANSFER RNA, MITOCHONDRIAL, |

| | | |
|-------|-------------|--|
| | | ISOLEUCINE; MTTI |
| MTTK | GDB:118936 | MERRF SYNDROME TRANSFER RNA, MITOCHONDRIAL, LYSINE; MTTK |
| MTTL1 | GDB:118937 | MERRF SYNDROME TRANSFER RNA, MITOCHONDRIAL, LEUCINE, 1; MTTL1 |
| MTTL2 | GDB:118938 | TRANSFER RNA, MITOCHONDRIAL, LEUCINE, 2; MTTL2 |
| MTTN | GDB:118940 | TRANSFER RNA, MITOCHONDRIAL, ASPARAGINE; MTTN |
| MTTP | GDB:118941 | TRANSFER RNA, MITOCHONDRIAL, PROLINE; MTTP |
| MTTS1 | GDB:118944 | TRANSFER RNA, MITOCHONDRIAL, SERINE, 1; MTTS1 |
| NAMSD | GDB:681237 | NEUROPATHY, MOTOR-SENSORY, TYPE II, WITH DEAFNESS AND MENTAL RETARDATION |
| NODAL | GDB:9848762 | NODAL, MOUSE, HOMOLOG OF |
| OCD1 | GDB:118846 | DISORDER-1; OCD1 |
| OPD2 | GDB:131394 | SYNDROME |
| PCK2 | GDB:137198 | PHOSPHOENOLPYRUVATE CARBOXYKINASE 2, MITOCHONDRIAL; PCK2 |
| PCLD | GDB:433949 | POLYCYSTIC LIVER DISEASE; PLD |

| | | |
|---------|-------------|---|
| PCOS1 | GDB:1391802 | STEIN-LEVENTHAL SYNDROME |
| PFKM | GDB:120277 | GLYCOGEN STORAGE DISEASE VII |
| PKD3 | GDB:127866 | KIDNEY DISEASE 3, AUTOSOMAL DOMINANT; PKD3 |
| PRCA1 | GDB:342066 | PROSTATE CANCER; PRCA1 |
| PRO1 | GDB:128585 | |
| PROP1 | GDB:9834318 | PROPHET OF PIT1, MOUSE, HOMOLOG OF; PROP1 |
| RBS | GDB:118862 | ROBERTS SYNDROME; RBS |
| RFXAP | GDB:9475355 | REGULATORY FACTOR X-ASSOCIATED PROTEIN; RFXAP |
| RP | GDB:9958158 | RETINITIS PIGMENTOSA-8 |
| SLC25A6 | GDB:125184 | ADENINE NUCLEOTIDE TRANSLOCATOR 3; ANT3 ADENINE NUCLEOTIDE TRANSLOCATOR 3, Y-CHROMOSOMAL; ANT3Y |
| SPG5B | GDB:250333 | SPASTIC PARAPLEGIA-5B, AUTOSOMAL RECESSIVE; SPG5B |
| STO | GDB:439375 | CEREBRAL GIGANTISM |
| SUOX | GDB:5584405 | SULFOCYSTEINURIA |
| TC21 | GDB:5573831 | ONCOGENE TC21 |

| | | |
|-----|------------|--------------------------|
| THM | GDB:439378 | FAMILIAL |
| TST | GDB:134043 | RHODANESE; RDS |
| TTD | GDB:230276 | TRICHOThIODYSTROPHY; TTD |

Equivalents:

The present invention is not to be limited in scope by the specific embodiments described herein. Indeed, various modifications of the invention in addition to those described will become apparent to those skilled in the art from the foregoing 5 description and accompanying figures. Such modifications are intended to fall within the scope of the appended claims.

Various publications are cited herein, the disclosures of which are incorporated by reference in their entireties.

What is claimed is:

1. A method of identifying a compound that modulates premature translation termination or nonsense-mediated mRNA decay, said method comprising:
 - (a) contacting a member of a library of compounds with a cell containing a first nucleic acid sequence and a second nucleic acid sequence, wherein the first nucleic acid sequence comprises a regulatory element operably linked to a reporter gene and the second nucleic acid sequence comprises a nucleotide sequence with a premature stop codon that encodes a regulatory protein that binds to the regulatory element of the first nucleic acid sequence and regulates the expression of the reporter gene; and
 - (b) detecting the expression of the reporter gene, wherein a compound that modulates premature translation termination or nonsense-mediated mRNA decay is identified if the expression of the reporter gene in the presence of the compound is altered relative to the expression of the reporter gene in the absence of the compound or the presence of a negative control.
2. A method of identifying a compound that modulates premature translation termination or nonsense-mediated mRNA decay, said method comprising:
 - (a) contacting a member of a library of compounds with a cell containing a first nucleic acid sequence, a second nucleic acid sequence and a third nucleic acid sequence, wherein (i) the first nucleic acid sequence comprises a nucleotide sequence encoding a first fusion protein comprising a DNA binding domain and a first protein, the nucleotide sequence of the first protein containing a premature stop codon, (ii) the second nucleic acid sequence comprises a nucleotide sequence encoding a second fusion protein comprising an activation domain and a second protein, the second protein interacting with the first protein to produce a regulatory protein, and (iii) the third nucleic acid sequence comprises a regulatory element operably linked to a reporter gene, the expression of the reporter gene being regulated by the binding of the regulatory protein to the regulatory element; and
 - (b) detecting the expression of the reporter gene, wherein a compound that modulates premature translation termination or nonsense-mediated mRNA decay is identified if the expression of the reporter gene in the presence of

the compound is altered relative to the expression of the reporter gene in the absence of the compound or the presence of a negative control.

3. A method of identifying a compound that modulates premature translation termination or nonsense-mediated mRNA decay, said method comprising:

- 5 (a) contacting a member of a library of compounds with a cell containing a first nucleic acid sequence, a second nucleic acid sequence and a third nucleic acid sequence, wherein (i) the first nucleic acid sequence comprises a nucleotide sequence encoding a first fusion protein comprising a DNA binding domain and a first protein, (ii) the second nucleic acid sequence comprises a nucleotide sequence encoding a second fusion protein comprising an activation domain and a second protein, the nucleotide sequence of the second protein containing a premature stop codon and the second protein interacting with the first protein to produce a regulatory protein, and (iii) the third nucleic acid sequence comprises a regulatory element operably linked to a reporter gene, the expression of the reporter gene being regulated by the binding of the regulatory protein to the regulatory element; and
- 10 (b) detecting the expression of the reporter gene, wherein a compound that modulates premature translation termination or nonsense-mediated mRNA decay is identified if the expression of the reporter gene in the presence of the compound is altered relative to the expression of the reporter gene in the absence of the compound or the presence of a negative control.
- 15

4. A method for identifying a compound that modulates premature translation termination or nonsense-mediated mRNA decay, said method comprising:

- 20 (a) contacting a member of a library of compounds with a cell-free translation mixture and a nucleic acid sequence comprising a regulatory element operably linked to a reporter gene, wherein the reporter gene contains a premature stop codon and the cell-free translation mixture is isolated from cells that have been incubated at about 0°C to about 10°C; and
- 25 (b) detecting the expression of the reporter gene, wherein a compound that modulates premature translation termination or nonsense-mediated mRNA decay is identified if the expression of the reporter gene in the presence of

the compound is altered relative to the expression of the reporter gene in the absence of the compound or the presence of a negative control.

5. A method for identifying a compound that modulates premature translation termination or nonsense-mediated mRNA decay, said method comprising:

- 5 (a) contacting a member of a library of compounds with a cell-free translation mixture and a nucleic acid sequence comprising a regulatory element operably linked to a reporter gene, wherein the reporter gene contains a premature stop codon and the cell-free translation mixture is a S10 to S30 cell-free extract; and
- 10 (b) detecting the expression of the reporter gene, wherein a compound that modulates premature translation termination or nonsense-mediated mRNA decay is identified if the expression of the reporter gene in the presence of the compound is altered relative to the expression of the reporter gene in the absence of the compound or the presence of a negative control.

15 6. The method of claim 4, wherein the cell-free translation mixture is a S10 to S30 cell-free extract.

7. The method of claim 5, wherein the cell-free translation mixture is a S12 cell-free extract.

8. The method of claim 6, wherein the cell-free translation mixture is a S12 cell-free extract.

9. A method of identifying a compound to be tested for its ability to prevent or treat a disease characterized by or associated with the presence of a premature stop codon in a gene, said method comprising:

- 25 (a) contacting a member of a library of compounds with a cell containing a nucleic acid sequence comprising a reporter gene with a premature stop codon; and
- (b) detecting the expression of the reporter gene,

so that if the expression of the reporter gene in the presence of the compound is altered relative to the expression of the reporter gene in the absence of the compound or the

presence of a negative control, then a compound to be tested for its ability to prevent or treat the disease is identified, wherein the disease is familial hypercholesterolemia, osteogenesis imperfecta, cirrhosis, ataxia telangiectasia or a lysosomal storage disease.

10. A method of identifying a compound to be tested for its ability to prevent or
5 treat a disease characterized by or associated with the presence of a premature stop codon in
a gene, said method comprising:

- (a) contacting a member of a library of compounds with a cell-free translation mixture and a nucleic acid sequence comprising a reporter gene with a premature stop codon; and
- 10 (b) detecting the expression of the reporter gene,

so that if the expression of the reporter gene in the presence of the compound is altered relative to the expression of the reporter gene in the absence of the compound or the presence of a negative control, then a compound to be tested for its ability to prevent or treat the disease is identified, wherein the disease is familial hypercholesterolemia, osteogenesis imperfecta, cirrhosis, ataxia telangiectasia or a lysosomal storage disease.
15

11. The method of claim 1, 2, 3, 4 or 5, wherein the method further comprises determining the structure of the compound that suppresses premature translation termination or nonsense-mediated mRNA decay.

12. The method of claim 9 or 10, wherein the method further comprises
20 determining the structure of the compound.

13. The method of claim 1, 2, 3, 4, 5, 9 or 10, wherein the reporter gene is firefly luciferase, renilla luciferase, click beetle luciferase, green fluorescent protein, yellow fluorescent protein, red fluorescent protein, cyan fluorescent protein, blue fluorescent protein, beta galactosidase, beta glucuronidase, beta lactamase, chloramphenicol acetyltransferase, or alkaline phosphatase.
25

14. The method of claim 1, 2, 3 or 9, wherein the cell is selected from the group consisting of 293T, HeLa, MCF7, Wi-38, SkBr3, Jurkat, CEM, THP1, 3T3, and Raw264.7 cells.

15. The method of claim 4, 5 or 10, wherein the cell-free translation mixture is a cell-free extract from 293T, HeLa, MCF7, Wi-38, SkBr3, Jurkat, CEM, THP1, 3T3, or Raw264.7 cells.

16. The method of claim 1, 2, 3, 4, 5, 9 or 10, wherein the compound is selected
5 from a combinatorial library of compounds comprising peptoids; random biooligomers; diversomers such as hydantoins, benzodiazepines and dipeptides; vinylogous polypeptides; nonpeptidal peptidomimetics; oligocarbamates; peptidyl phosphonates; peptide nucleic acid libraries; antibody libraries; carbohydrate libraries; and small organic molecule libraries.

17. The method of claim 16, wherein the small organic molecule libraries are
10 libraries of benzodiazepines, isoprenoids, thiazolidinones, metathiazanones, pyrrolidines, morpholino compounds, or diazepindiones.

18. The method of claim 1, 2, 3, 4, 5, 9 or 10, wherein the premature stop codon is UAG or UGA.

19. The method of claim 1, 2, 3, 4, 5, 9 or 10, wherein the premature stop codon
15 context is UAGA, UAGC, UAGG, UAGU, UGAA, UGAC, UGAG or UGAU.

WILD-TYPE RNA IN HeLa TRANSLATION EXTRACT

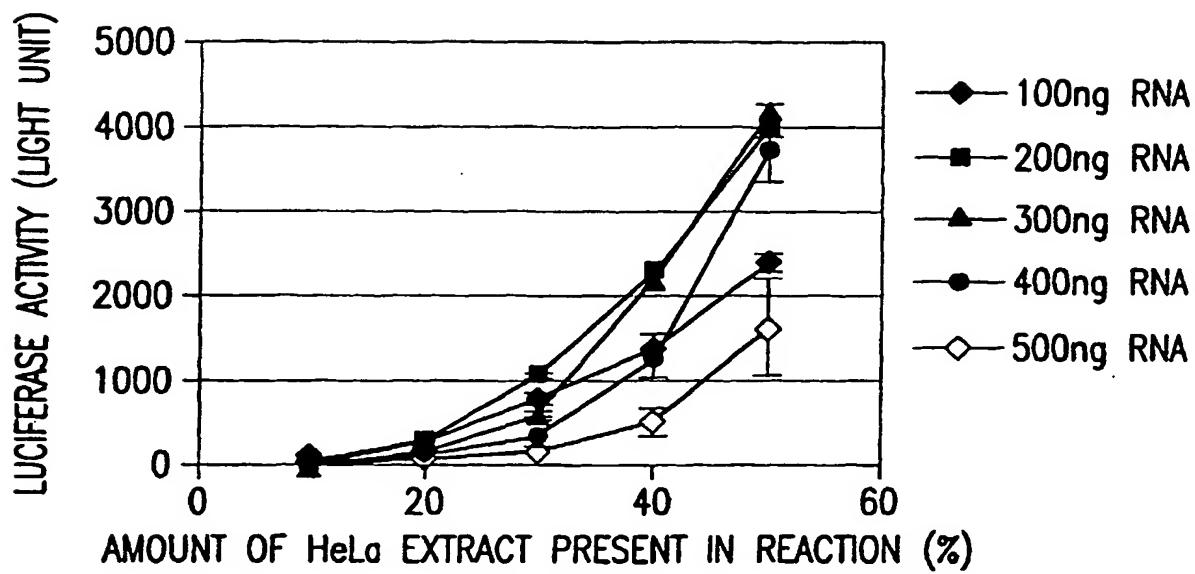


FIG. 1

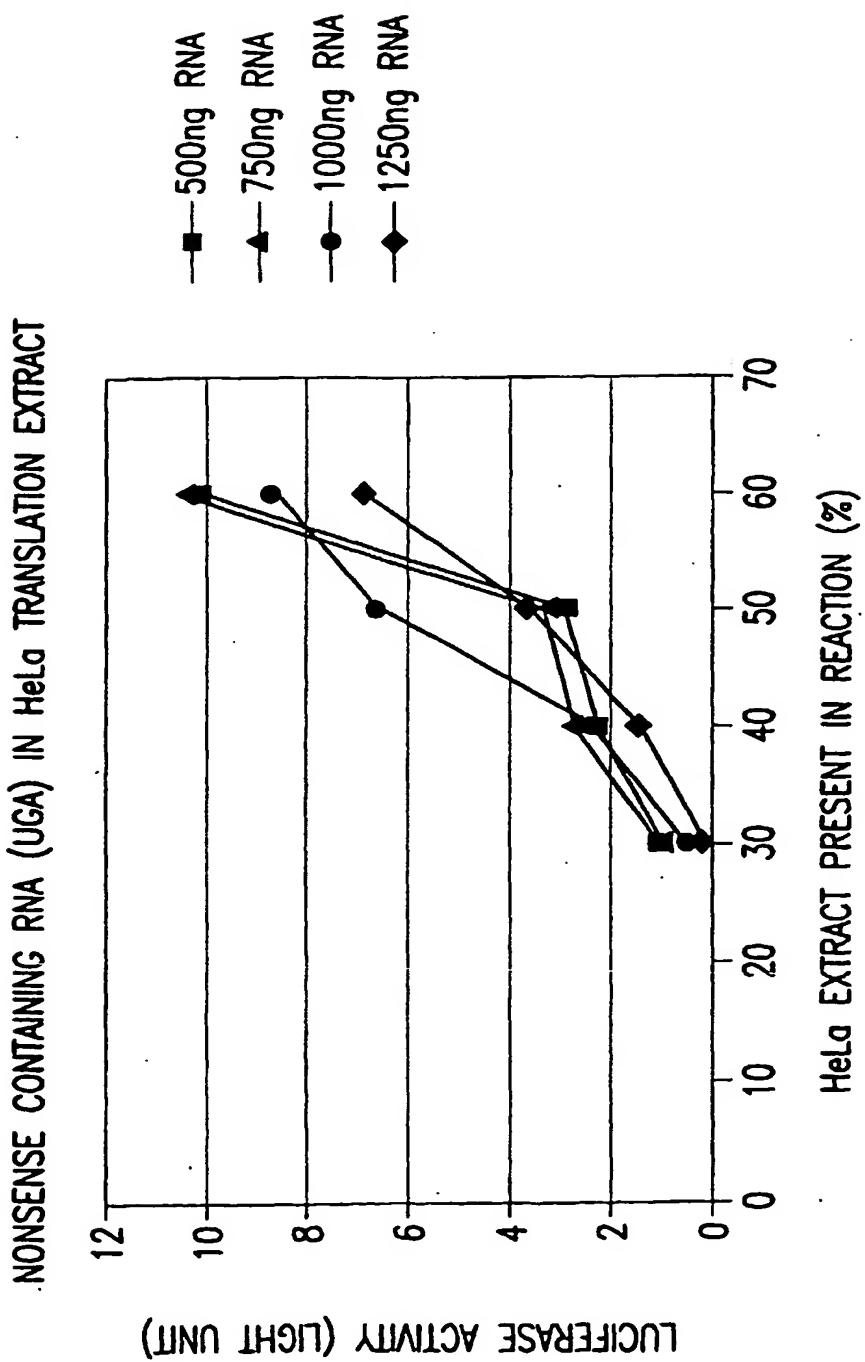


FIG. 2

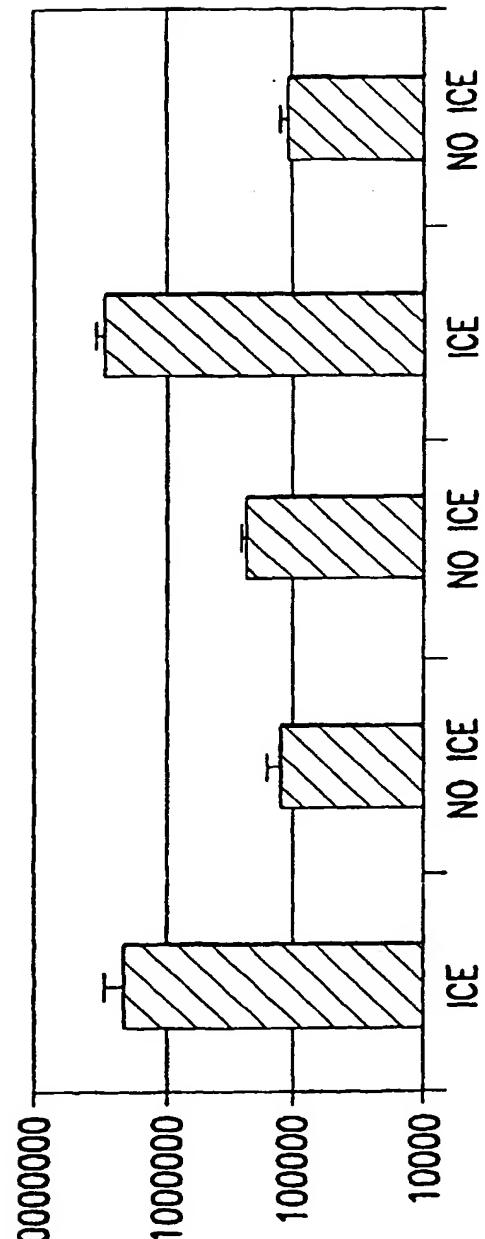


FIG. 3

LUCIFERASE ACTIVITY (LOG)

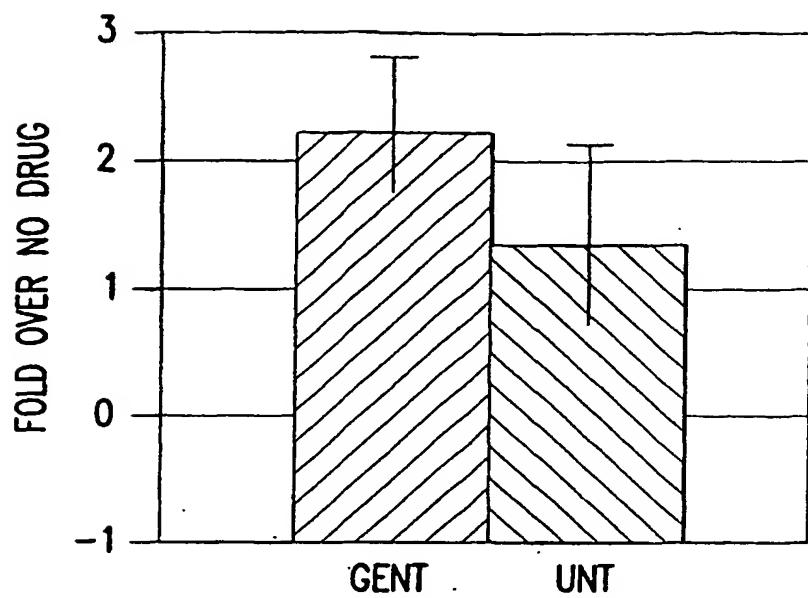


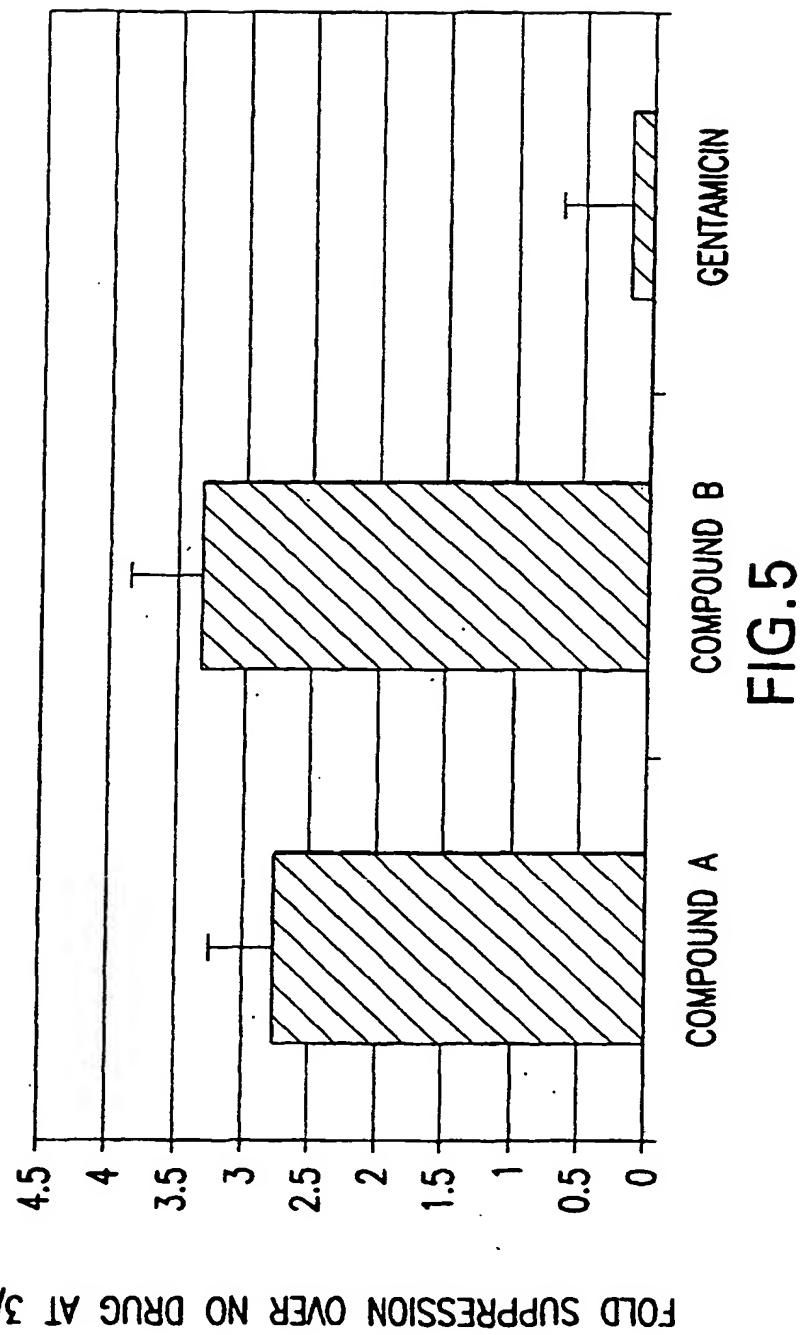
FIG.4

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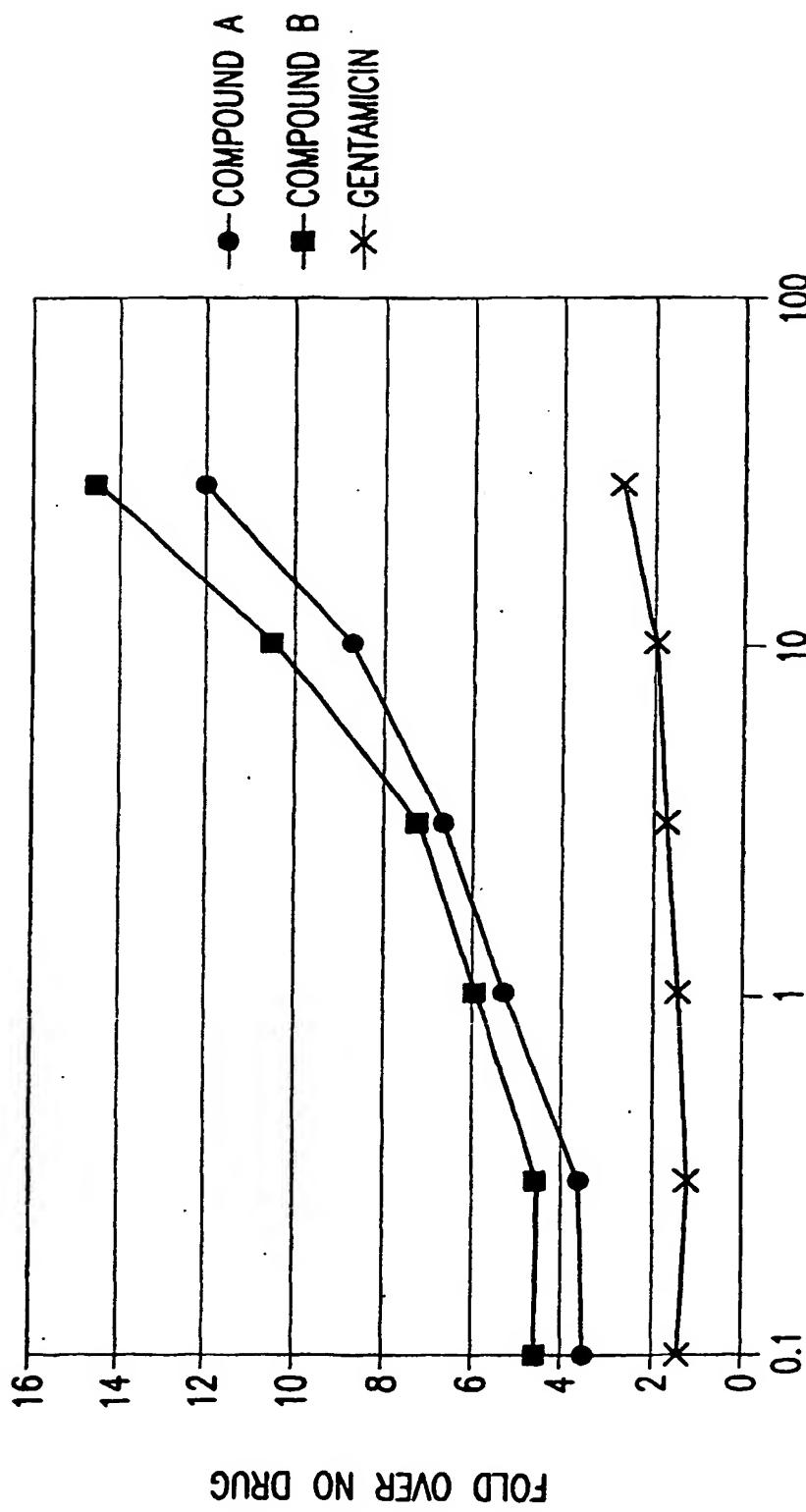
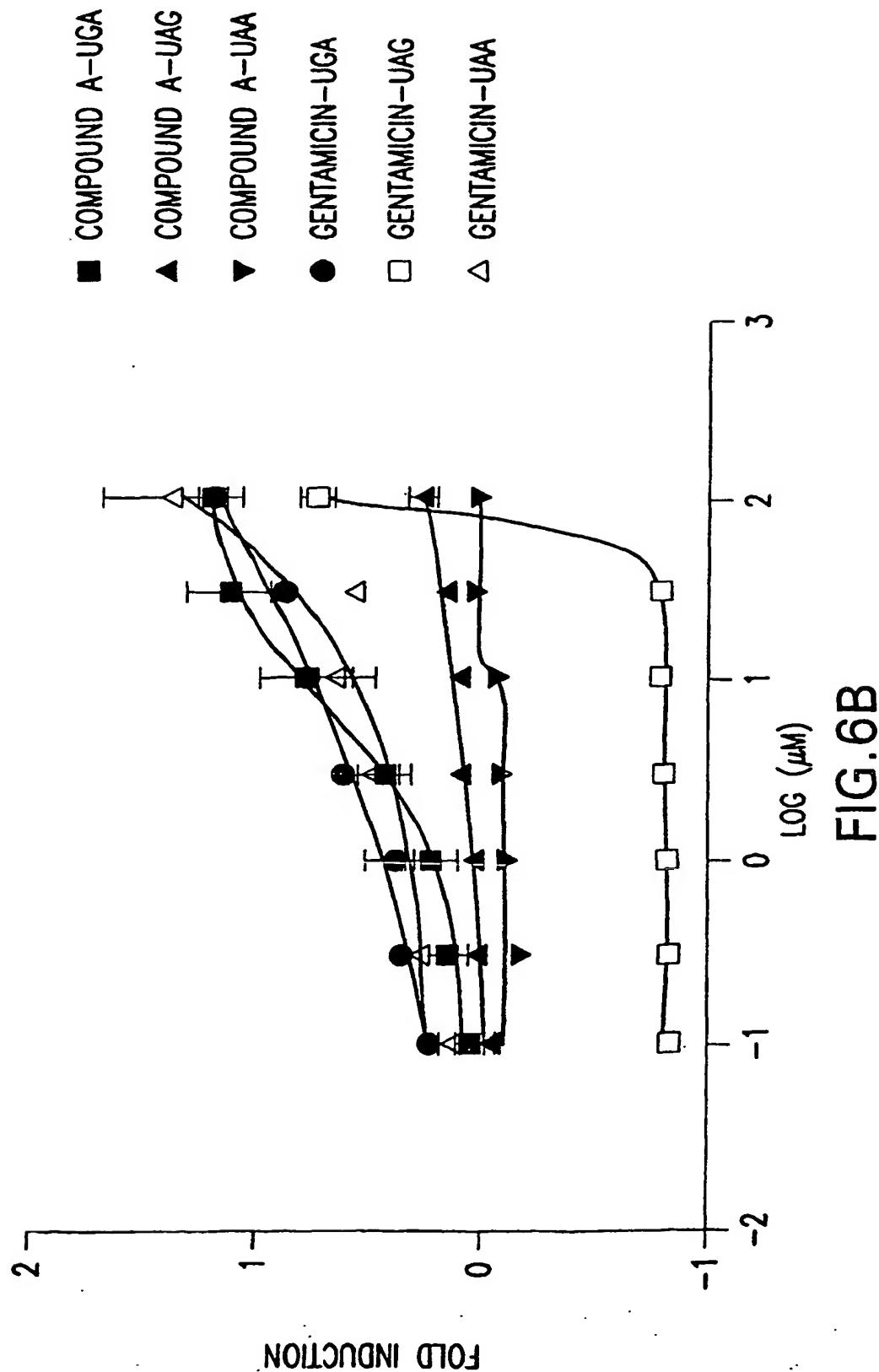


FIG. 6A



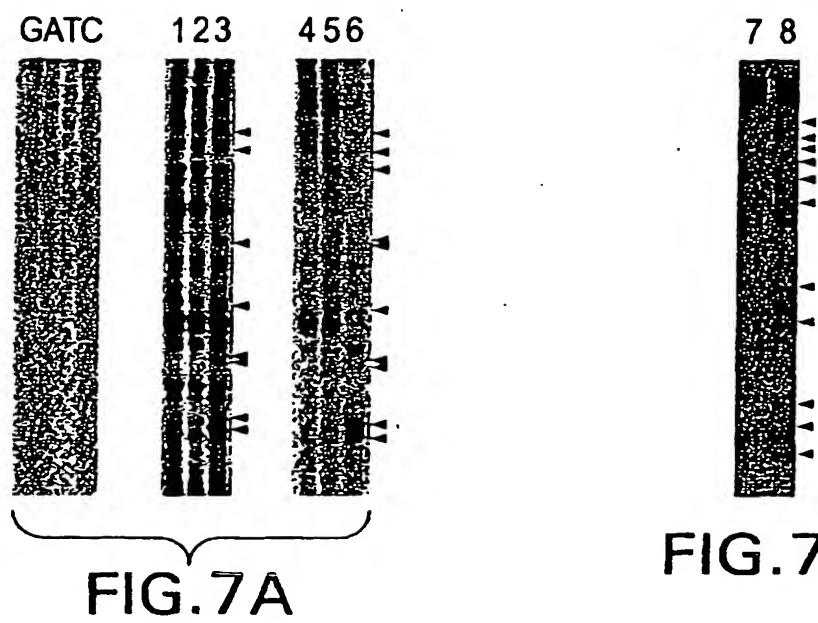


FIG.7B

FIG.7A

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3 TO 4-FOLD INCREASE IN Cl-
CHANNEL ACTIVITY (SPQ) AT 20 μ M

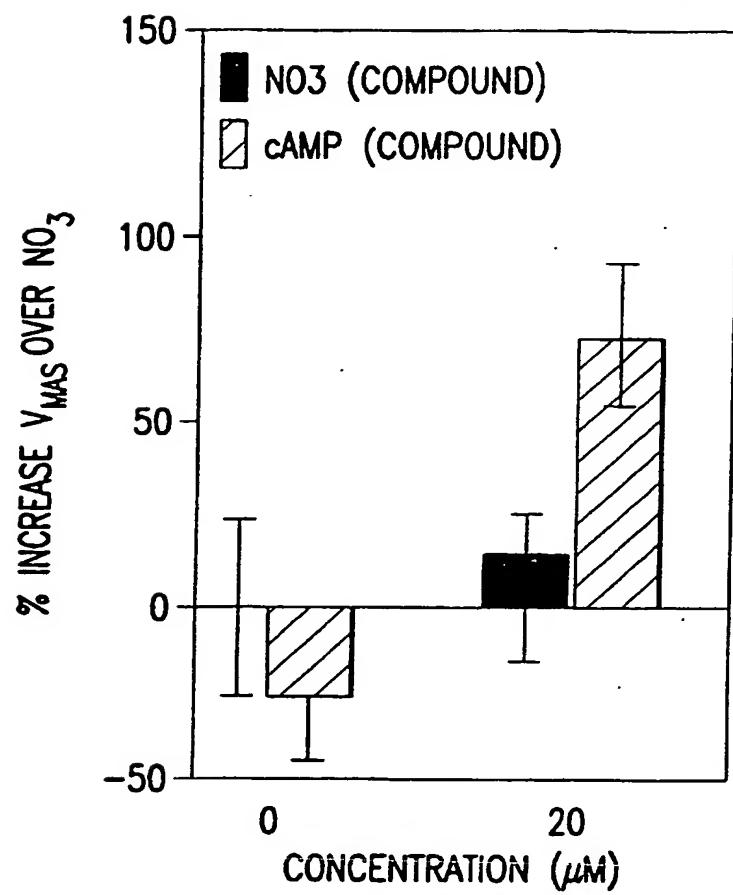
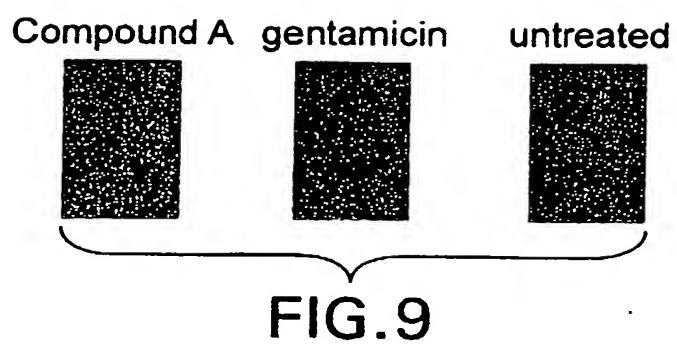


FIG.8



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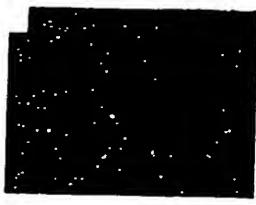


FIG. 10A

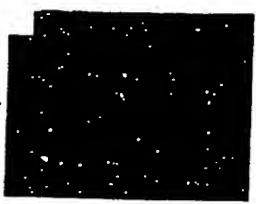


FIG. 10B

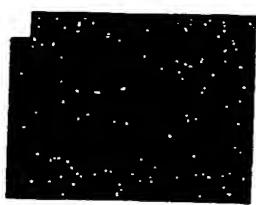


FIG. 10C

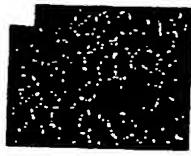


FIG. 10D



FIG. 10E



FIG. 10F